

Catalytic *in vivo* protein knockdown

Technical Journal Club

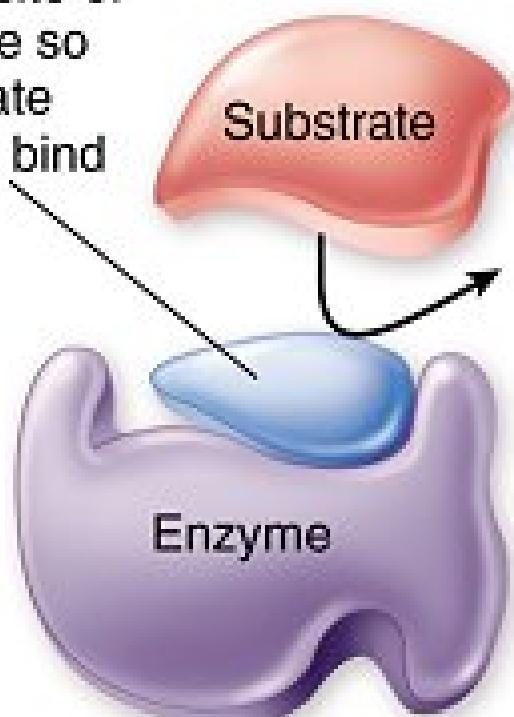
Mario Nuvolone

November 15th 2016

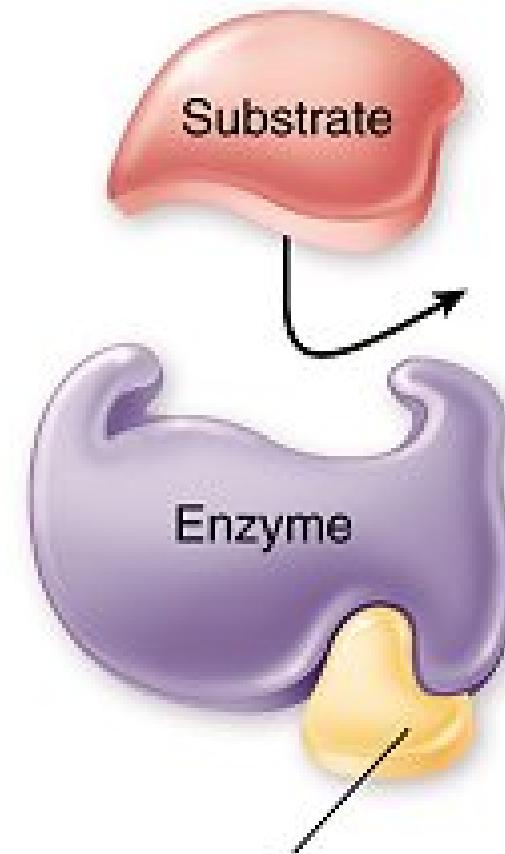


Current inhibitory-based pharmacologic approach

Competitive inhibitor interferes with active site of enzyme so substrate cannot bind



(a) Competitive inhibition



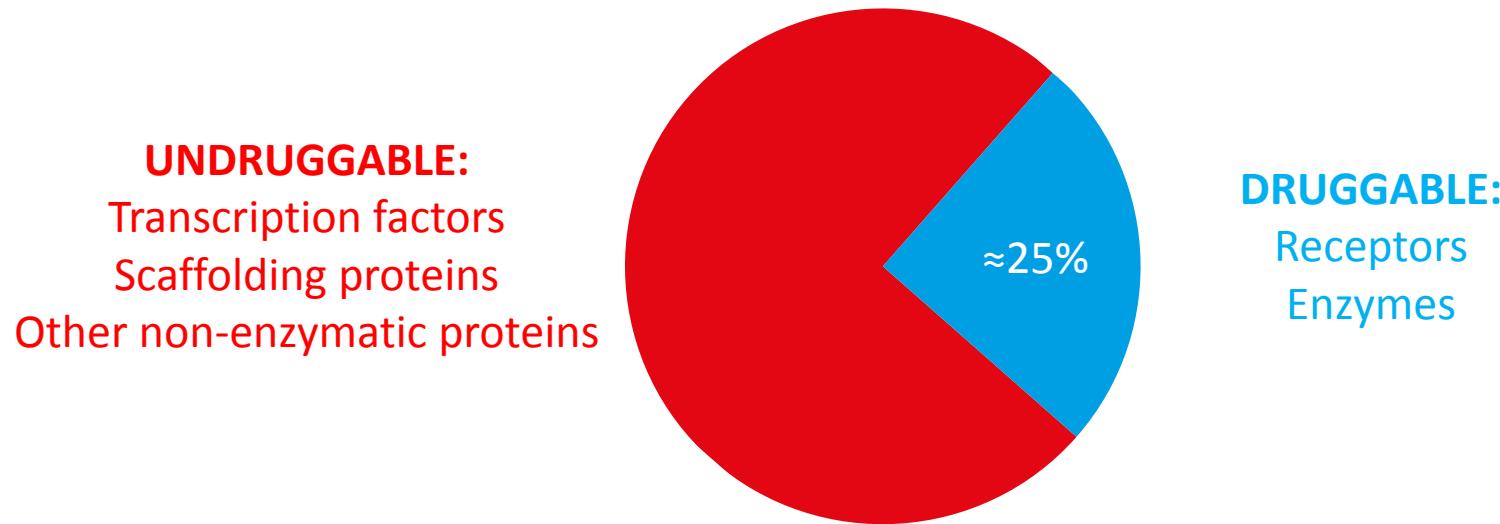
Noncompetitive inhibitor changes shape of enzyme so it cannot bind to substrate

(b) Noncompetitive inhibition

Limitations of inhibitory-based pharmacologic approach

1) Limited to proteins with tractable ligand binding site

→ Limited drug space ($\approx 25\%$ of human proteome)



2) Need to maintain high systemic exposure for high *in vivo* inhibition

3) Potential off-target binding and unwanted side effects

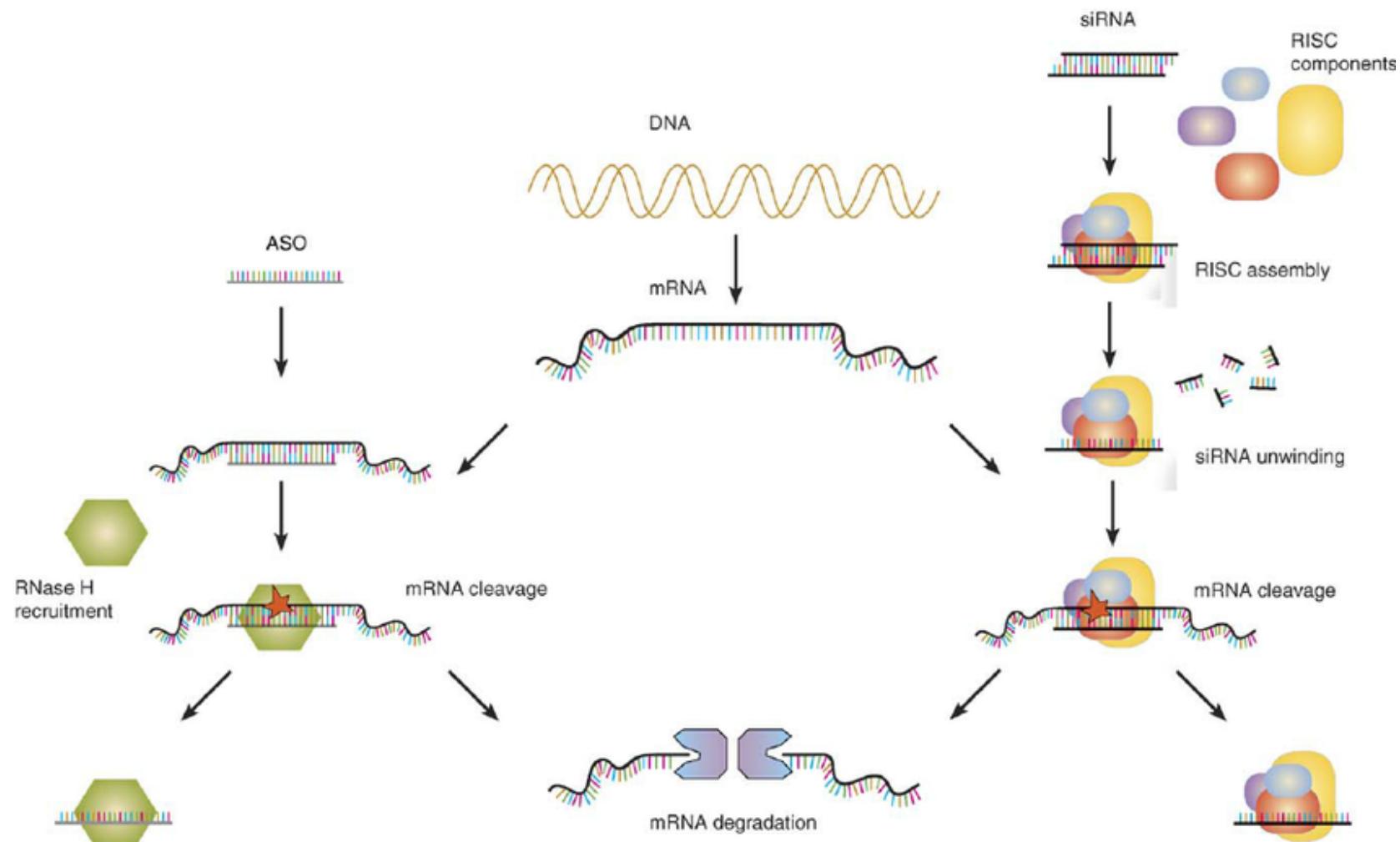
Alternatives approaches

1) Gene knockdown

2) Protein knockdown

Alternatives approaches I: Gene knockdown

- 1) Based on anti-sense oligonucleotides (ASOs) or RNA interference (RNAi)
- 2) Aims at decreasing translation of transcripts from disease-causing genes



Wacheck & Zangemeister-Wittke. Crit Rev Hematol Oncol 2006

Limitations of Gene knockdown

1) Pharmacokinetics profile:

- Limited stability of double-stranded RNA for RNAi
- ASOs preferentially accumulate in the liver

2) Off-target issues

3) Requires time to achieve efficient knockdown (especially for target proteins with long half-life)

→ Limited therapeutic applications

Toure & Crews. Angew Chem Int Ed 2016
Ohoka et al. Curr Cancer Drug Targets 2016

Protein knockdown

1) PROTACs

2) Phtalimide conjugation

Alternatives approaches II: PROTAC-based Protein knockdown

PROTAC: PROteolysis TArgeting Chimeras

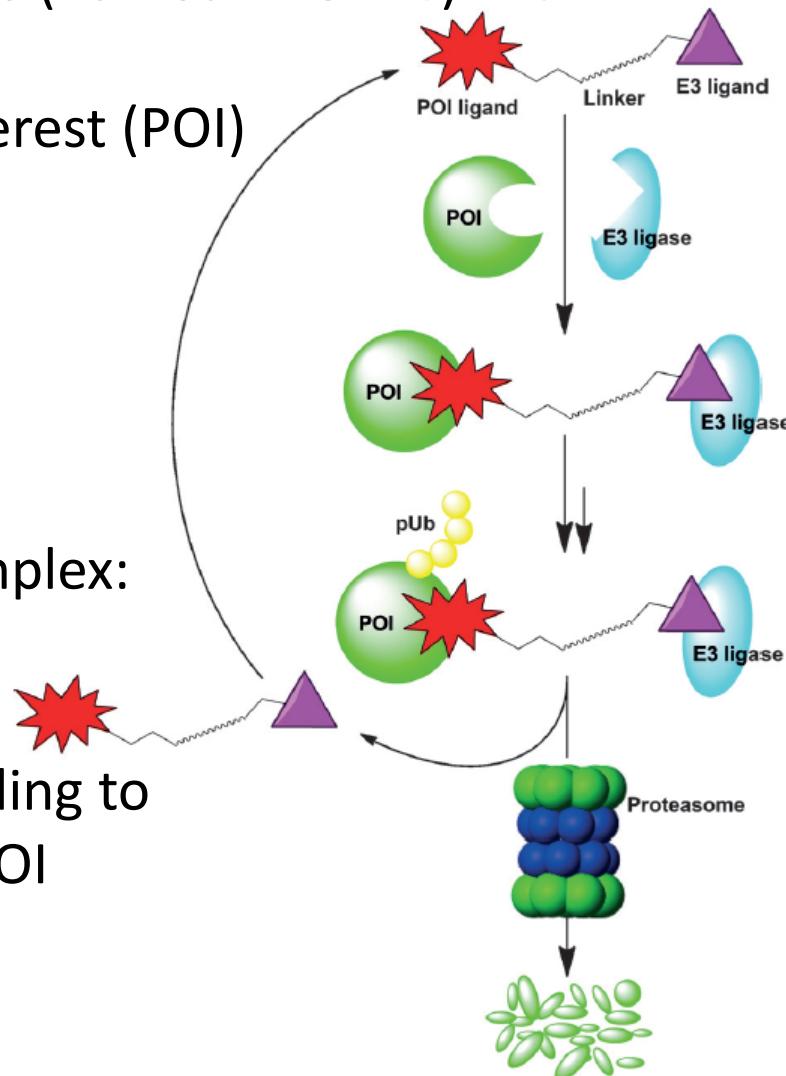
1) Based on heterobifunctional compounds (named PROTAC) with:

- Ligand specific to the Protein Of Interest (POI)
- Linker
- Ligand specific to an E3 ligase

2) Favours the formation of a ternary complex:

POI – PROTAC – E3

3) E3 ligase poly-ubiquitinates the POI, leading to proteasome-mediated degradation of POI



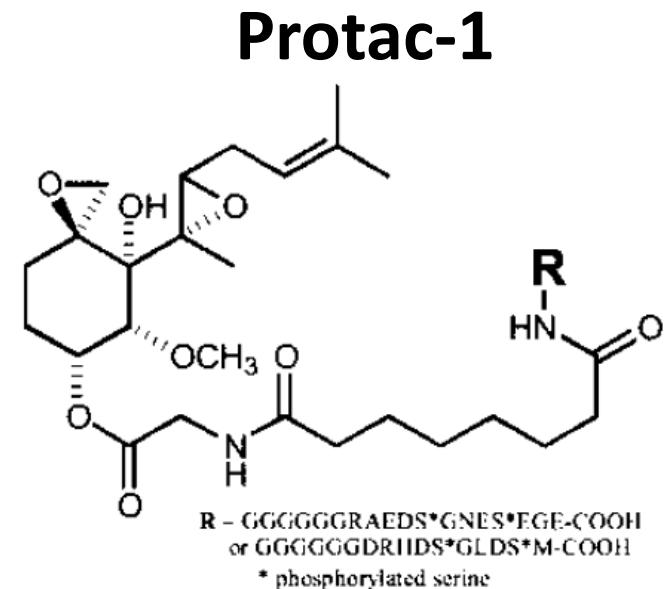
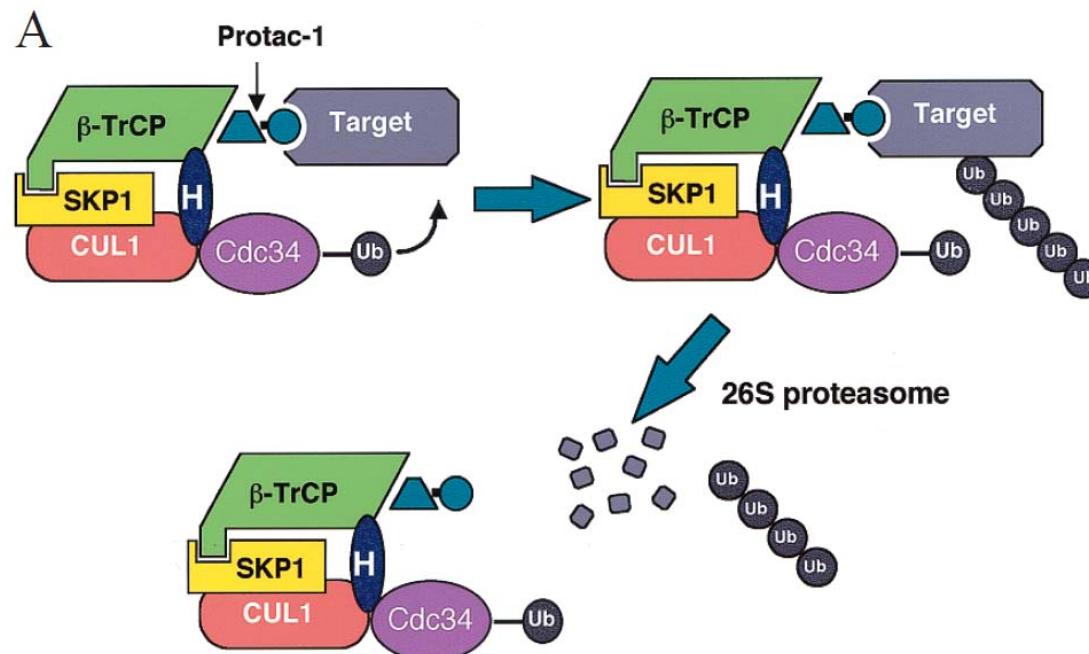
Toure & Crews. Angew Chem Int Ed 2016

Ohoka et al. Curr Cancer Drug Targets 2016

Protacs: Chimeric molecules that target proteins to the Skp1–Cullin–F box complex for ubiquitination and degradation

Kathleen M. Sakamoto^{*†‡}, Kyung B. Kim[§], Akiko Kumagai[†], Frank Mercurio[¶], Craig M. Crews[§], and Raymond J. Deshaies^{†‡||}

^{*}Department of Pediatrics and Pathology, Mattel Children's Hospital at University of California Los Angeles, University of California Los Angeles School of Medicine, Gwynn Hazen Cherry Memorial Laboratories, and Jonsson Comprehensive Cancer Center, Los Angeles, CA 90095-1752; [†]Division of Biology, and ^{||}Howard Hughes Medical Institute, California Institute of Technology, Pasadena, CA 91125; [§]Department of Molecular, Cellular, and Developmental Biology, Yale University, New Haven, CT 06520; and [¶]Signal Division, Celgene Pharmaceuticals, La Jolla, CA 92121



Protacs: Chimeric molecules that target proteins to the Skp1–Cullin–F box complex for ubiquitination and degradation

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Main result:

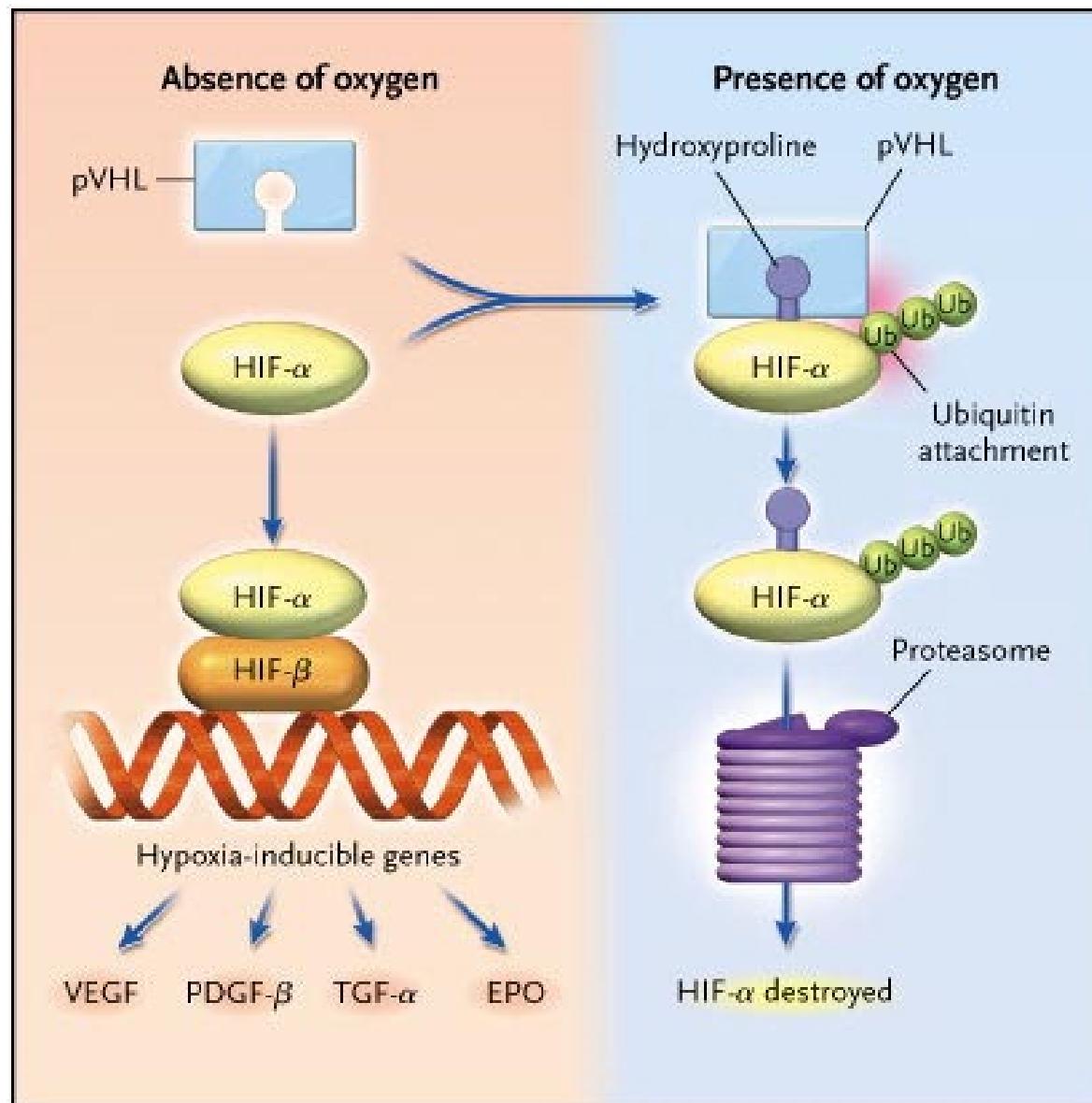
First proof-of-concept for the PROTAC technology

Caveat:

No cell permeability due to peptidic E3-recruiting moiety of PROTAC-1

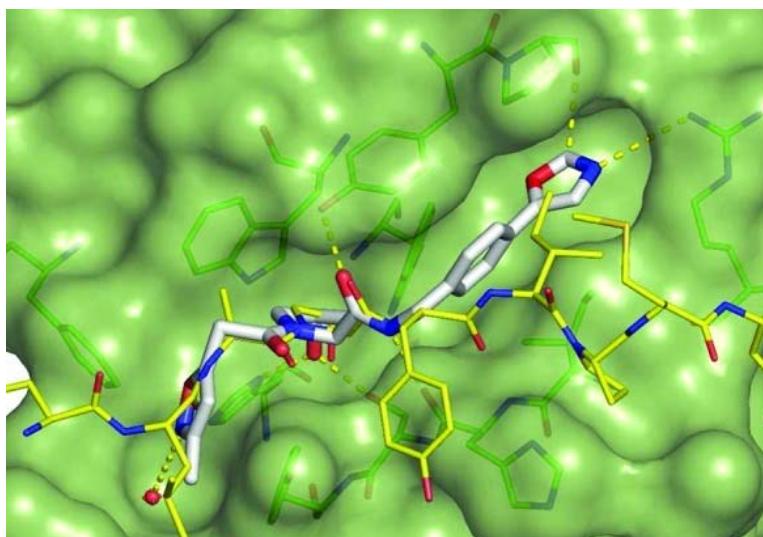
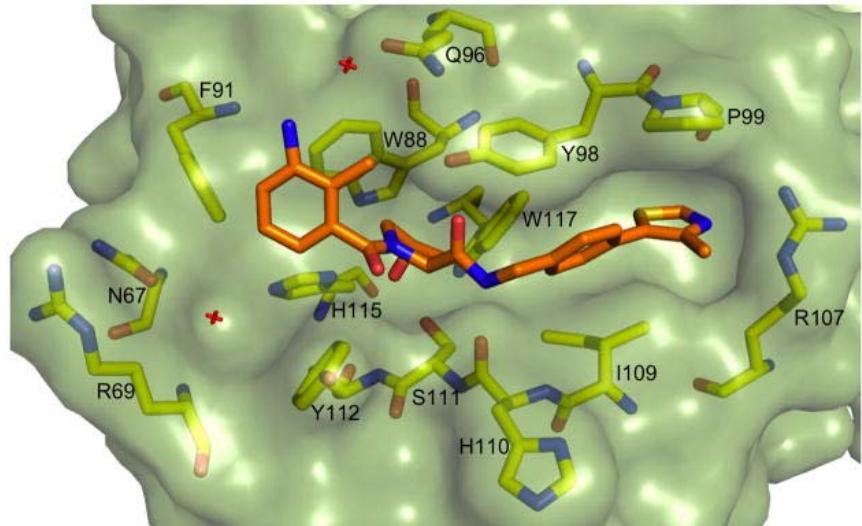
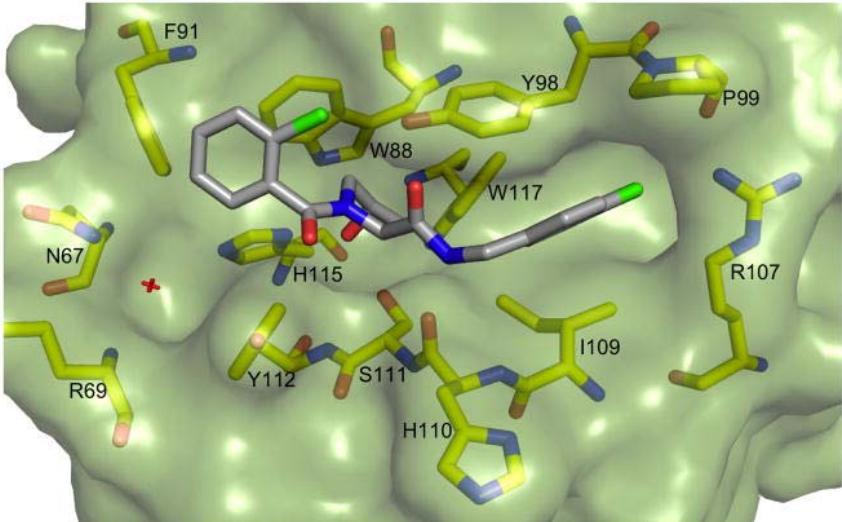
Category	Specific Name	Target (Ligand)	E3 Ligase (Ligand)	Supplement	Proposed Mechanism	Refs.
PROTAC	PROTAC-1	MetAP-2 (ovalicin)	SCF ^β -TRCP (IkBα-phos-peptide)		SCF ^β -TRCP-mediated ubiquitylation and proteasomal degradation of MetAP-2 <i>in vitro</i> .	[50]
	PROTAC-2	ERα (estradiol)	SCF ^β -TRCP (IkBα-phos-peptide)		SCF ^β -TRCP-mediated ubiquitylation and proteasomal degradation of ERα <i>in vitro</i> .	[51]
	PROTAC-3	AR (DHT)	SCF ^β -TRCP (IkBα-phos-peptide)		Proteasomal degradation of the GFP-AR protein in the microinjected cells.	[51]
	PROTAC-4	FKBP12(F36V) (AP21998)	VHL E3 complex (HIF-1α-peptide)	Poly-D-Arg (tag)	VHL-mediated degradation of the EGFP-FKBP12 protein.	[52]
	PROTAC-5	AR (DHT)	VHL E3 complex (HIF-1α-peptide)	Poly-D-Arg (tag)	Proteasomal degradation of the GFP-AR protein.	[52]
	Apigenin-PROTAC	AhR (apigenin)	VHL E3 complex (HIF-1α-peptide)		Proteasomal degradation of the AhR protein. Repression of AhR-dependent transcriptional activation.	[53]
	PROTAC-A PROTAC-AA	AR (DHT)	VHL E3 complex (HIF-1α-peptide)	Poly-D-Arg (tag)(-AA)	Ubiquitylation and proteasomal degradation of AR. Cell cycle arrest of AR-dependent prostate cancer cells.	[54]
	PROTAC-B	ERα (estradiol)	VHL E3 complex (HIF-1α-peptide)		Ubiquitylation and proteasomal degradation of ERα. Cell cycle arrest of ERα-dependent breast cancer cells.	[54]
	SARM-nutlin PROTAC	AR (SARM)	MDM2 (nutlin)		Proteasomal degradation of the AR protein.	[56]
	Phospho-PROTAC (^{TrkA} PP _{FRS2α})	FRS2α (TrkA-peptide)	VHL E3 complex (HIF-1α-peptide)	Poly-D-Arg (tag)	NGF-dependent ubiquitylation and proteasomal degradation of FRS2α. Inhibition of NGF-dependent neuronal differentiation.	[59]
	Phospho-PROTAC (^{ErbB3} PP _{PI3K})	PI3K (ErbB3-peptide)	VHL E3 complex (HIF-1α-peptide)	Poly-D-Arg (tag)	NRG-dependent ubiquitylation and proteasomal degradation of PI3K. Inhibition of ovarian tumor growth in xenograft model.	[59]

Von Hippel Lindau (VHL) – Hypoxia inducible factor 1 α (HIF1 α)



George & Kaelin New Engl J Med 2003

Small molecule inhibitors of VHL– HIF1 α interaction



Development of potent small-molecule inhibitors of the interaction between VHL and HIF1 α

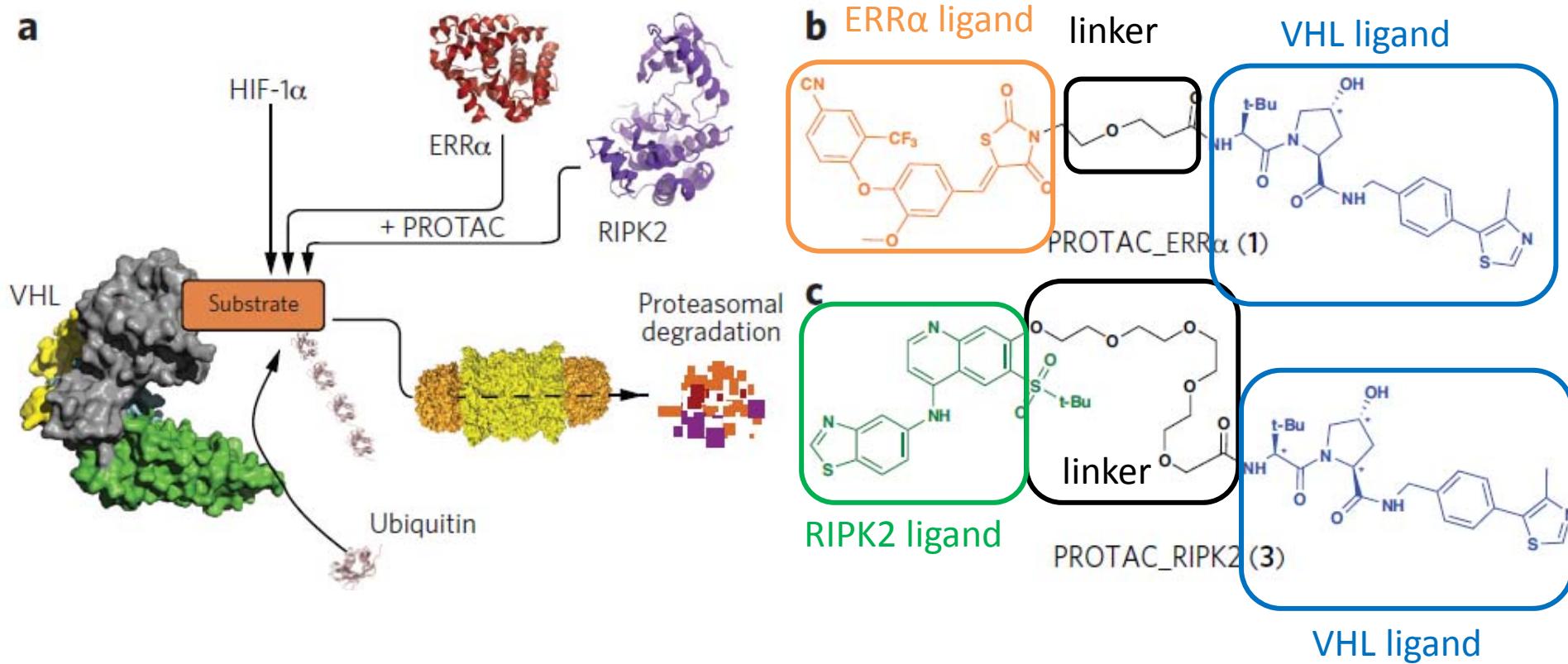
Buckley et al. J Am Chem Soc 2012

Buckley et al. Angew Chem Int Ed Engl 2012

Catalytic *in vivo* protein knockdown by small-molecule PROTACs

Daniel P Bondeson^{1,9}, Alina Mares^{2,9}, Ian E D Smith^{2,9}, Eunhwa Ko¹, Sebastien Campos², Afjal H Miah², Katie E Mulholland², Natasha Routly², Dennis L Buckley¹, Jeffrey L Gustafson¹, Nico Zinn³, Paola Grandi³, Satoko Shimamura³, Giovanna Bergamini³, Maria Faelth-Savitski³, Marcus Bantscheff³, Carly Cox¹, Deborah A Gordon⁴, Ryan R Willard⁴, John J Flanagan⁴, Linda N Casillas⁵, Bartholomew J Votta⁵, Willem den Besten⁶, Kristoffer Famm², Laurens Kruidenier², Paul S Carter², John D Harling², Ian Churcher^{2*} & Craig M Crews^{1,7,8*}

Small molecule PROTACs: overview



VHL ligand designed based on previous structural studies
 K_d : 320 nM

PROTAC_ERR α : design

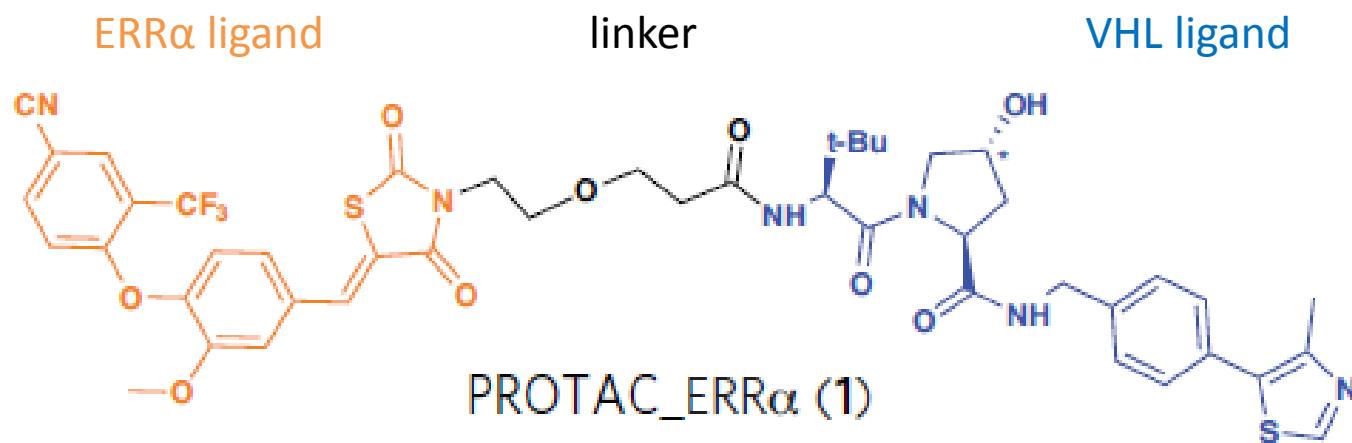
ERR α :

(Estrogen-related receptor α)

- Orphan nuclear hormone receptor
- Master regulator of cellular energy homeostasis

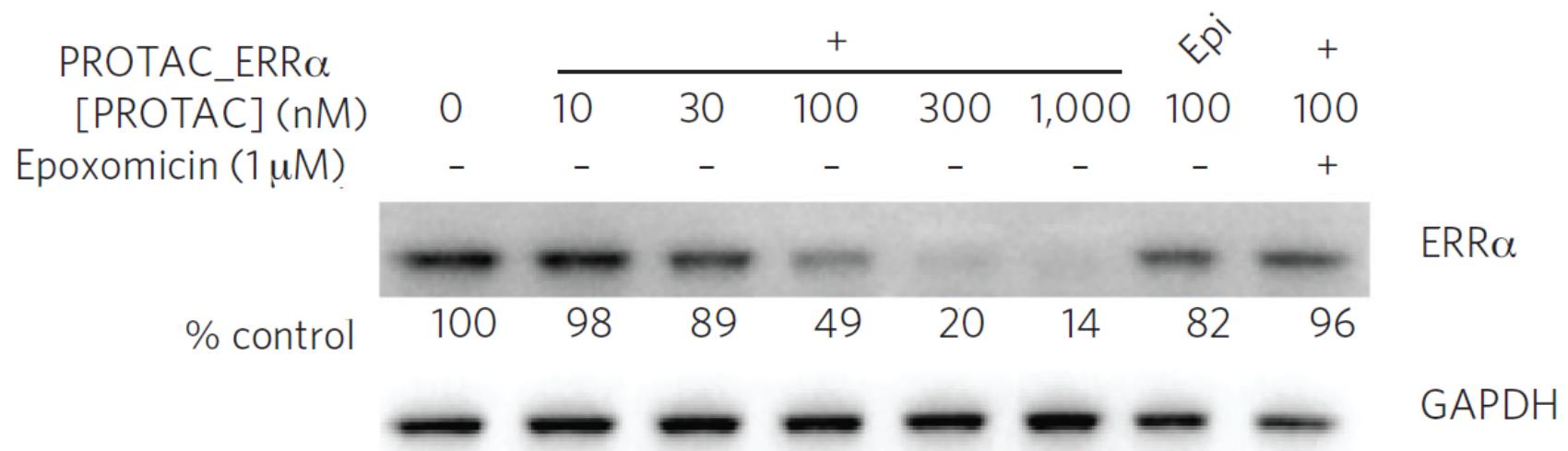
ERR α ligand:

- Thiazolinedione-based
- Selective for binding to ERR α over other ERR isoforms
- Not able to degrade ERR α alone



PROTAC_ERR α : functionality

In MCF-7 breast cancer cell line

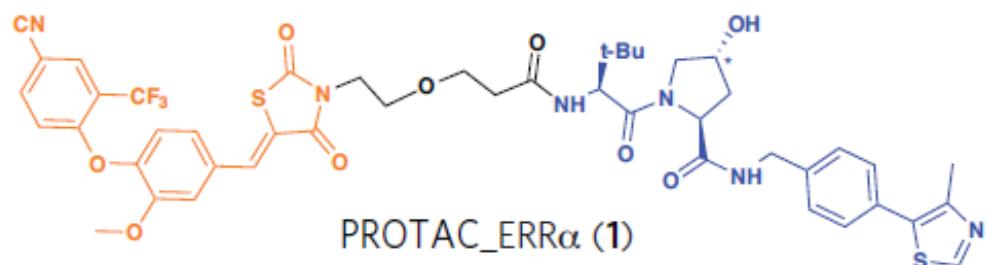


PROTAC and proteasome required

DC₅₀ of PROTAC_ERR : \approx 100 nM

Epi: PROTAC_ERR α epi (inverted stereochemistry at *)

Epoxomicin: Proteasome inhibitor



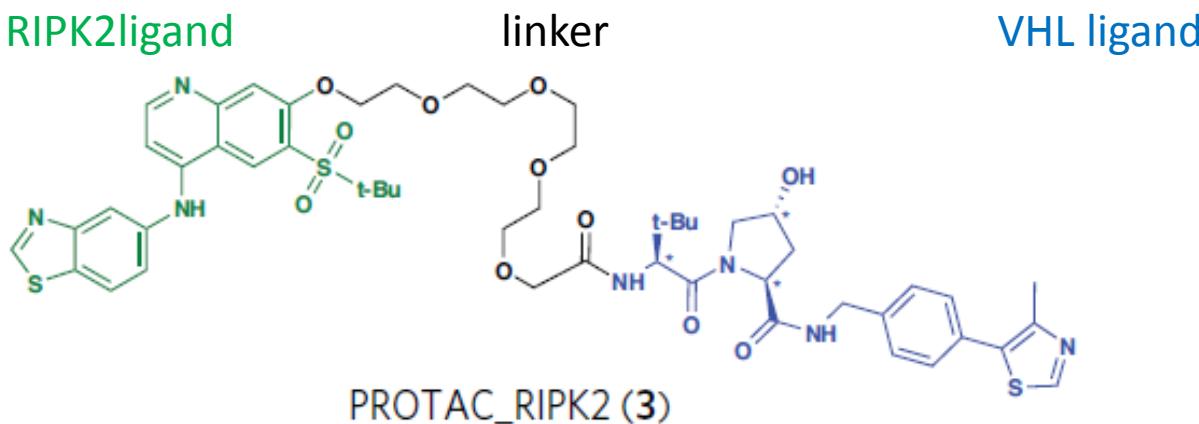
PROTAC_RIPK2: design

RIPK2:

- Serine-threonine kinase
- Important mediator of innate immune signaling
- Involved in NF-κB and MAPK activation
- Implicated in Blau syndrome and sarcoidosis

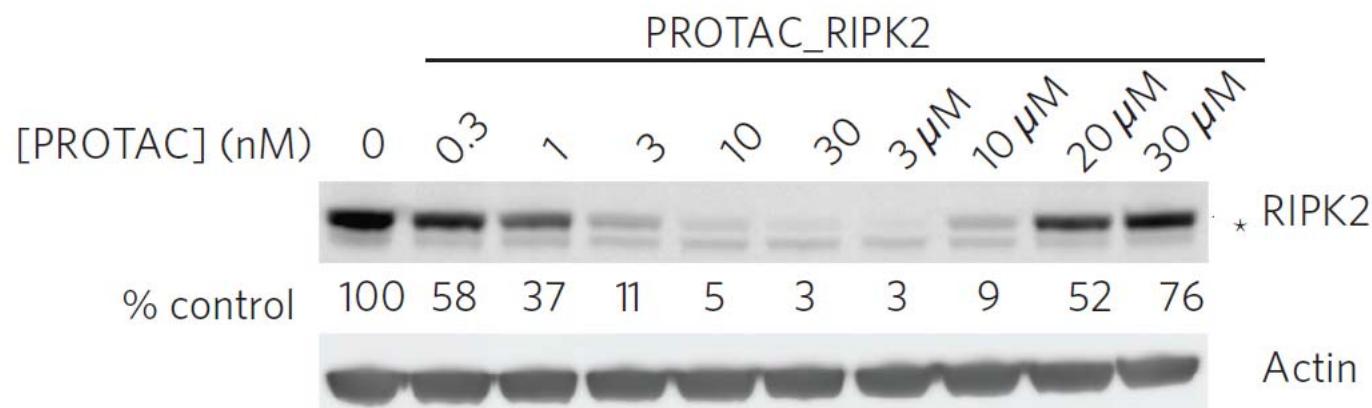
RIPK2 ligand:

- Modification of a previously identified inhibitor
- Optimized through study of structure-activity relationship



PROTAC_RIPK2: functionality I

In THP-1 human monocyte cell line

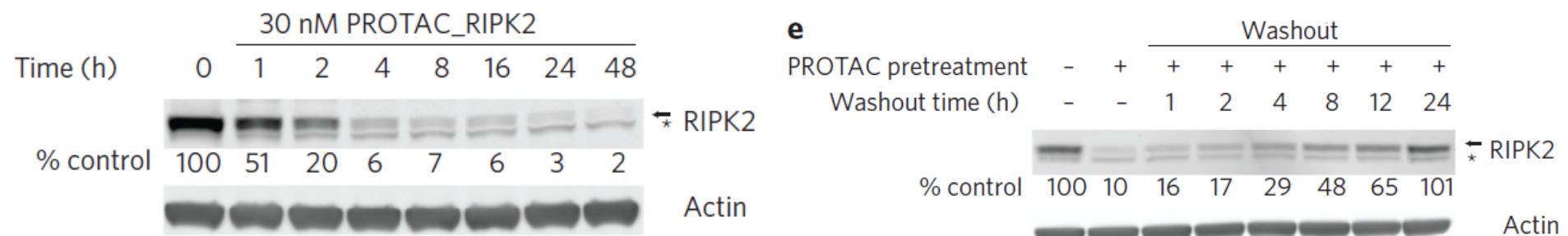


→ DC₅₀ of PROTAC_RIPK2 : ≈ 1.4 nM

→ Biphasic response

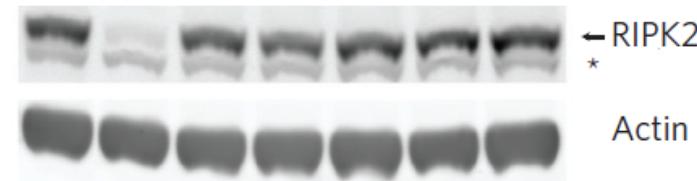
PROTAC_RIPK2: functionality II

In THP-1 human monocyte cell line



Early degradation
(RIPK2 half life is ≈60h)

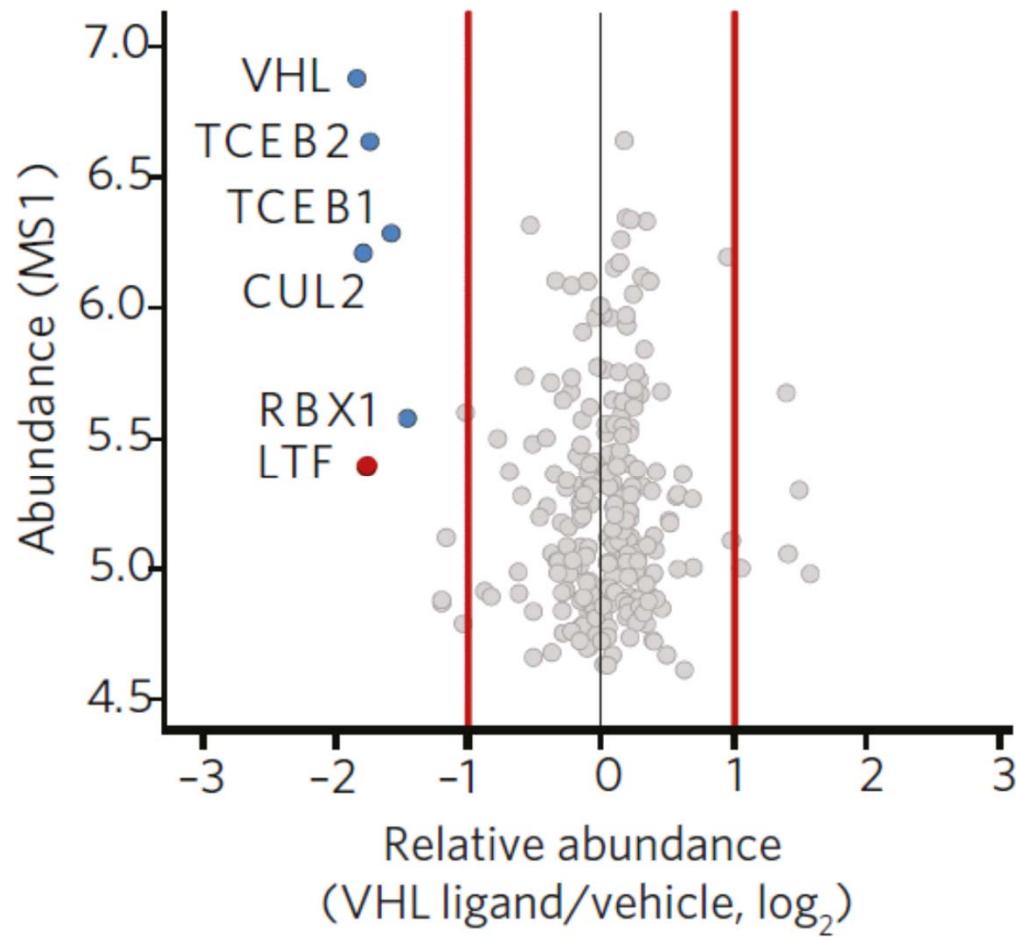
RIPK2 levels recover after PROTAC washout



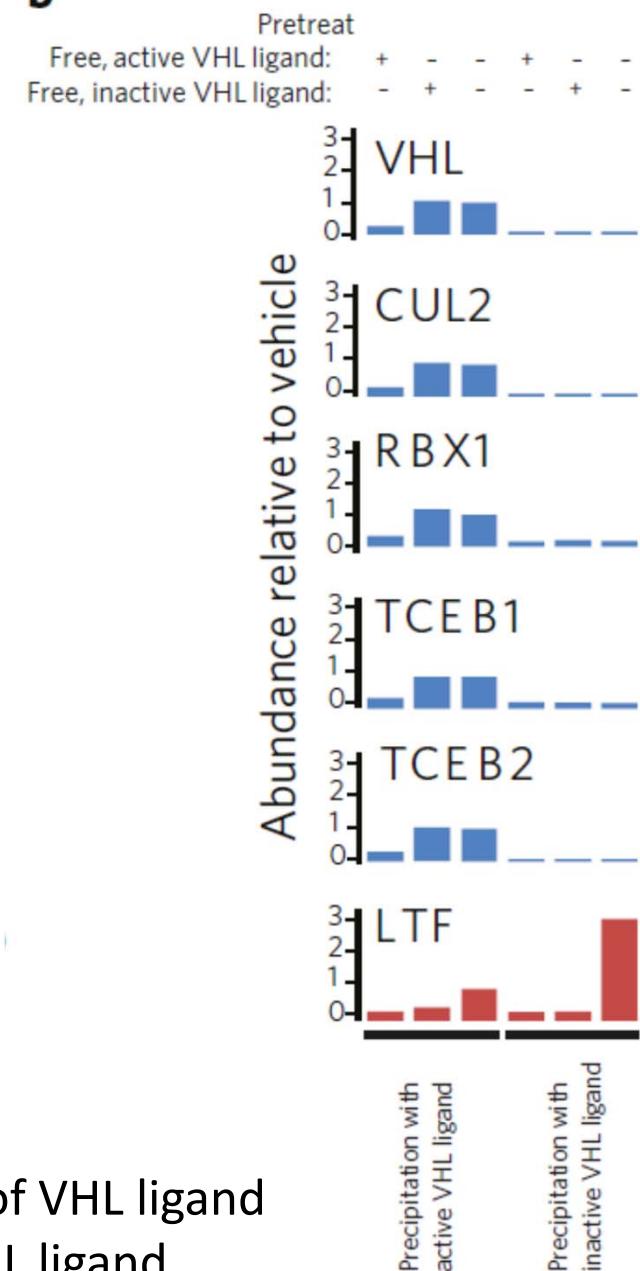
PROTAC and proteasome required
RIPK2 ligand and VHL ligand alone insufficient

PROTAC: specificity of VHL ligand

Chemoproteomic pulldown with VHL ligand



b

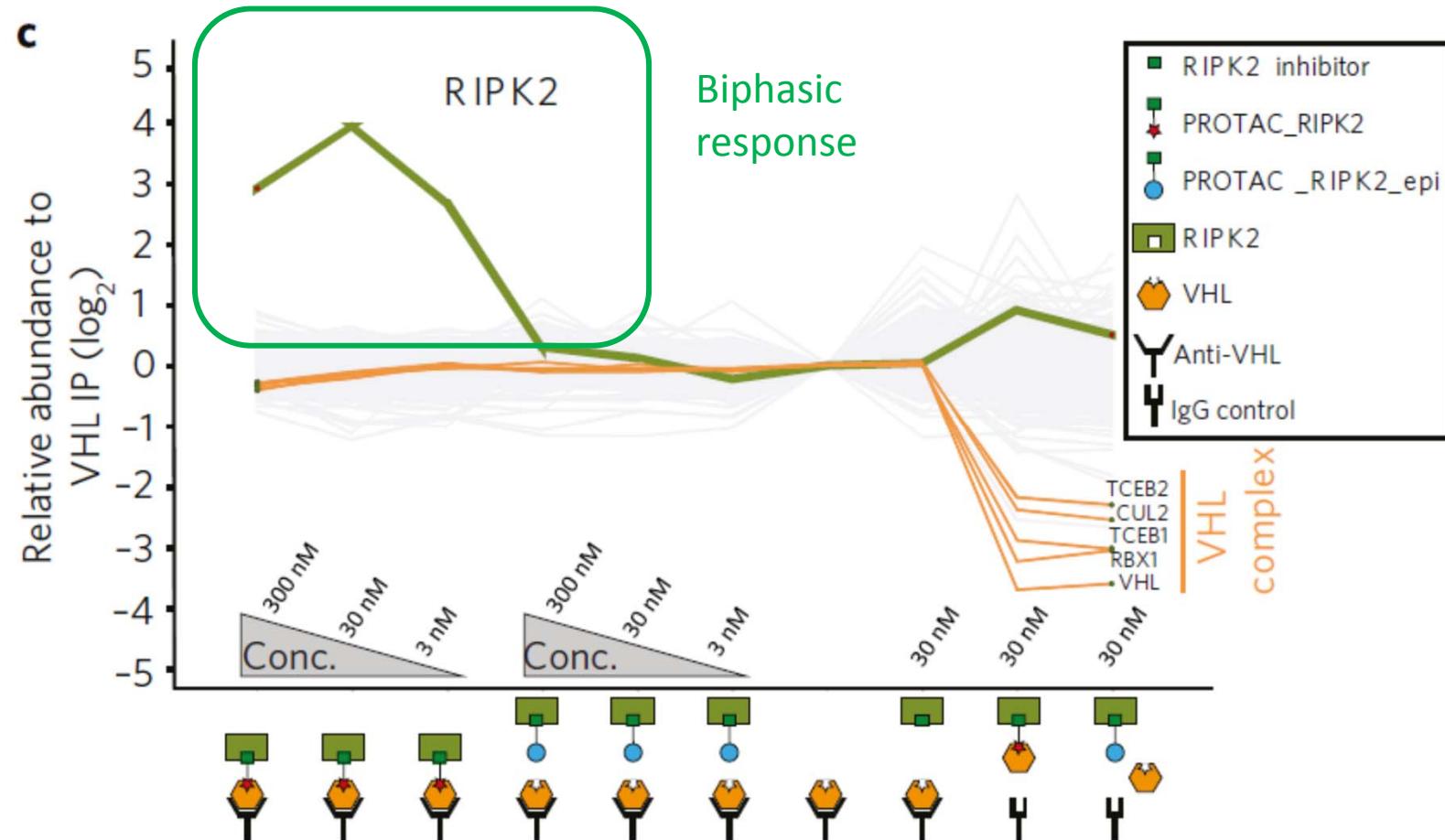


Member of the VHL complex are the only major targets of VHL ligand
LTF: unspecific binding, also with the control, inactive VHL ligand

PROTAC: ternary complex I

Immunoprecipitation of VHL from THP-1 lysates

Multiplex quantitative MS analysis with isobaric mass tags (TMT10 labels)

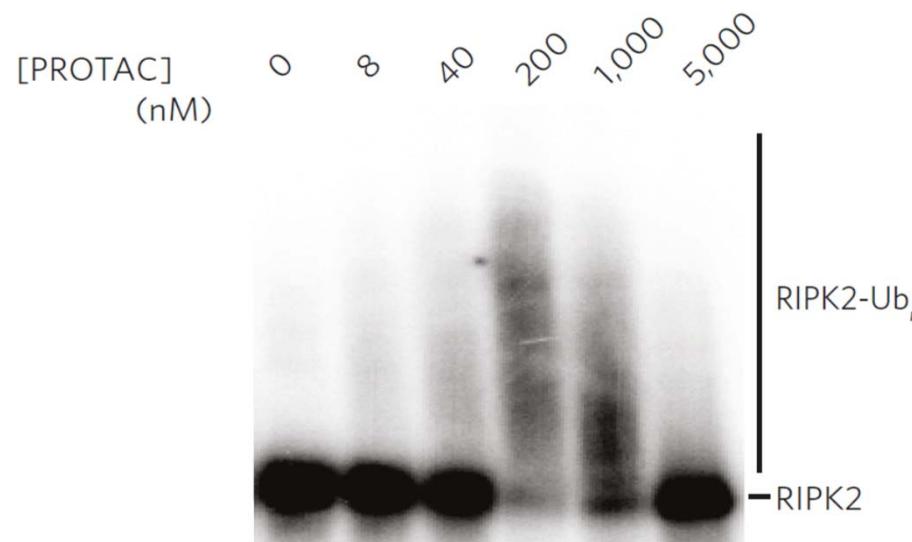


In the presence of PROTAC_RIPK2, not PROTAC_RIPK2_epi, there is co-immunoprecipitation of VHL and RIPK2

PROTAC: ternary complex II

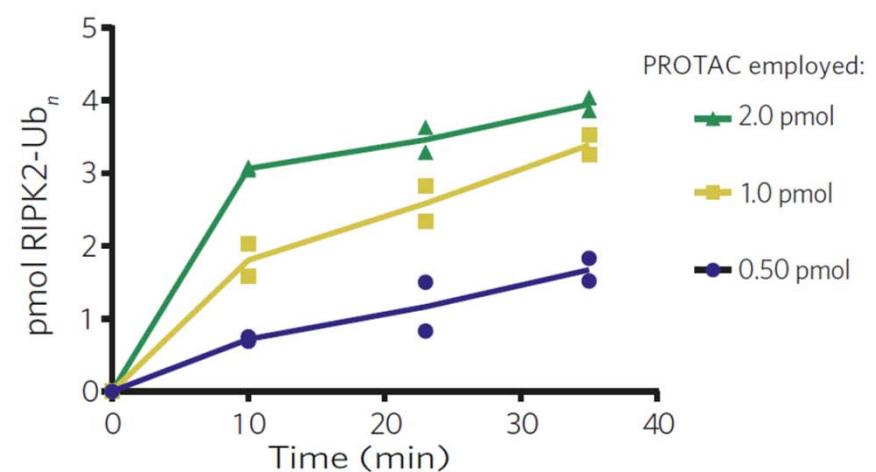
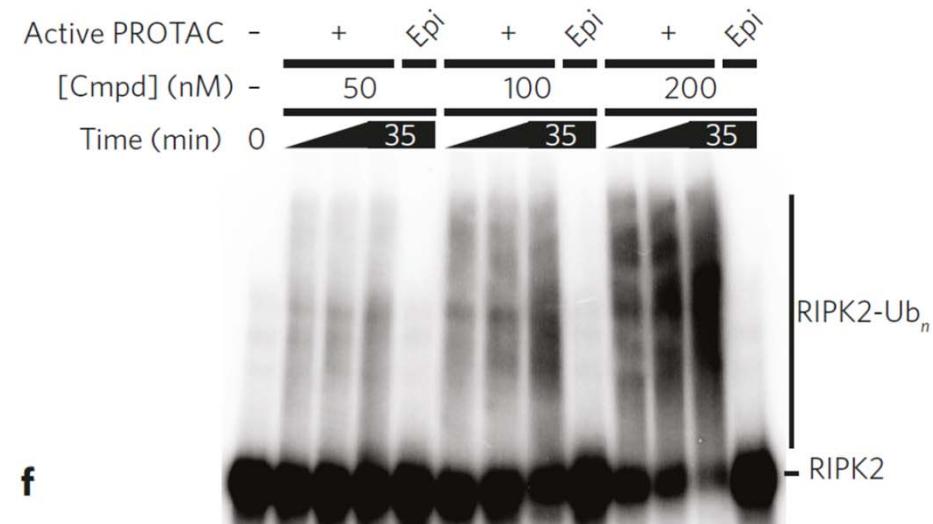
In vitro ubiquitination of radiolabelled RIPK2

Quantification through PAGE/autoradiography and liquid scintillation of excised bands



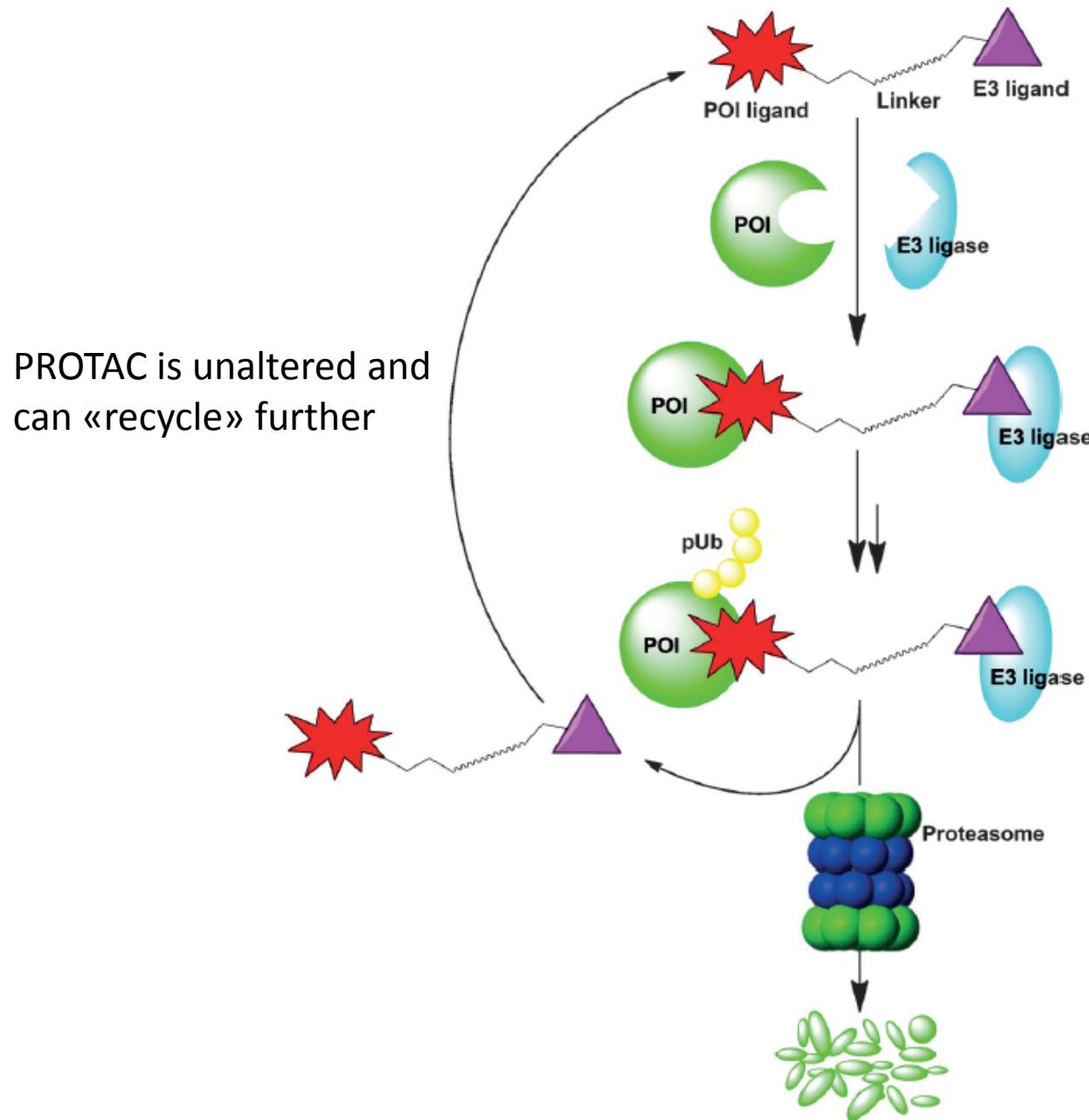
PROTAC required for ubiquitination
Biphasic response

Sub-stoichiometric catalysis



PROTAC (pmol):	0.5	1.0	2.0
RIPK2-Ub (pmol):	1.7	3.4	4.0
Stoichiometry:	3.3	3.4	2.0

PROTAC: catalytically active

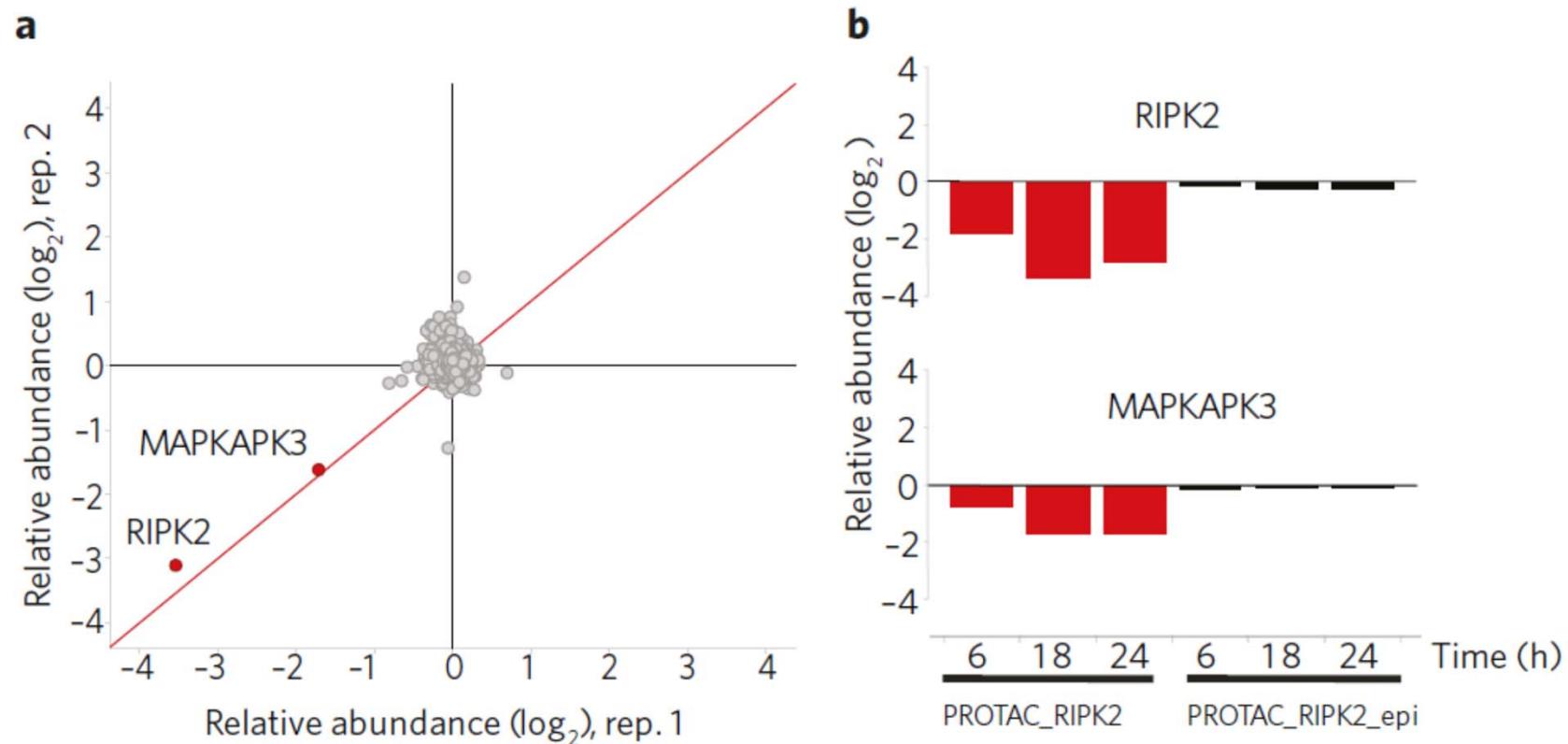


PROTAC_RIPK2: specificity for RIPK2 degradation

Cellular expression proteomics

THP-1 treated with PROTAC_RIPK2, PROTAC_RIPK2_epi or RIPK2 alone

≈7000 quantified proteins

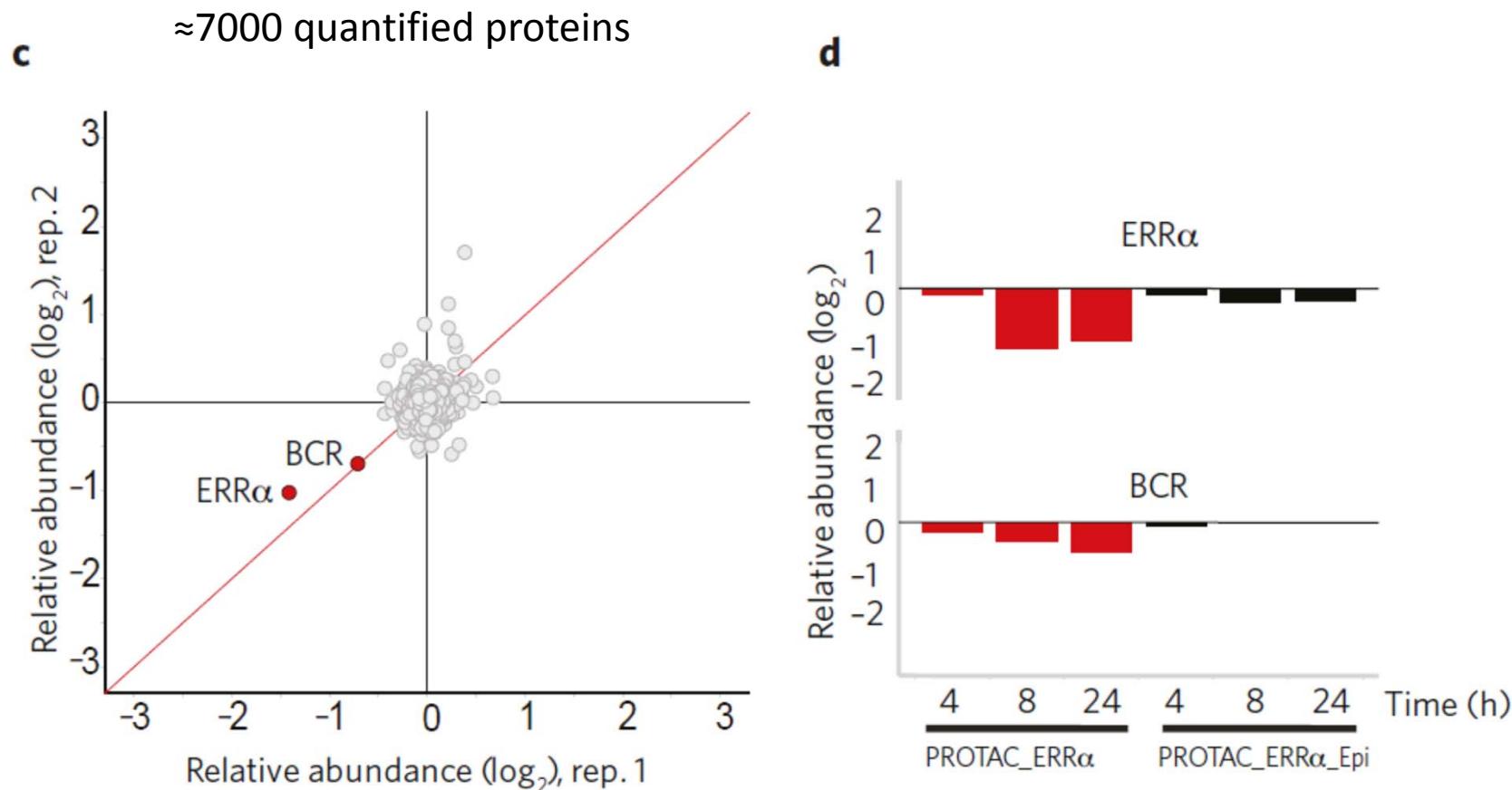


Only RIPK2 and the unrelated MAPKAPK3 were underrepresented in PROTAC_RIPK2 treated cells

PROTAC_ERR α : specificity for ERR α degradation

Cellular expression proteomics

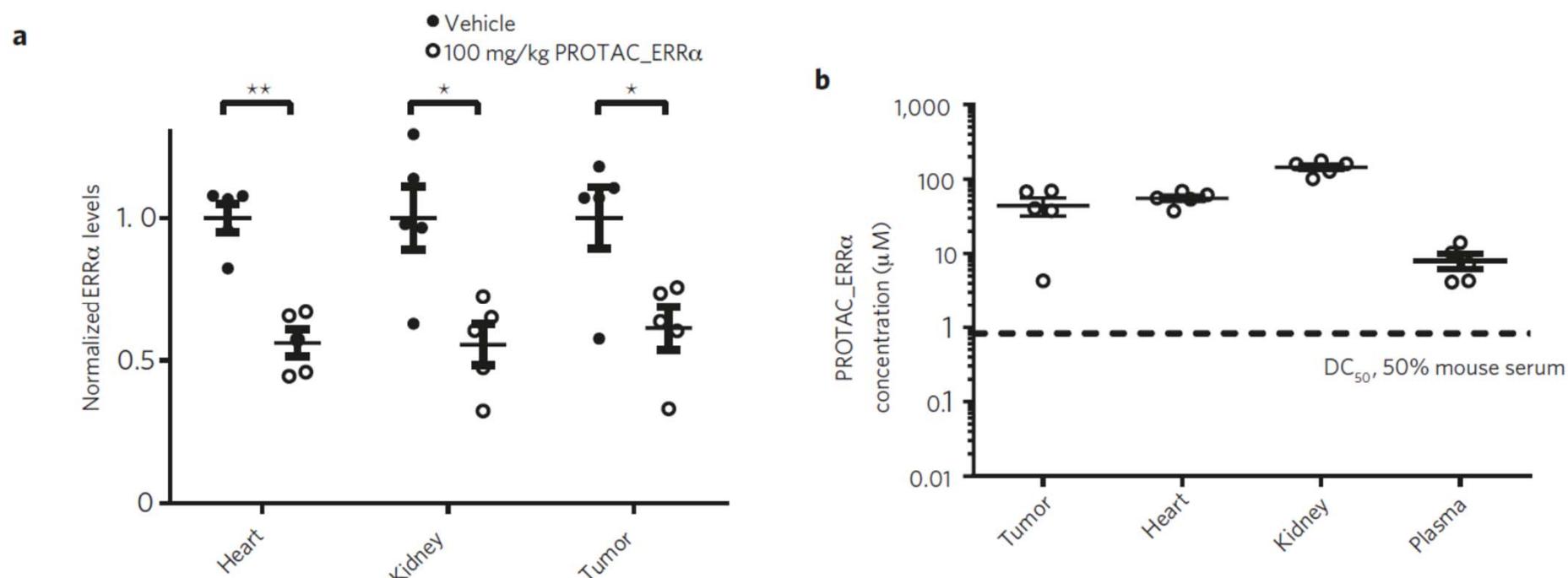
MCF-7 treated with PROTAC_ERR α or PROTAC_ERR α _epi



Only ERR α and the unrelated BCR were underrepresented in PROTAC_ERR α treated cells

PROTAC_ERR α : *in vivo* activity

In mice bearing MDA-MB-231 tumors
PROTAC_ERR α (100 mg/Kg) or vehicle, 3x daily, i.p.



Reduced ERR α levels in different tissues of PROTAC_ERR α -treated mice

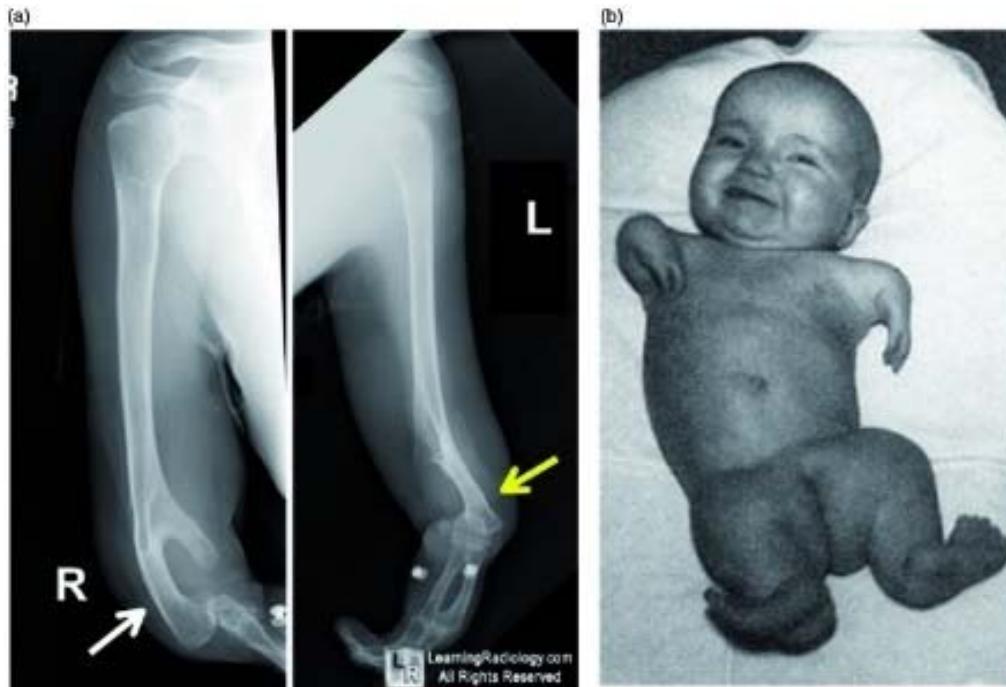
Protein knockdown

1) PROTACs

2) Phtalimide conjugation

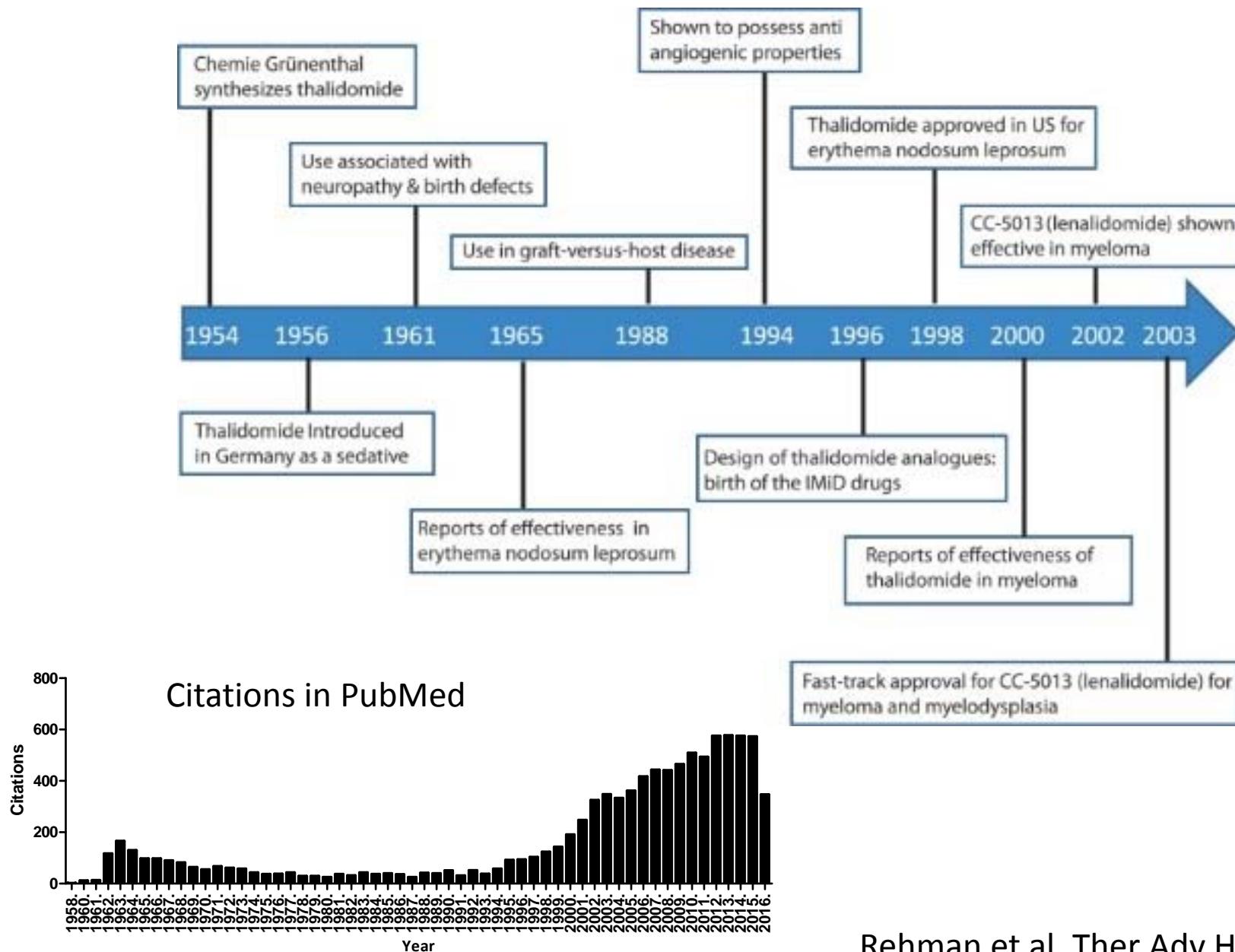
Thalidomide: the rise and fall... and rise

- Synthetic glutamic-acid derivative
- Sedative and anti-emetic activity
- Prescribed also to pregnant women for morning sickness
- In 1961, reports of birth defects led to market withdrawal



Bartlett et al. Nat Rev Cancer 2004
Rehman et al. Ther Adv Hematol 2011

Thalidomide: the rise and fall... and rise

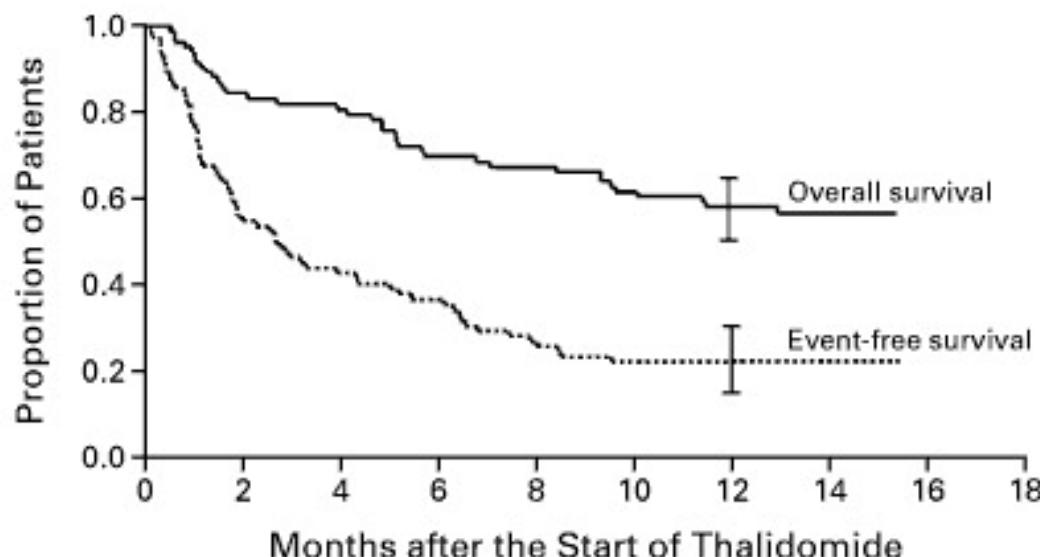




ANTITUMOR ACTIVITY OF THALIDOMIDE IN REFRACTORY MULTIPLE MYELOMA

ANTITUMOR ACTIVITY OF THALIDOMIDE IN REFRACTORY
MULTIPLE MYELOMA

SEEMA SINGHAL, M.D., JAYESH MEHTA, M.D., RAMAN DESIKAN, M.D., DAN AYERS, M.S., PAULA ROBERSON, PH.D.,
PAUL EDDLEMON, B.S., NIKHIL MUNSHI, M.D., ELIAS ANAISSE, M.D., CARLA WILSON, M.D., PH.D.,
MADHAV DHODAPKAR, M.D., JEROME ZELDIS, M.D., AND BART BARLOGIE, M.D., PH.D.



No. AT RISK

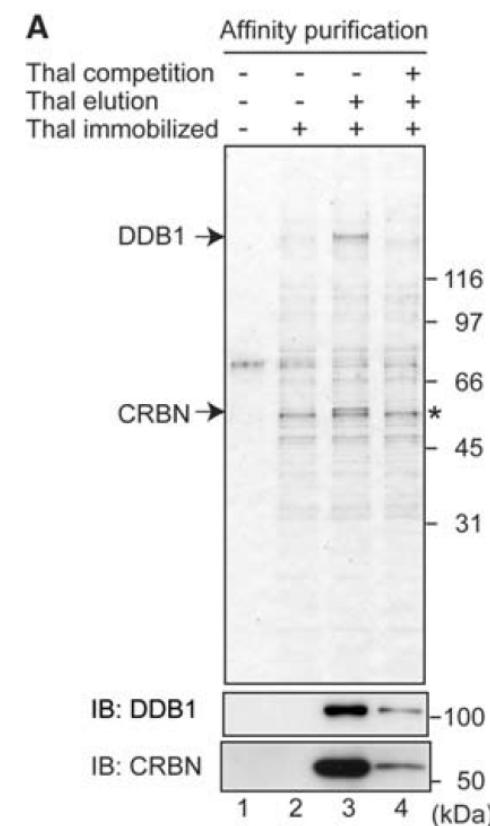
Overall survival	84	78	69	64	58	56	51	34
Event-free survival	84	65	39	32	24	19	18	11

Identification of a Primary Target of Thalidomide Teratogenicity

Takumi Ito,^{1,*} Hideki Ando,^{2,*} Takayuki Suzuki,^{3,4} Toshihiko Ogura,³ Kentaro Hotta,² Yoshimasa Imamura,⁵ Yuki Yamaguchi,² Hiroshi Handa^{1,2†}

Thalidomide binds to cereblon (CRBN), a component of a cullin-RING ubiquitin ligase complex

CRBN binding mediates thalidomide teratogenicity



Lenalidomide Causes Selective Degradation of IKZF1 and IKZF3 in Multiple Myeloma Cells

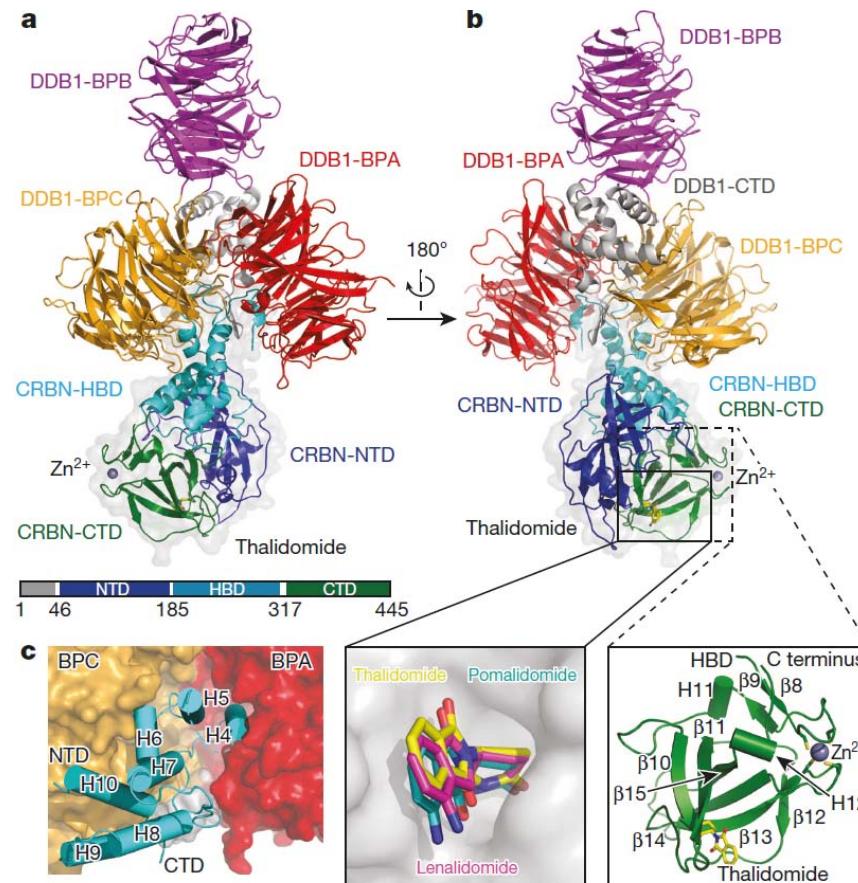
Jan Krönke,¹ Namrata D. Udeshi,² Anupama Narla,¹ Peter Grauman,¹ Slater N. Hurst,¹ Marie McConkey,¹ Tanya Svinkina,² Dirk Heckl,¹ Eamon Comer,² Xiaoyu Li,² Christie Ciarlo,² Emily Hartman,² Nikhil Munshi,³ Monica Schenone,² Stuart L. Schreiber,² Steven A. Carr,² Benjamin L. Ebert^{1,2*}

The Myeloma Drug Lenalidomide Promotes the Cereblon-Dependent Destruction of Ikaros Proteins

Gang Lu,¹ Richard E. Middleton,^{1,2} Huahang Sun,^{1,2} Mark Vic Naniong,^{1,2} Christopher J. Ott,¹ Constantine S. Mitsiades,¹ Kwok-Kin Wong,^{1,2} James E. Bradner,¹ William G. Kaelin Jr.^{1,3*}

Structure of the DDB1–CRBN E3 ubiquitin ligase in complex with thalidomide

Eric S. Fischer^{1,2}, Kerstin Böhm^{1,2}, John R. Lydeard³, Haidi Yang⁴, Michael B. Stadler^{1,2,5}, Simone Cavadini^{1,2}, Jane Nagel⁴, Fabrizio Serluca⁴, Vincent Acker⁶, Gondichatnahalli M. Lingaraju^{1,2}, Ritesh B. Tichkule⁴, Michael Schebesta⁴, William C. Forrester⁴, Markus Schirle⁴, Ulrich Hassiepen⁶, Johannes Ottl⁶, Marc Hild⁴, Rohan E. J. Beckwith⁴, J. Wade Harper³, Jeremy L. Jenkins⁴ & Nicolas H. Thomä^{1,2}



DRUG DEVELOPMENT

Phthalimide conjugation as a strategy for *in vivo* target protein degradation

Georg E. Winter,^{1,*} Dennis L. Buckley,^{1,*} Joshiawa Paulk,¹ Justin M. Roberts,¹
Amanda Souza,¹ Sirano Dhe-Paganon,² James E. Bradner^{1,3†}

BRD4 as a target

- Transcriptional coactivator
- Binds to acetylated Lys of:
 - histone proteins
 - transcription factors
- Its downregulation leads:
 - *MYC* downregulation
 - antiproliferation response
- Implicated in:
 - Cancer
 - Inflammation
 - Heart disease
- Desirable target for selective degradation

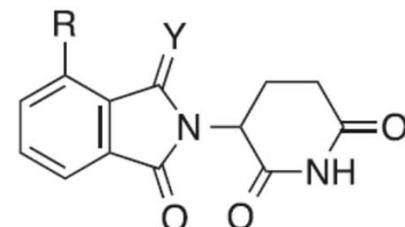
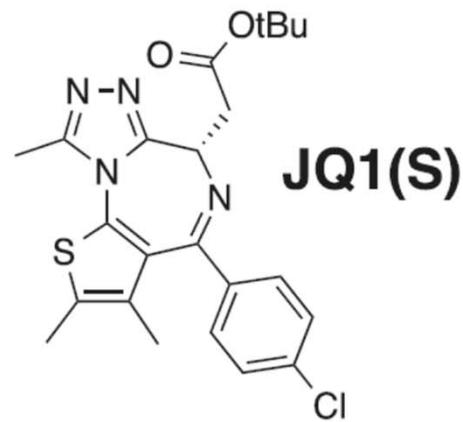
dBET1 to target BRD4

JQ1 displaces BRD4 from chromatin

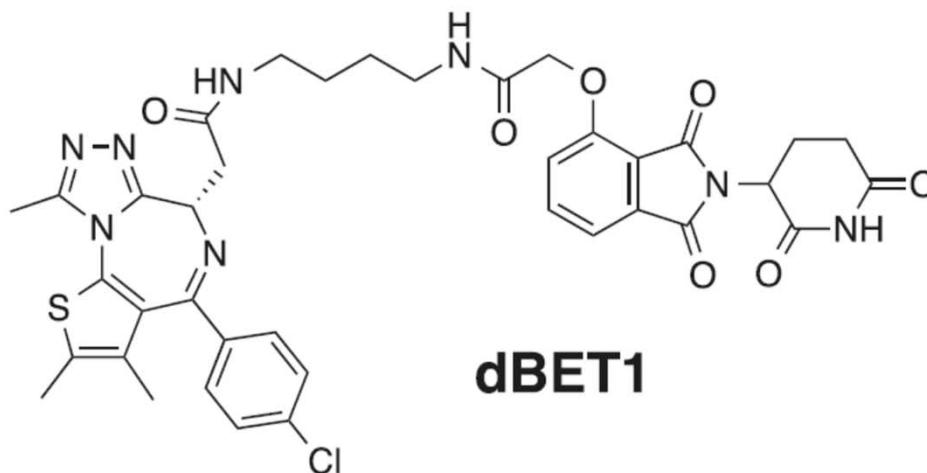
Carboxyl group tolerates chemical substitution

Thalidomide binds CRBN ligase

Aryl ring tolerates chemical substitution



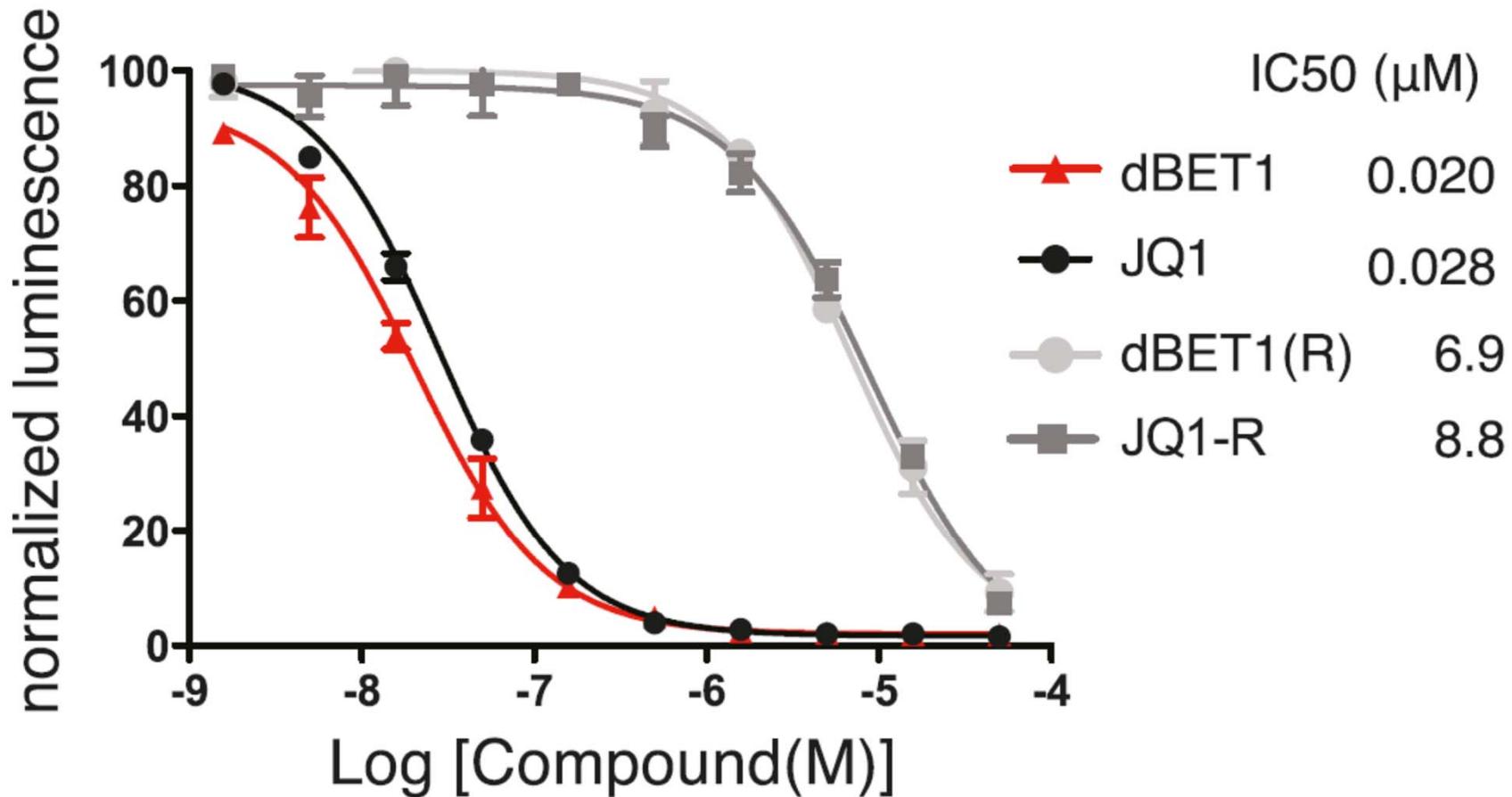
thalidomide: $\text{Y}=\text{O}, \text{R}=\text{H}$
lenalidomide: $\text{Y}=\text{CH}_2, \text{R}=\text{NH}_2$
pomalidomide: $\text{Y}=\text{O}, \text{R}=\text{NH}_2$



→ dBET1 was designed to have preserved BRD4 and

dBET1 binding to BRD4: affinity

BRD4 displacement assay



→ dBET1, not the stereochemical control dBET1(R), has preserved BRD4 binding

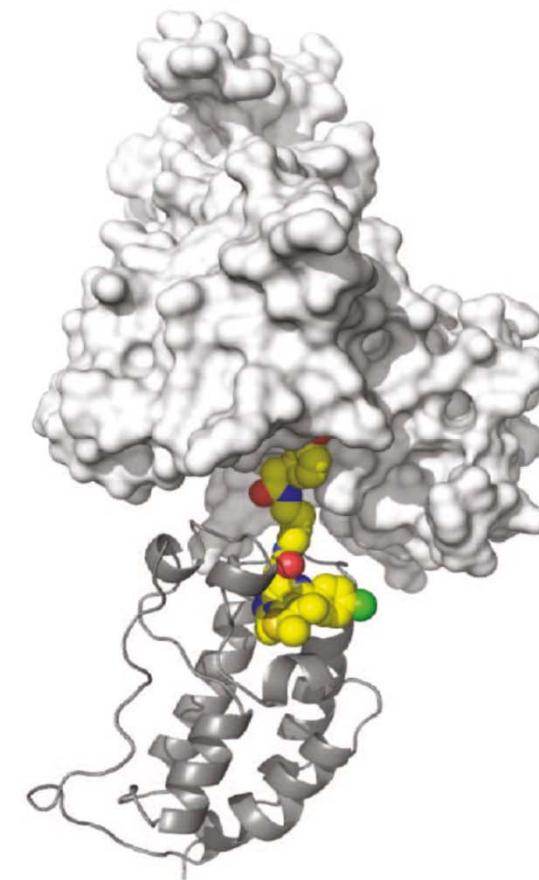
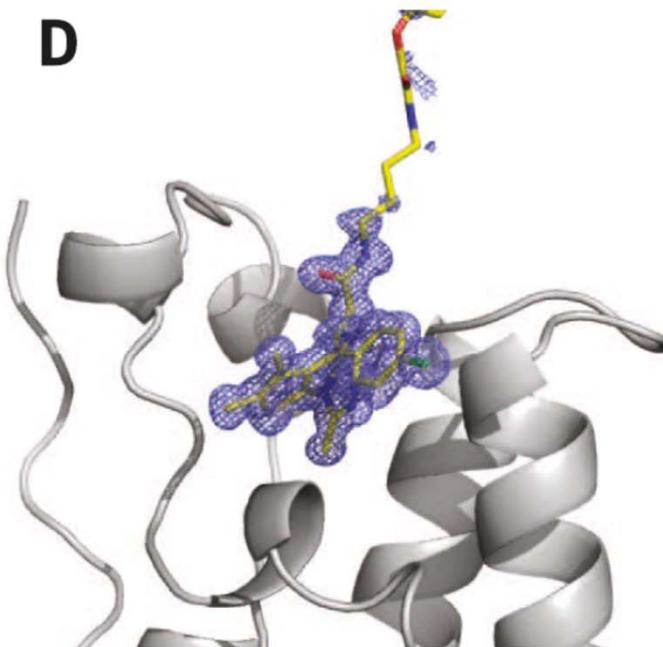
dBET1 binding to BRD4: structure

High-resolution crystal structure (1.0 Å)
of dBET1 bound to BRD4:

Molecular recognition similar to JQ1

Docking of D into published structure
of CRBN bound to thalidomide:

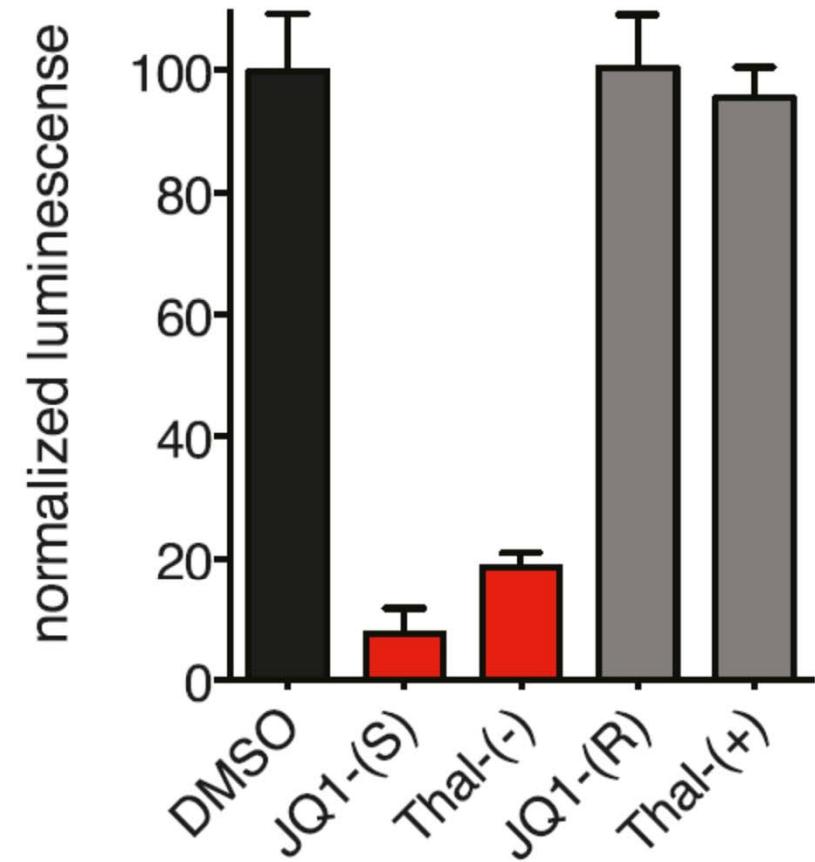
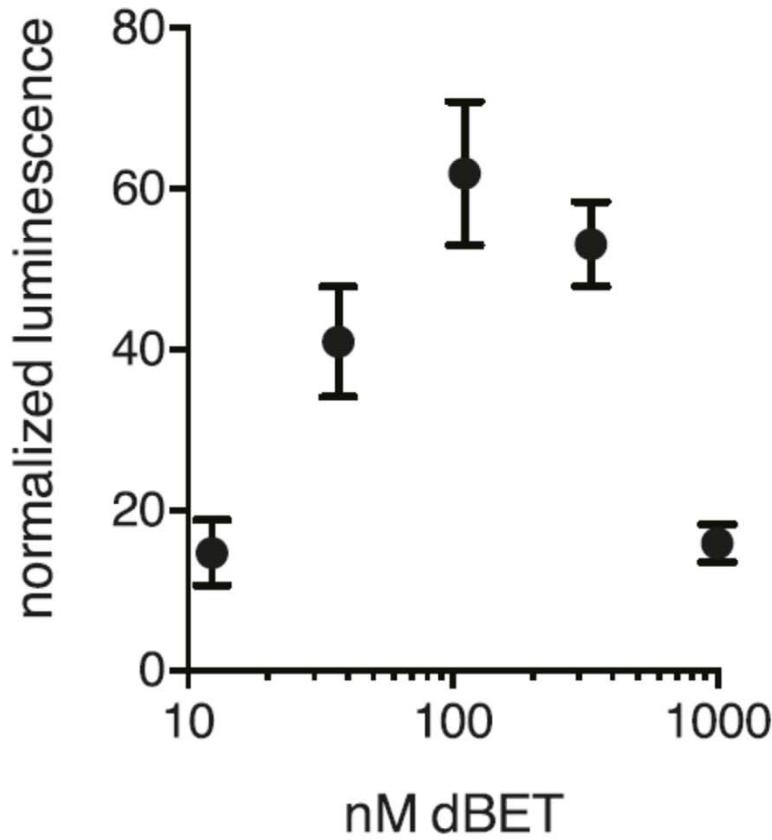
dBET1 can bridge BRD4 and CRBN
without destructive steric interactions



dBET1 binding to BRD4: ternary complex formation

Homogeneous proximity assay

Luminescence arises from proximity of CRBN complex and BRD4-bound acceptor beads

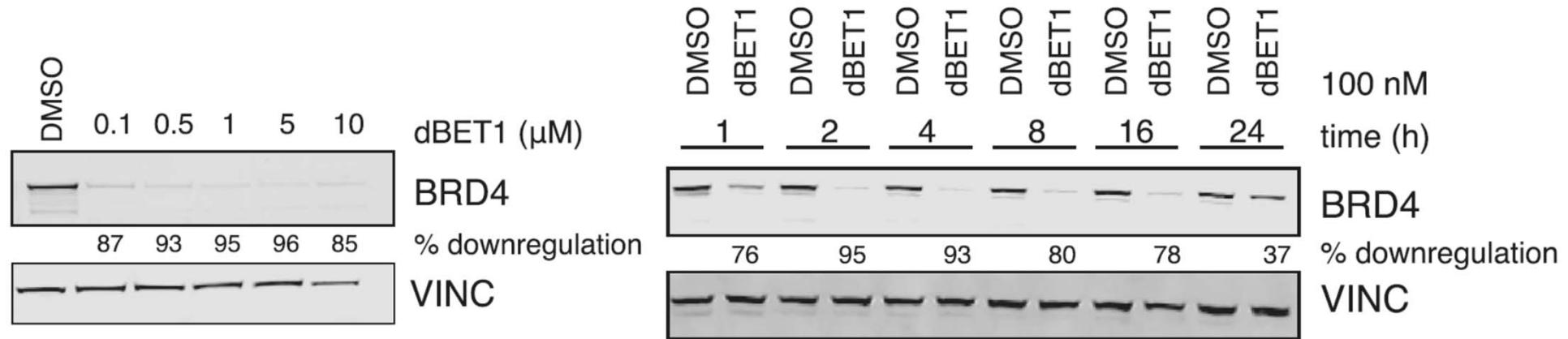


Biphasic response

Free JQ1 or thalidomide inhibits ternary complex formation in a stereospecific manner

dBET1: effect on cells

In MV4;11 cells (human AML cell line)



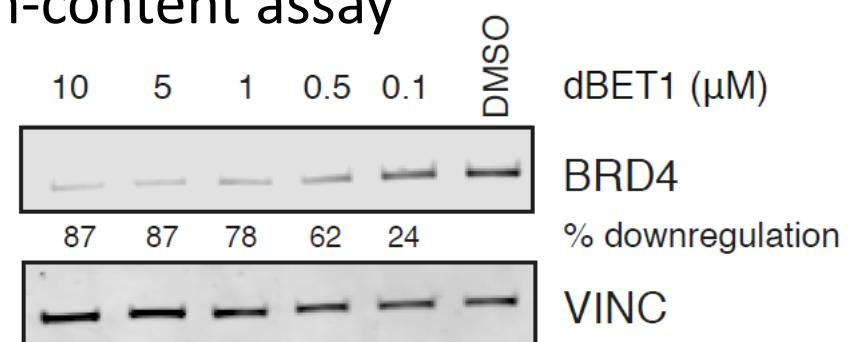
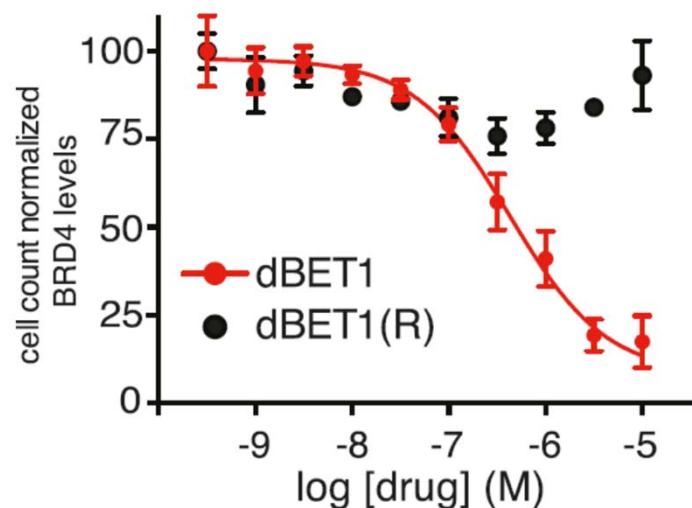
RIPK2 levels

Early degradation

Partial recovery of BRD4 levels at 24h: dBET1 instability?

In SUM149 cells (human breast cancer cell line)

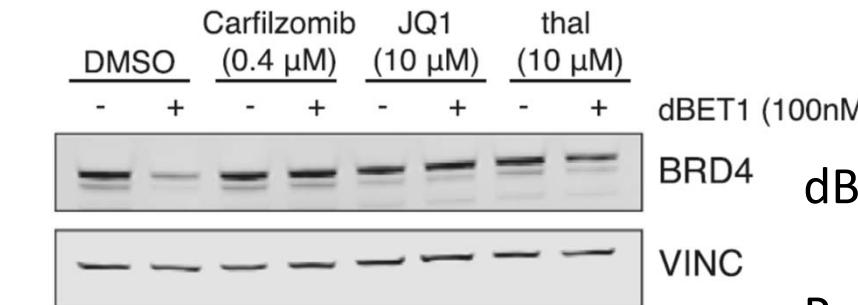
Based on cell-count normalized, high-content assay



Half maximal effective dose (EC_{50}): 430 nM
dBET1(R) inactive

dBET1: mechanism of action

In MV4;11 cells (human AML cell line)

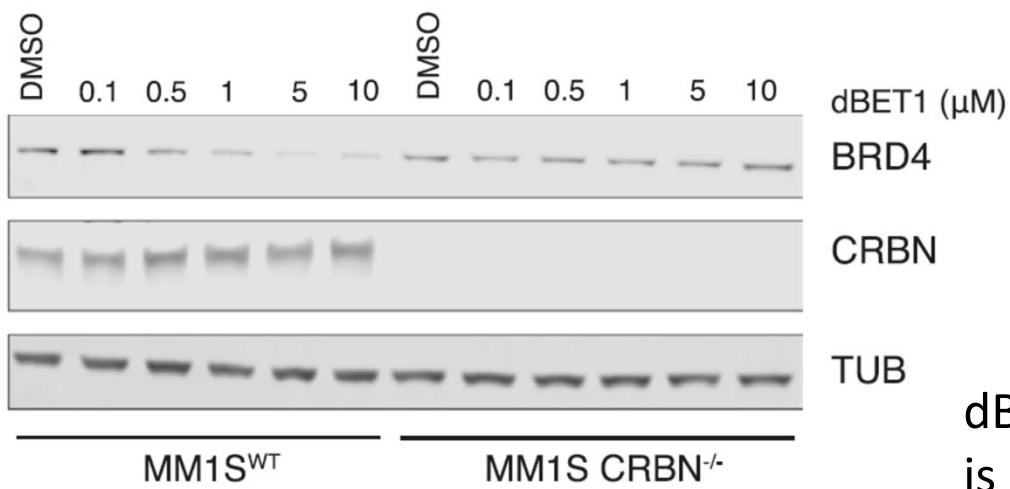


Carfilzomib: proteasome inhibitor

dBET1 and proteasome required

Pre-treatment with JQ1 or thalidomide prevent dBET1-mediated BRD4 downregulation
→ Requirement for both BRD4 and CBN engagement

In MM1.S cells (human MM cell line, either WT or CRBN^{-/-})



dBET1-mediated BRD4 downregulation
is CRBN-dependent

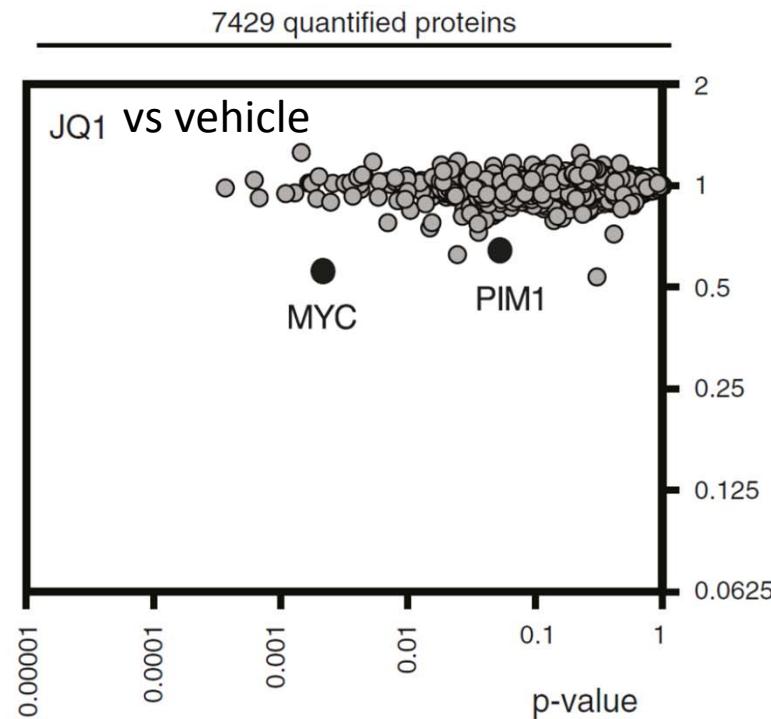
dBET1: specificity

Cellular expression proteomics

MV4;11 treated with dBET1, JQ1, or vehicle for 2 h

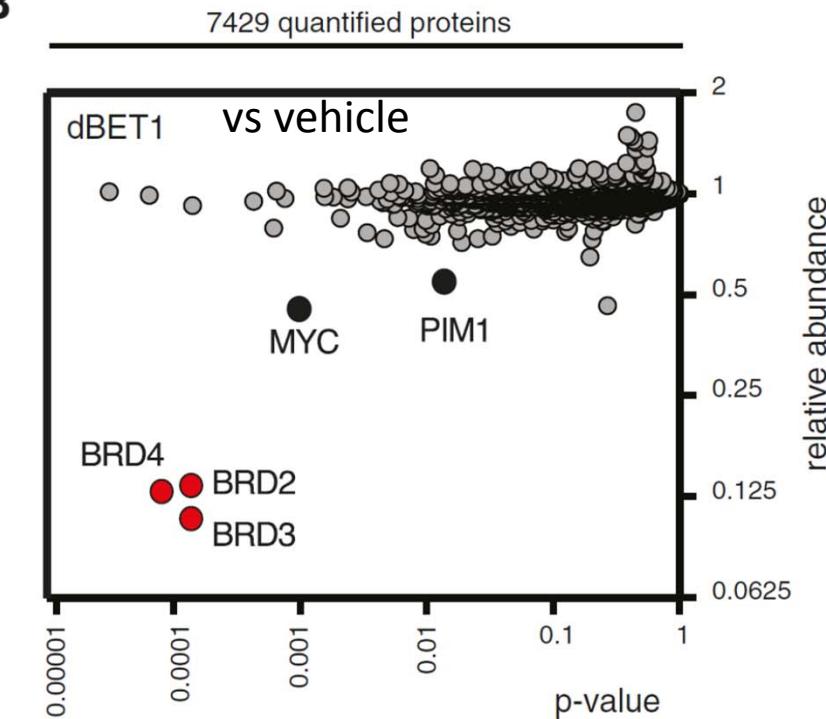
Multiplex quantitative MS analysis with isobaric mass tags

A



JQ1 resulted in MYC and PIM1
downregulation

B



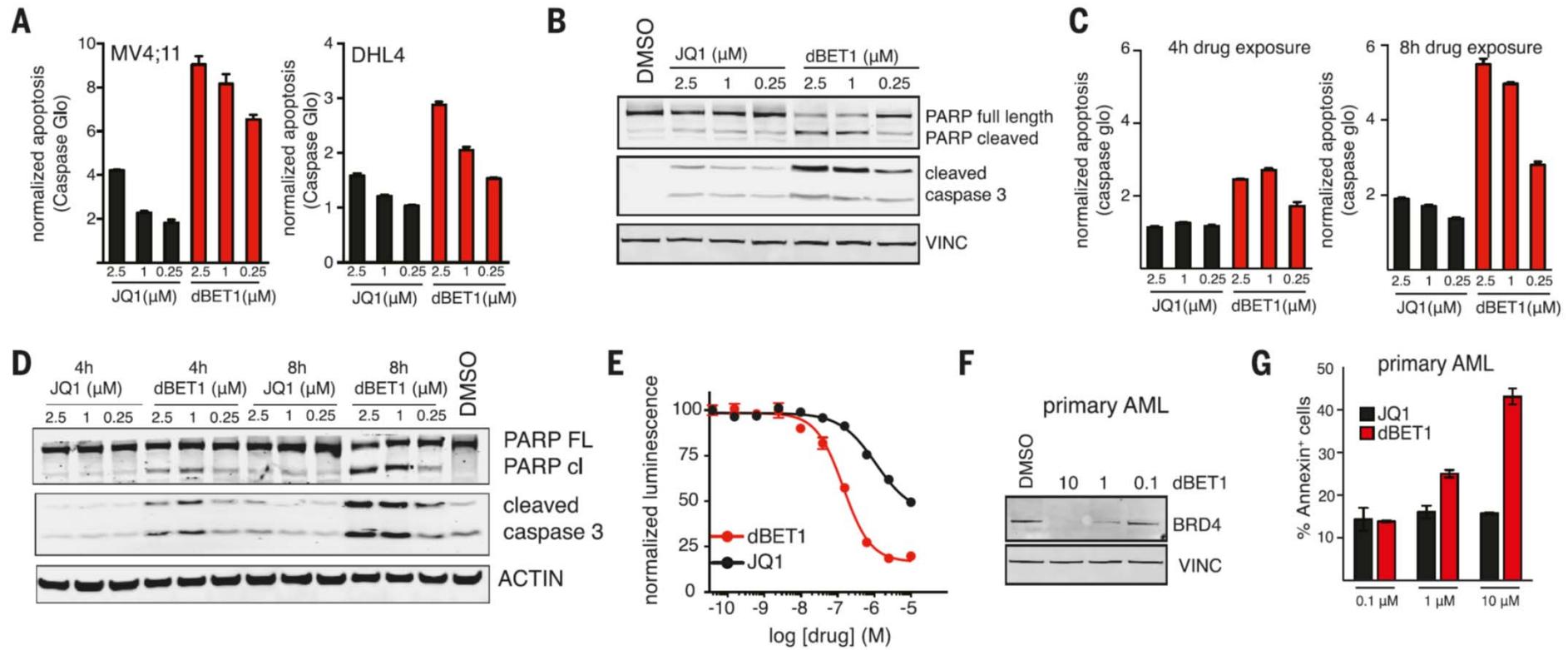
dBET1 resulted in MYC, PIM1 and BRD2, 3 and 4
downregulation

Changes in BRD4 at protein, not transcript level, as confirmed by immunoblot and RT-PCR

dBET1: anti-proliferative response *in vitro*

MV4;11 cells: human AML cell line

DHL4: human lymphoma cell line

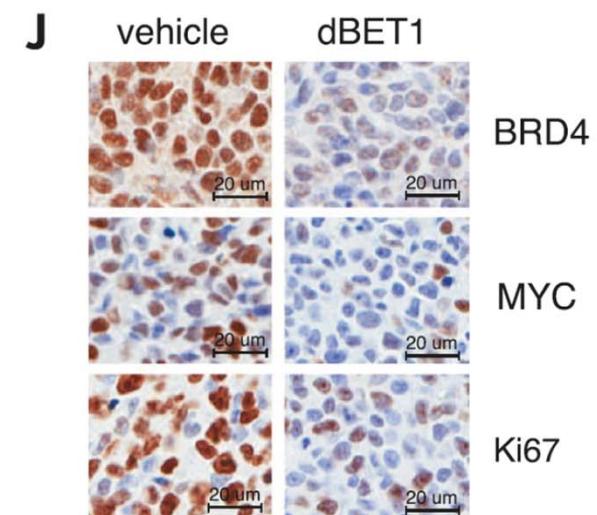
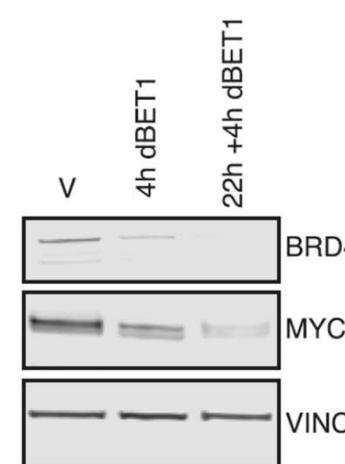
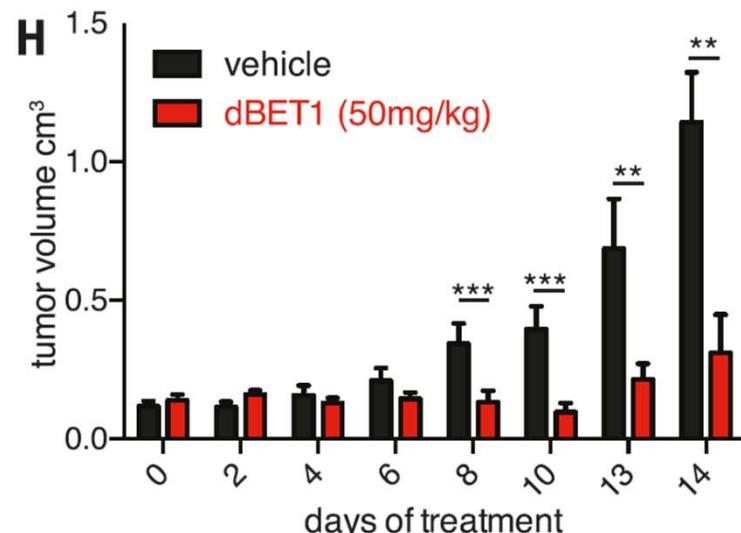


dBET1 treatment results in increased apoptosis with respect to JQ1 in different cell lines and in primary AML blasts

dBET1: anti-proliferative response *in vivo* I

Murine hind-limb xenograft model of MV4;11 AML

dBET (50 mg/kg) or vehicle 1x day i.p. for 14 d

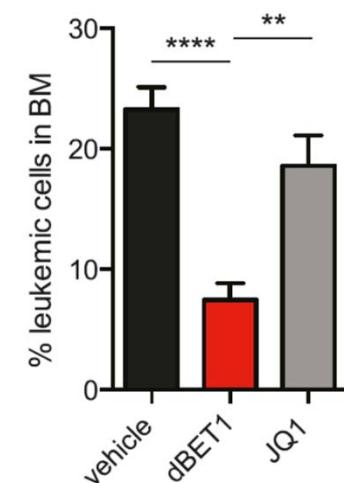


dBET1 treatment results in reduced tumor mass, BRD4/MYC levels and proliferation

Murine disseminated leukemia MV4;11 AML model

dBET or JQ1 (both at 63.8 $\mu\text{mol}/\text{kg}$) or vehicle 1x day i.p. for 19 d

dBET1 and JQ1 treatments result in reduced tumor burden



Catalytic *in vivo* protein knockdown

Advantages:

- Potentially applicable to broad spectrum of endogenous proteins
- Catalytic activity makes PROTACs more potent than the target-binding proteins from which they are derived
- Slower recovery from inhibition, because *de novo* synthesis of the target is required

Disadvantages:

- PROTAC linker length and composition have to be optimized for each target
- Limited to cytosolic proteins (?)
- Possible poor oral bioavailability of PROTACs

Bondeson et al. Nat Chem Biol 2015

Winter et al. Science 2015

Deshaires. Nat Chem Biol 2015

Toure & Crews Angew Chem Int Ed 2015

Ohoka et al. Curr Cancer Drug Targets 2016

