

single-particle cryo-electron microscopy

technical journal club
30.05.2017

Manuela Pfammatter

outline

introduction

single-particle cryo-electron microscopy

Fernandez Leiro & Scheres, Nature, 2016

Frank, Nat Protoc, 2017

rotational states in a V-ATPase

Zhao et al., Nature, 2015

spiral architecture of Hsp104 disaggregase

Yokom et al., Nat Struct Mol Biol, 2016

conclusion & outlook

why structure determination?



protein function



interactions



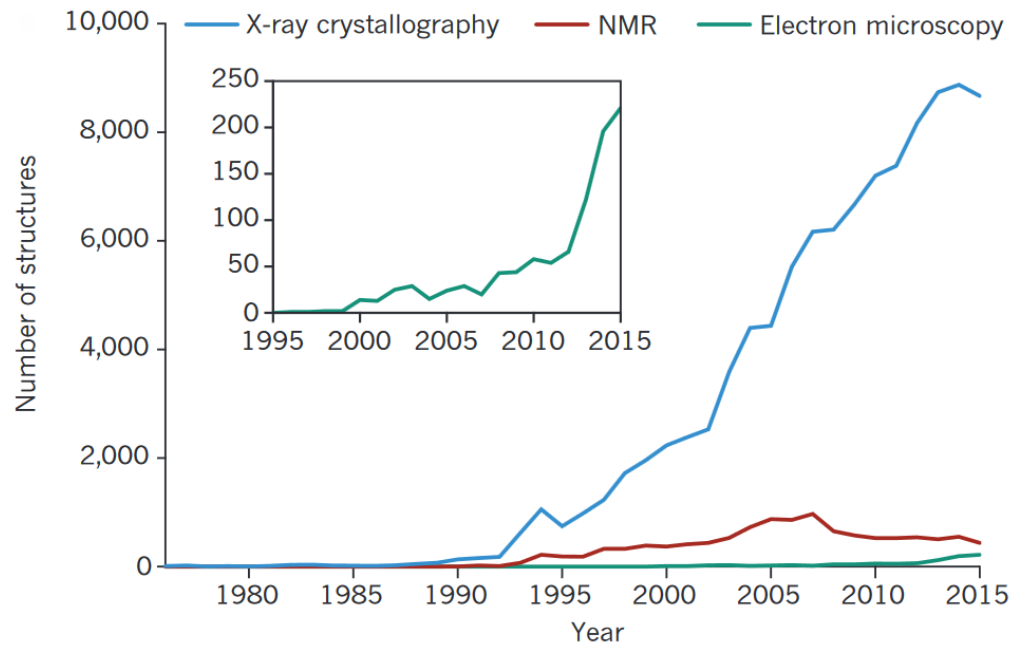
dynamics



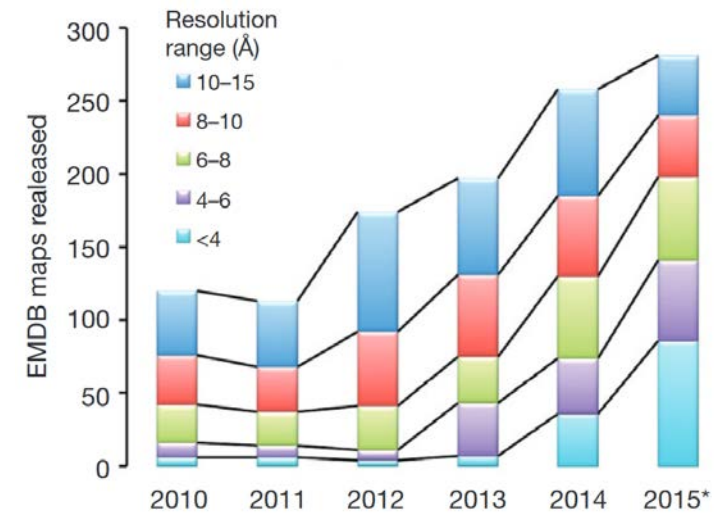
structure

elucidate **structures** to understand the **mechanisms** underlying **biological function**

methods for structure determination

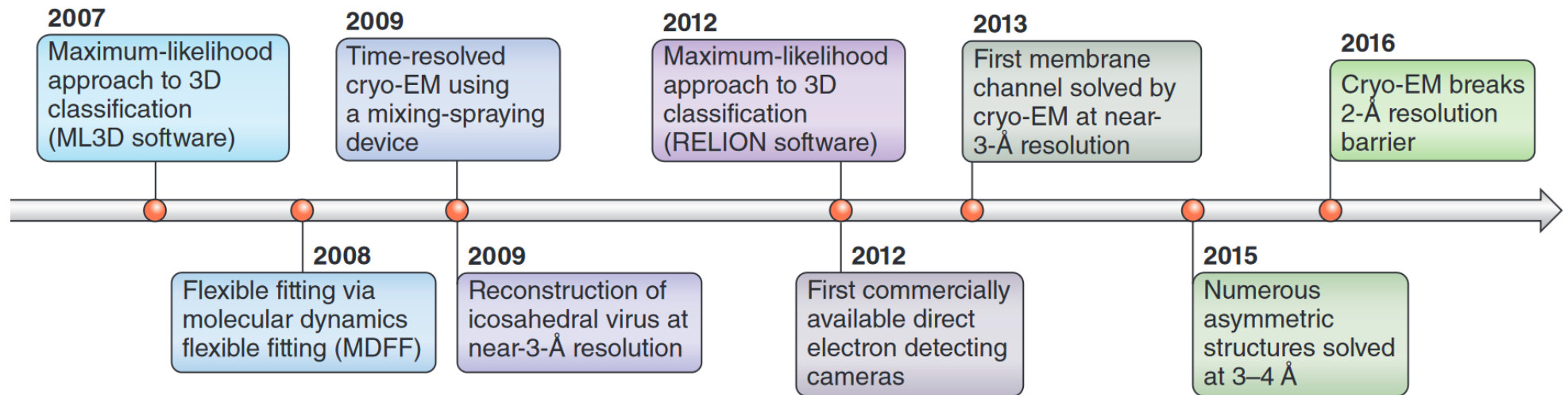


Fernandez-Leiro, 2016



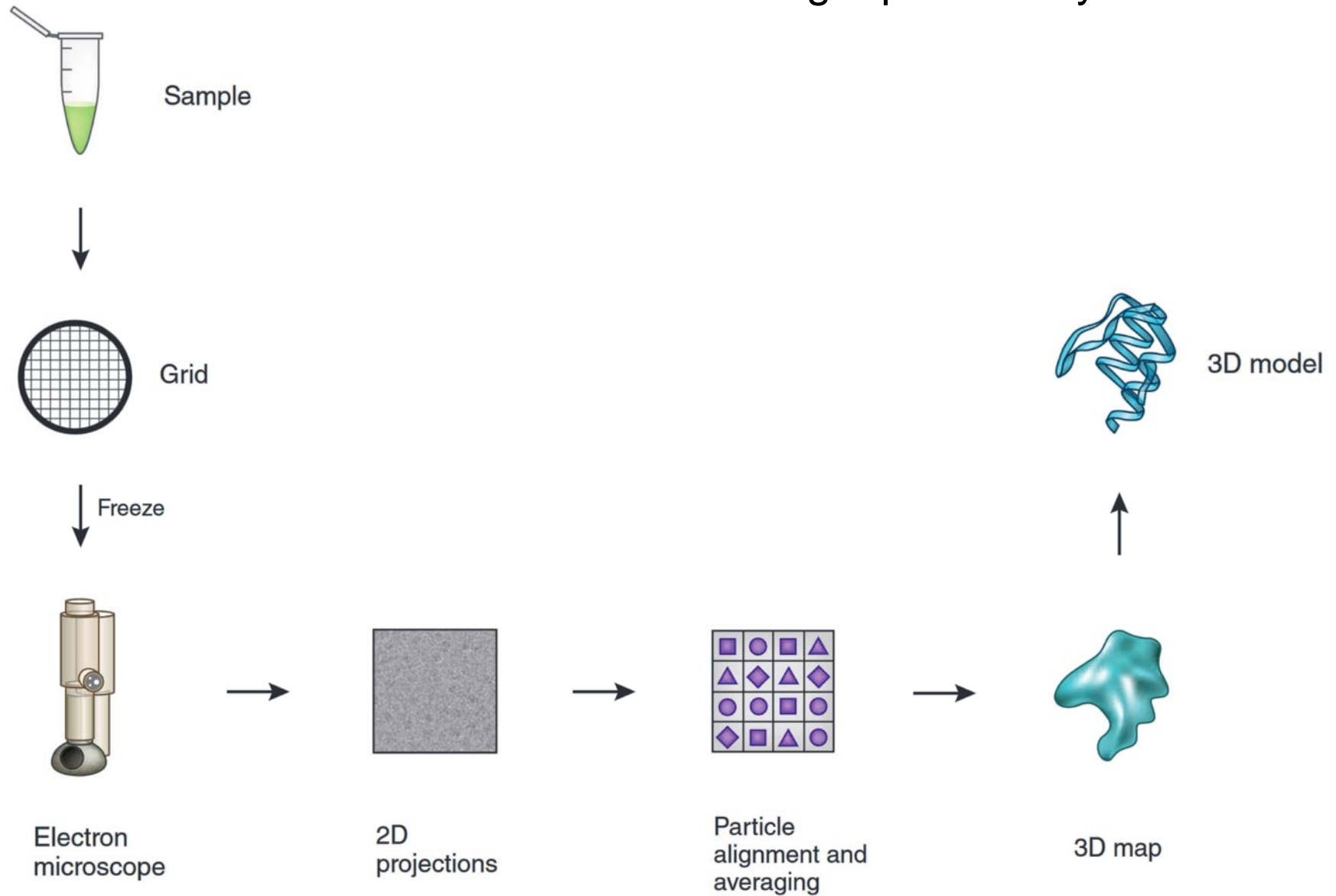
Nogales, 2016

recent advances in cryo-EM

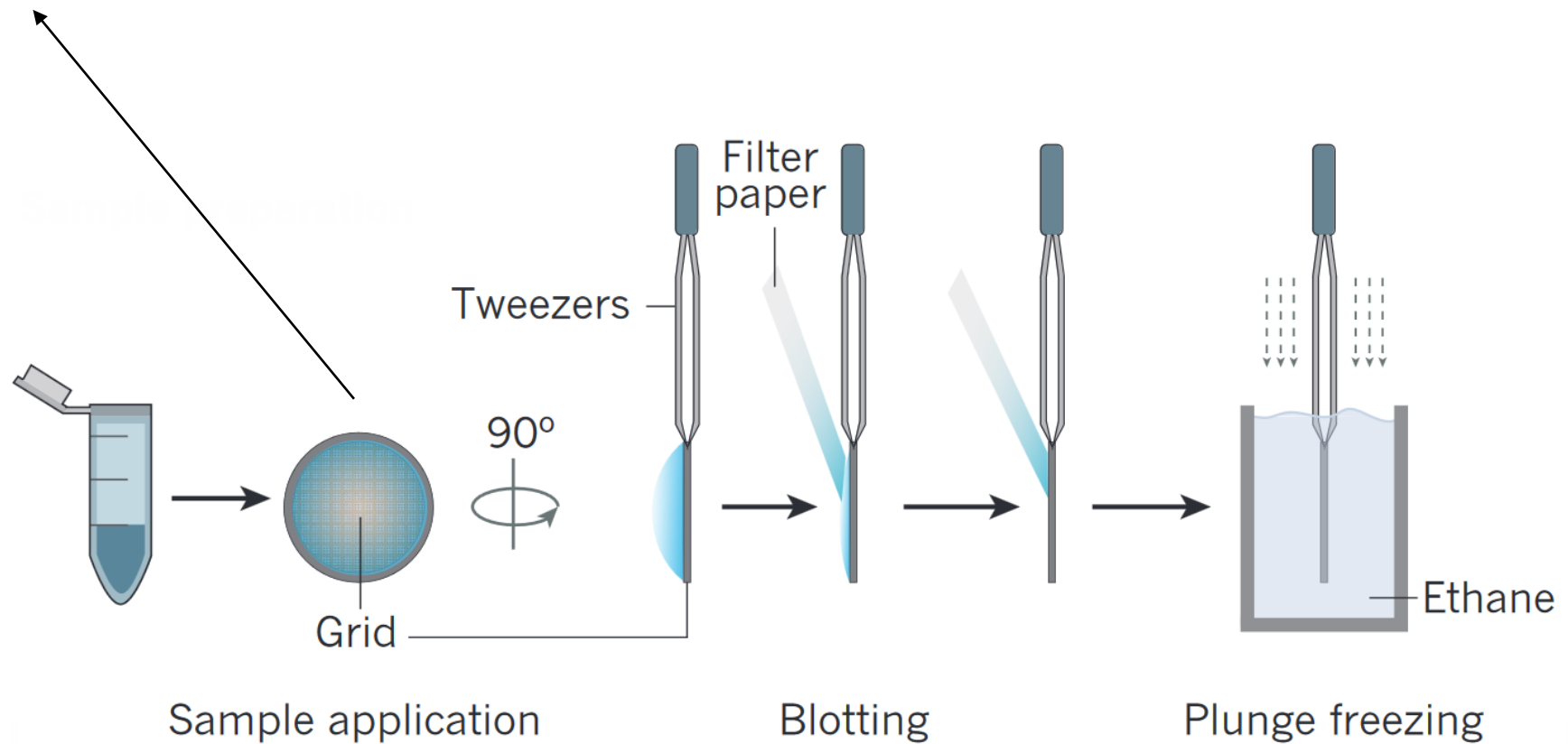
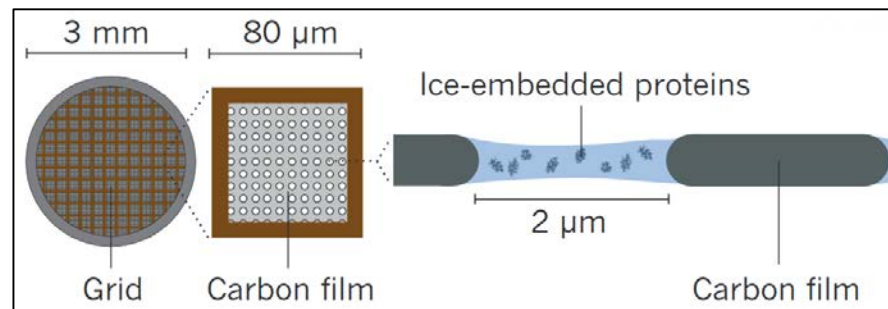


Frank, 2017

single-particle cryo-EM workflow

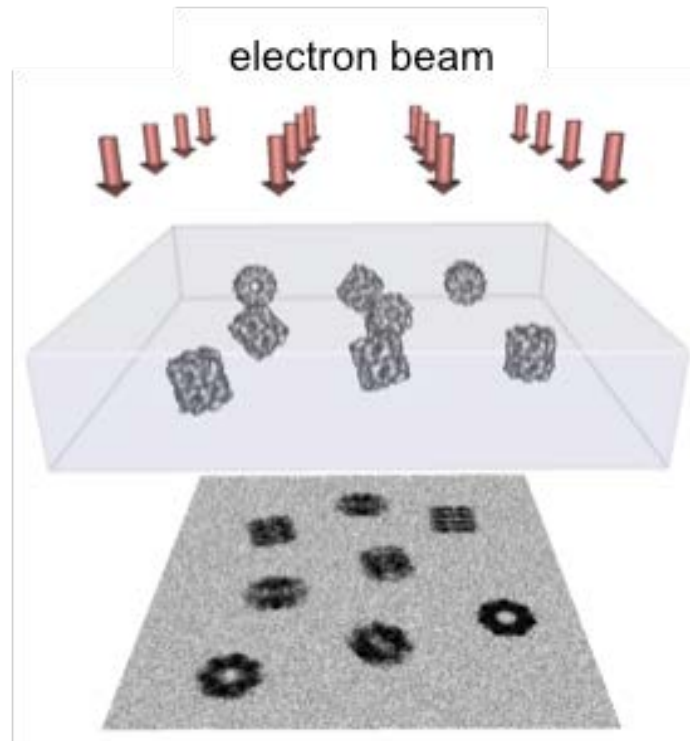


i. sample grid preparation

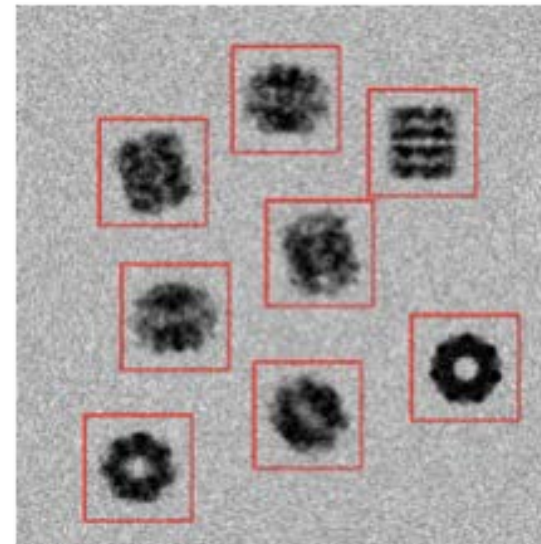


adapted from *Fernandez-Leiro, 2016*

ii. data collection: 2D projections

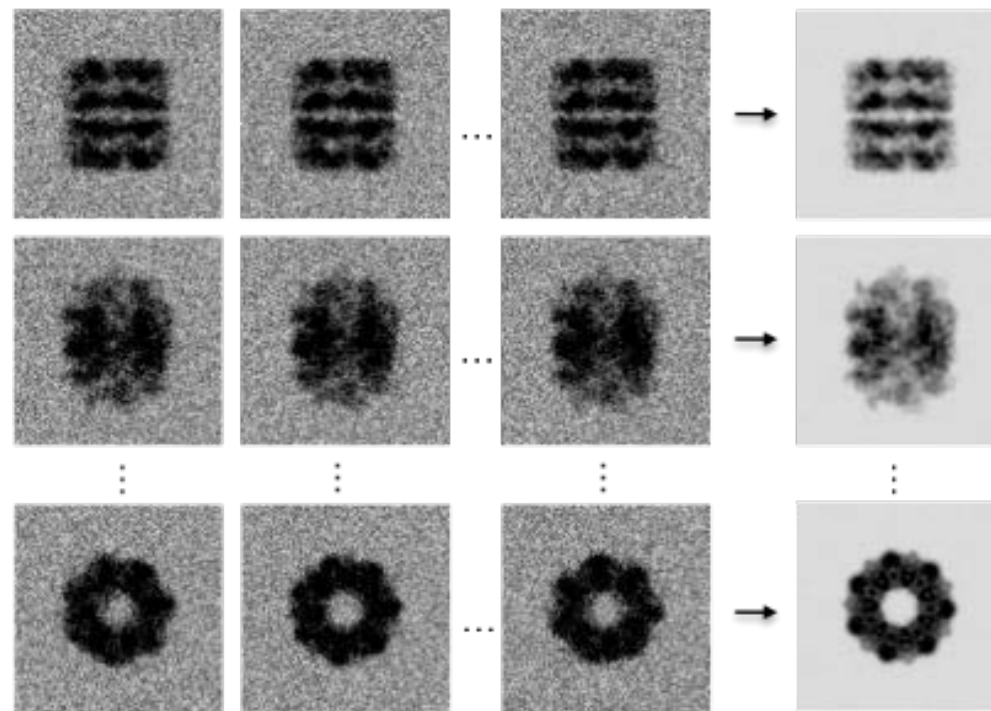


2D projections



particle boxing

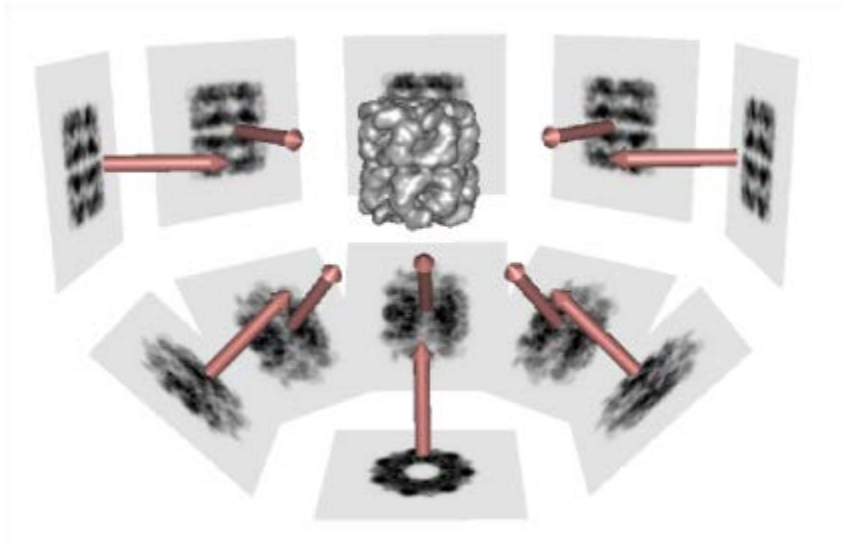
iii. data processing: particle alignment and averaging



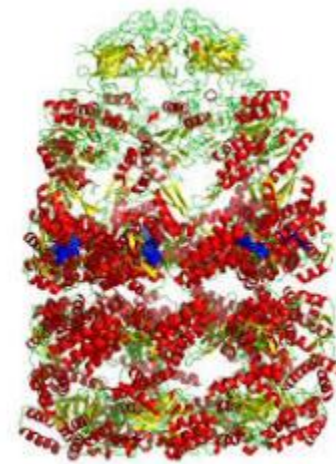
clustering

averaging

iv. data processing: 3D reconstruction



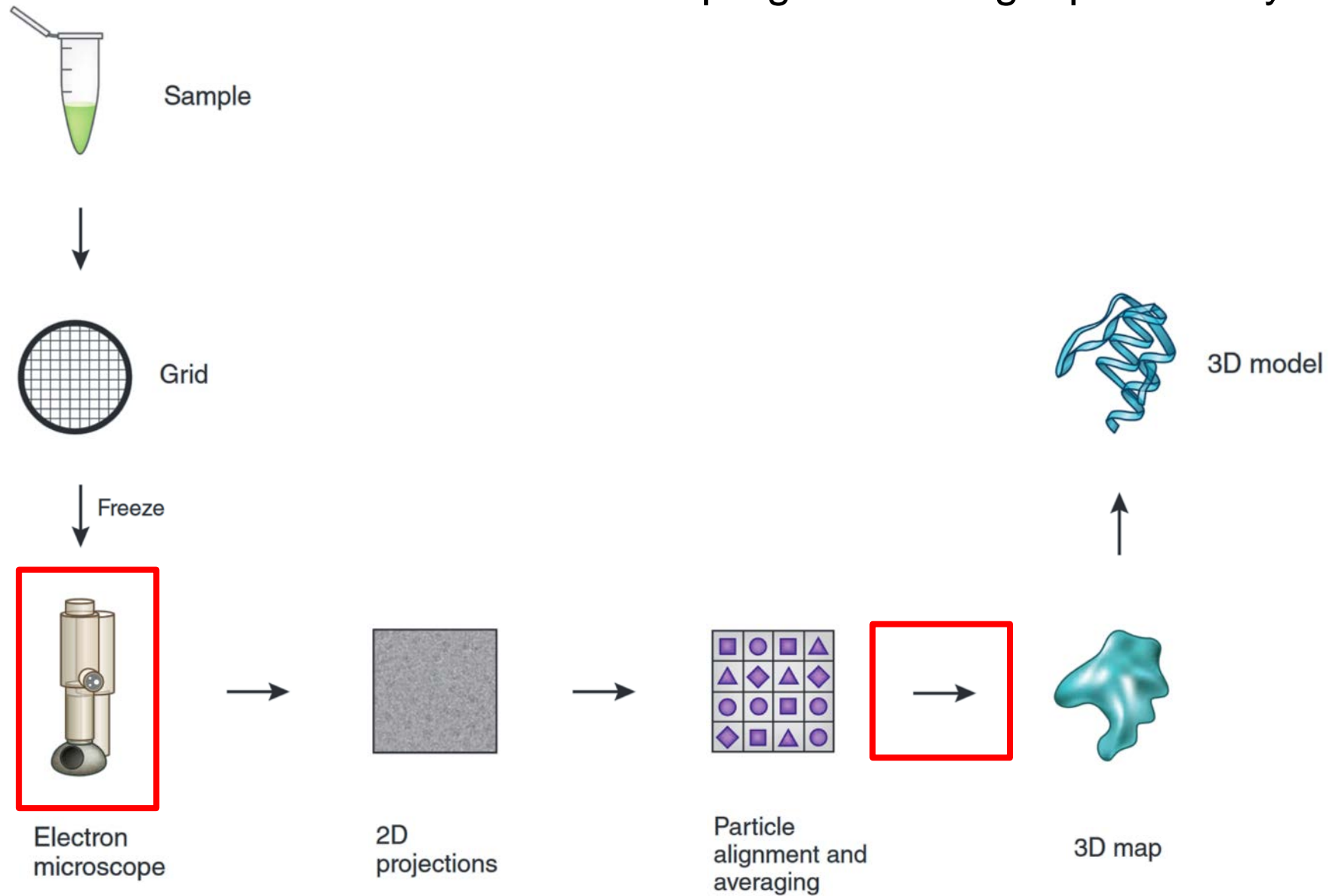
3D map



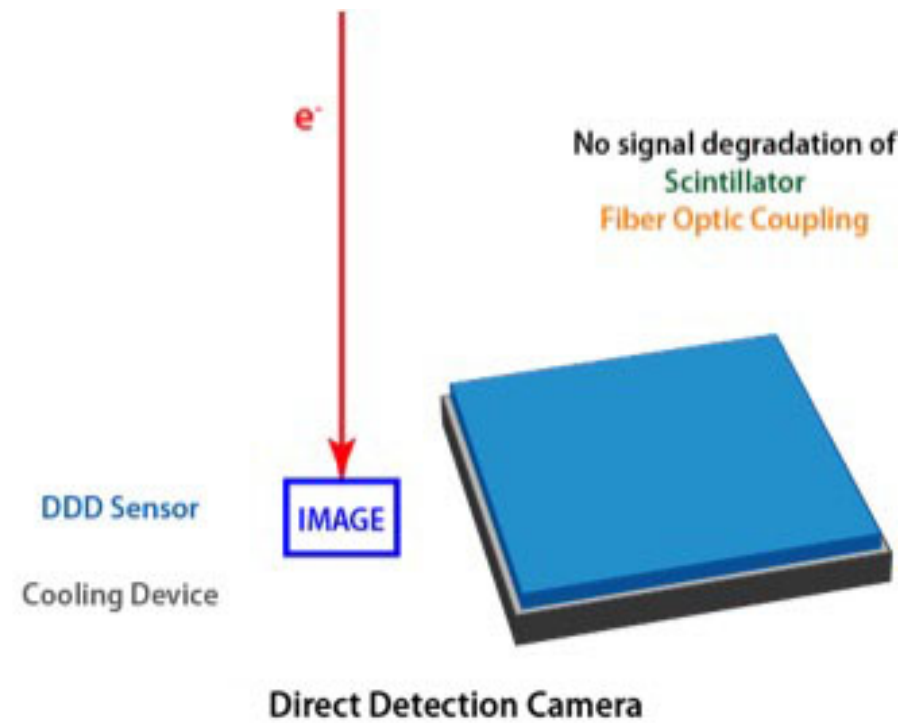
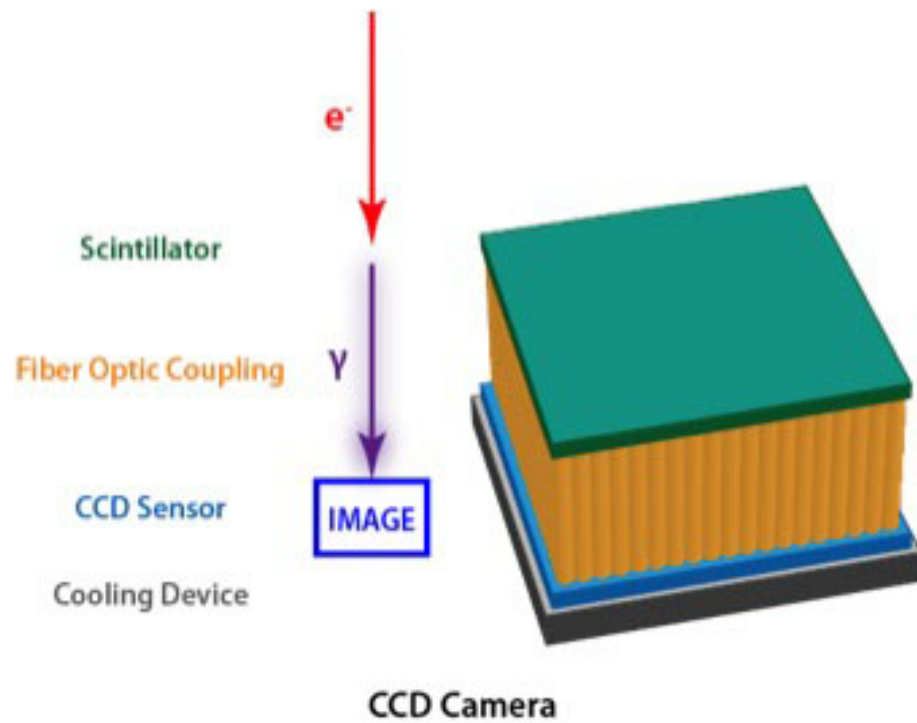
3D model

adapted from *Pintilie (online)*

recent progress in single-particle cryo-EM



principle of direct detection



courtesy of Direct Electron

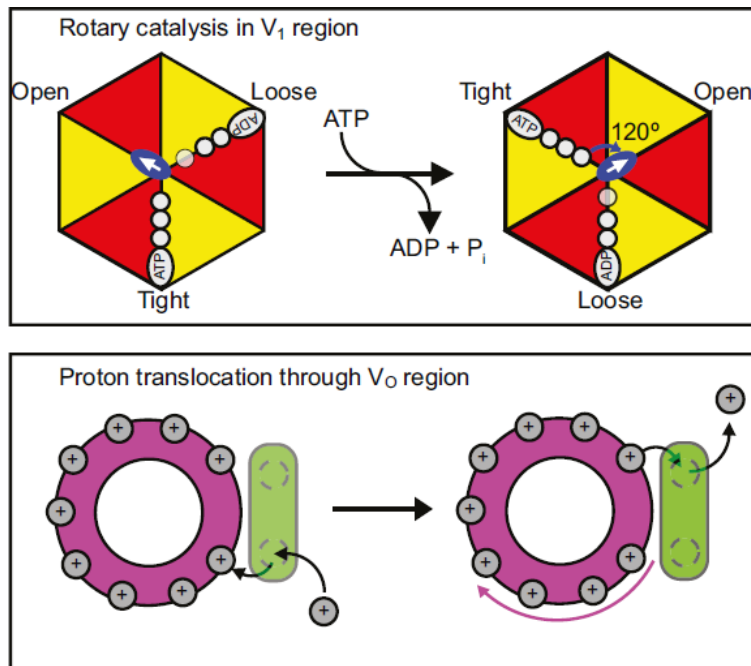
nature

Electron cryomicroscopy observation of rotational states in a eukaryotic V-ATPase

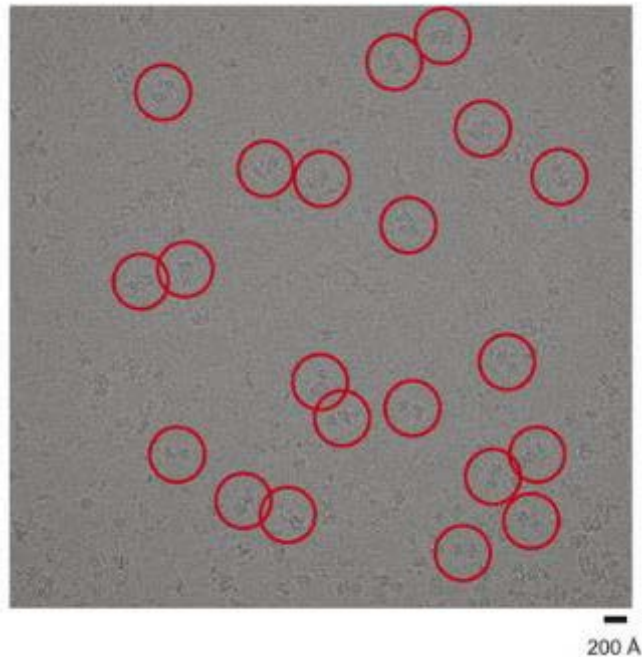
Jianhua Zhao^{1,2*}, Samir Benlekbir^{1*} & John L. Rubinstein^{1,2,3}

eukaryotic V-ATPase

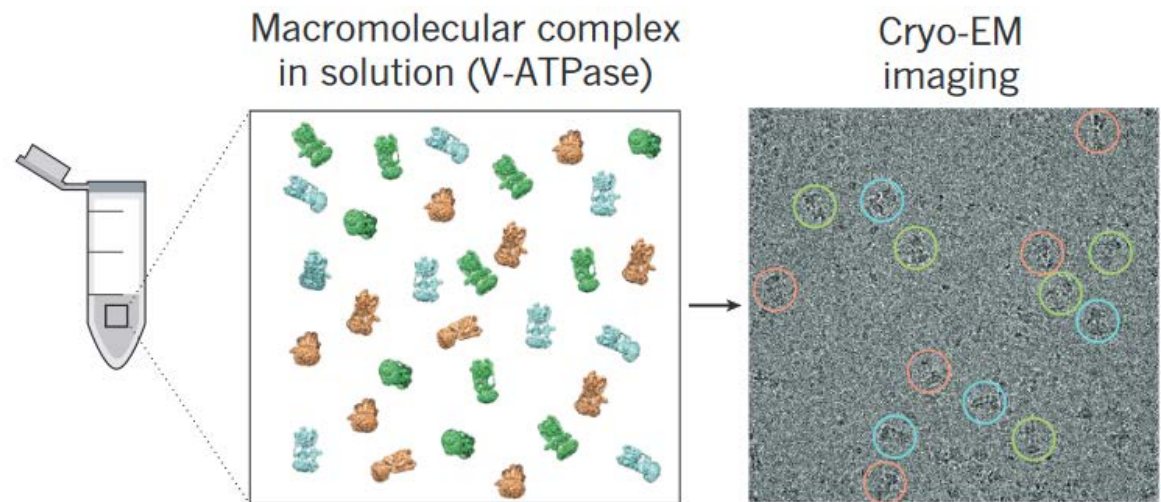
eukaryotic vacuolar H⁺-ATPase



V-ATPase: data collection and processing

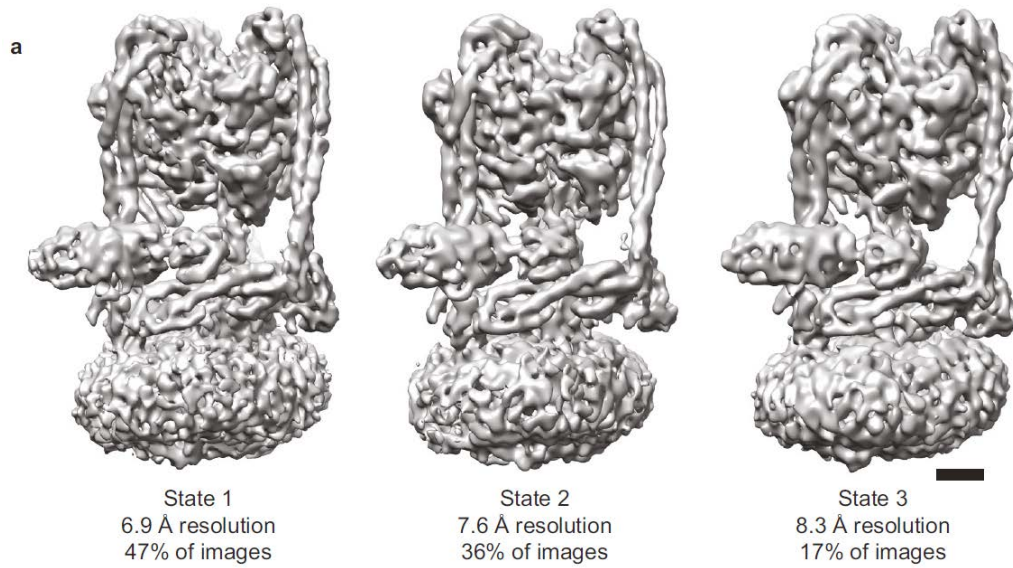


2D projection

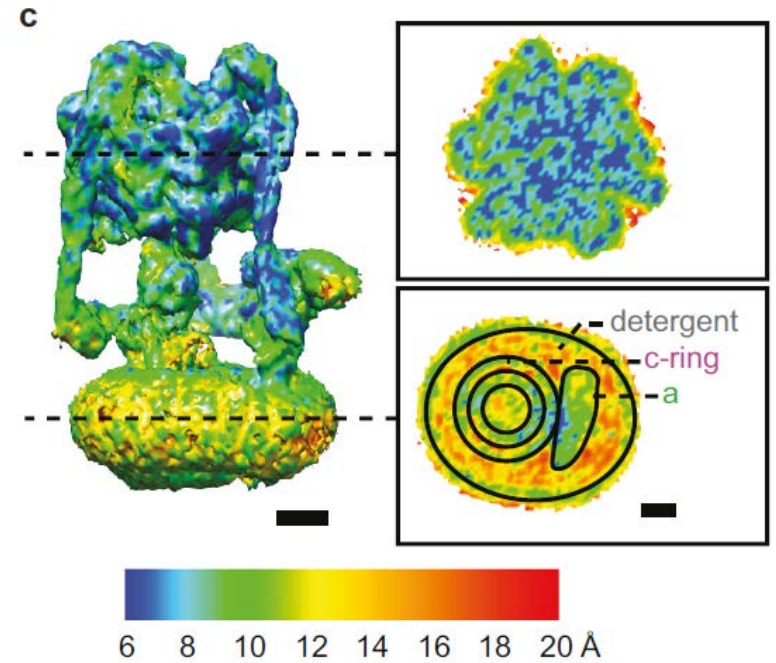


particle boxing and classification
(106 445 particles)

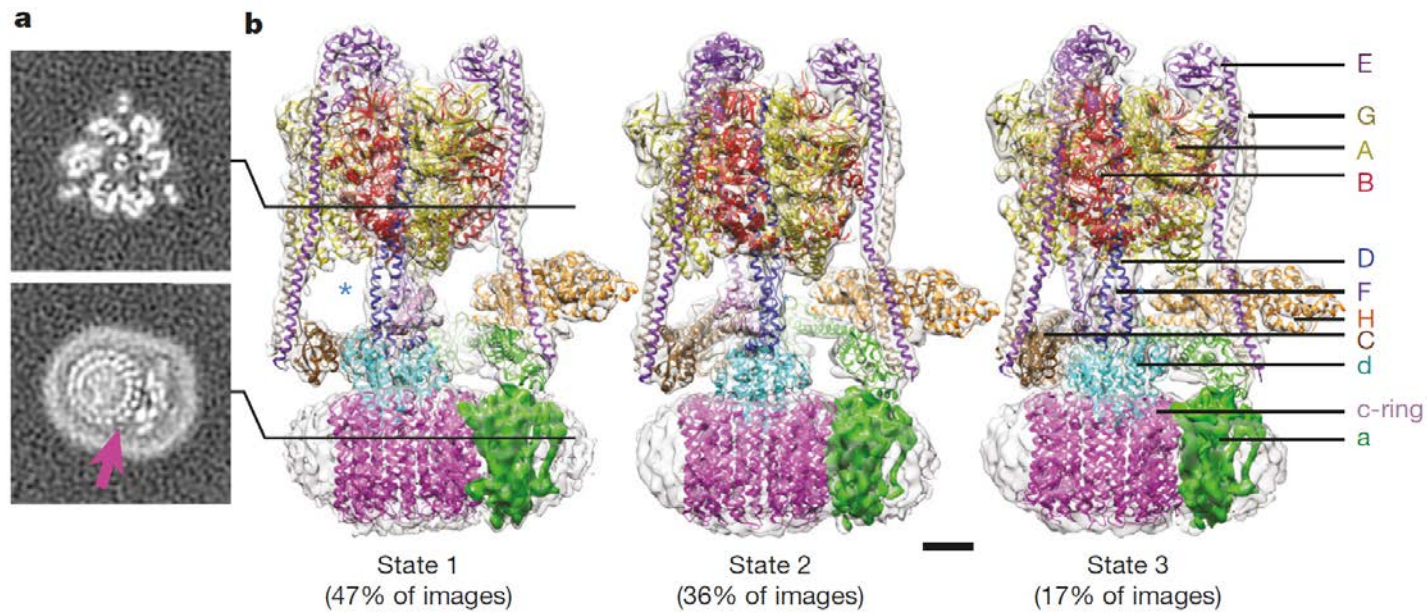
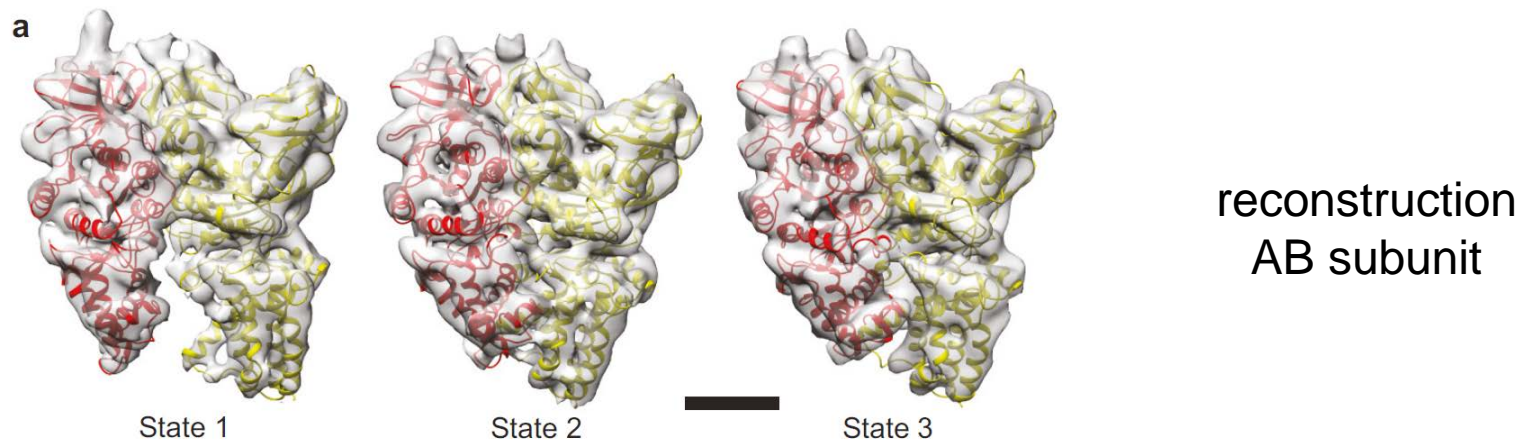
V-ATPase: data processing



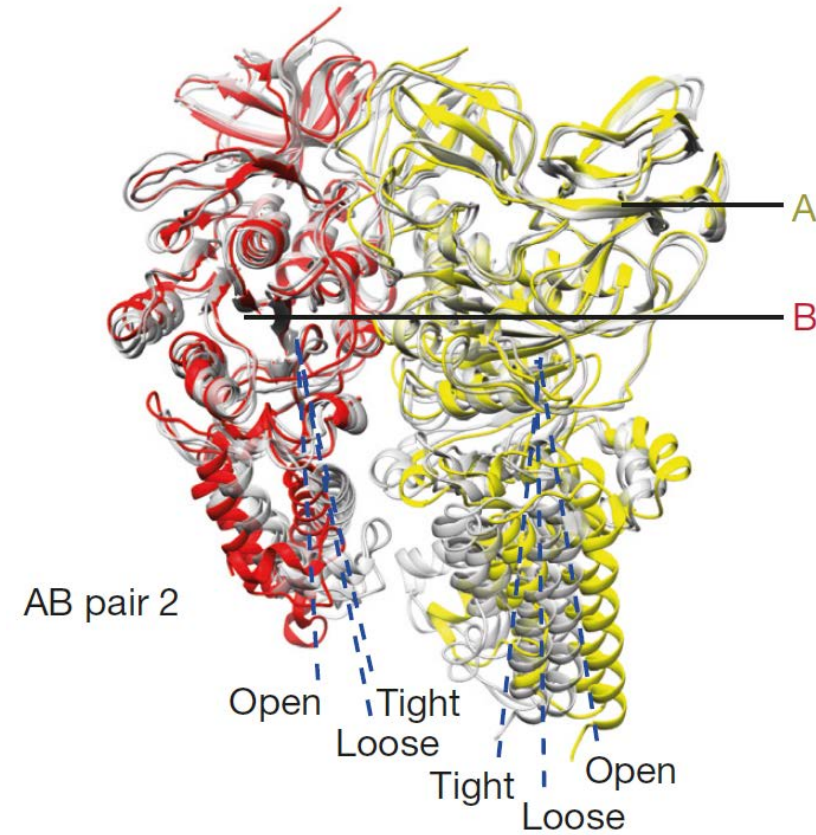
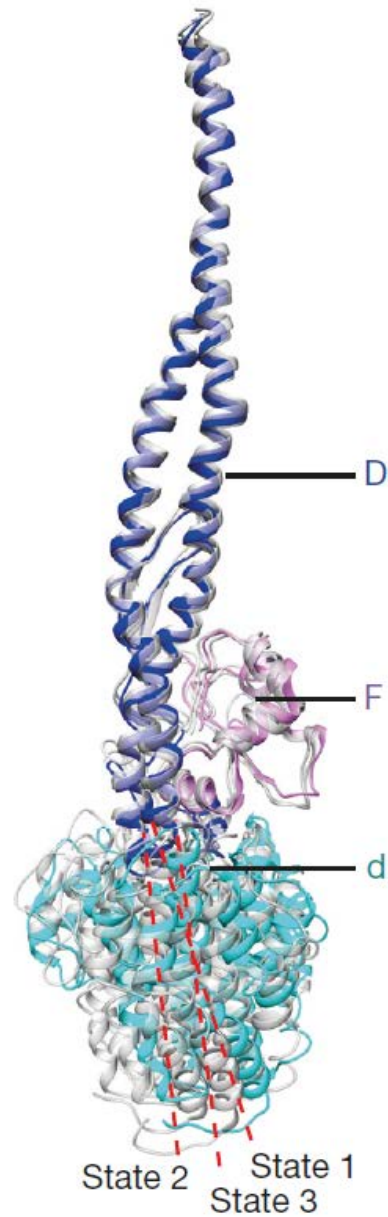
3D maps



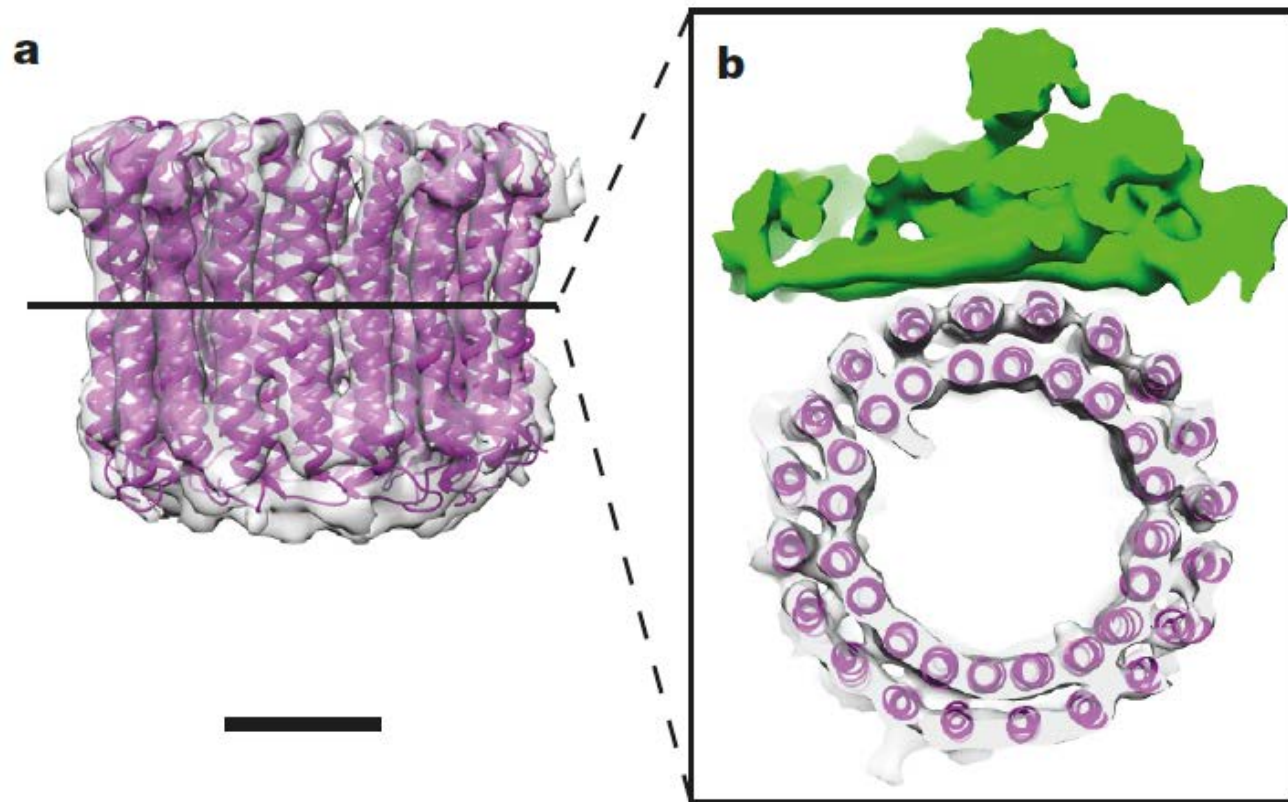
V-ATPase: data processing



central rotor and soluble V_1 region

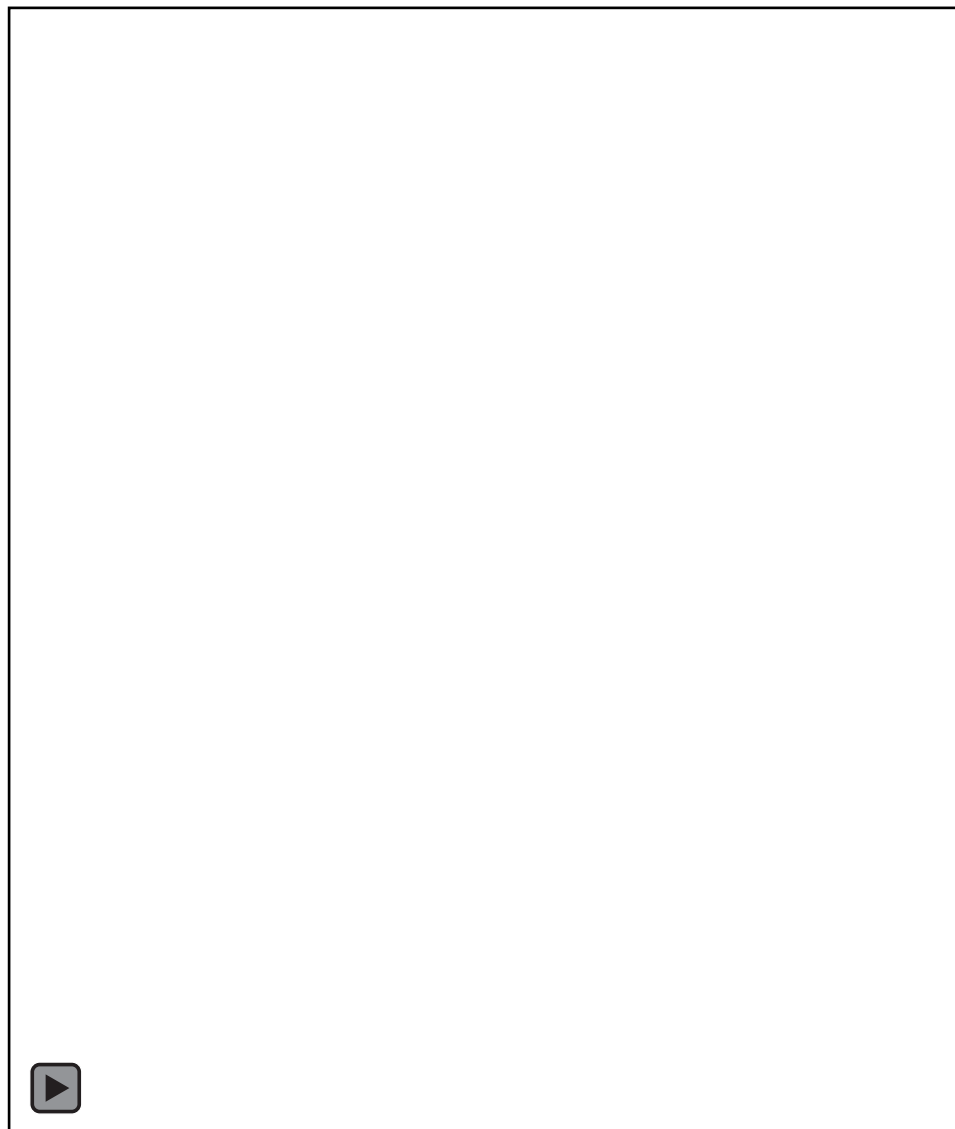


membrane-bound V_0 region



ATP:H⁺ = 3:10

structural changes occurring during rotary catalysis



nature
structural &
molecular biology

Spiral architecture of the Hsp104 disaggregase reveals the basis for polypeptide translocation

Adam L Yokom^{1,2}, Stephanie N Gates^{1,2}, Meredith E Jackrel³, Korrie L Mack^{3,4}, Min Su¹, James Shorter^{3,4} & Daniel R Southworth¹

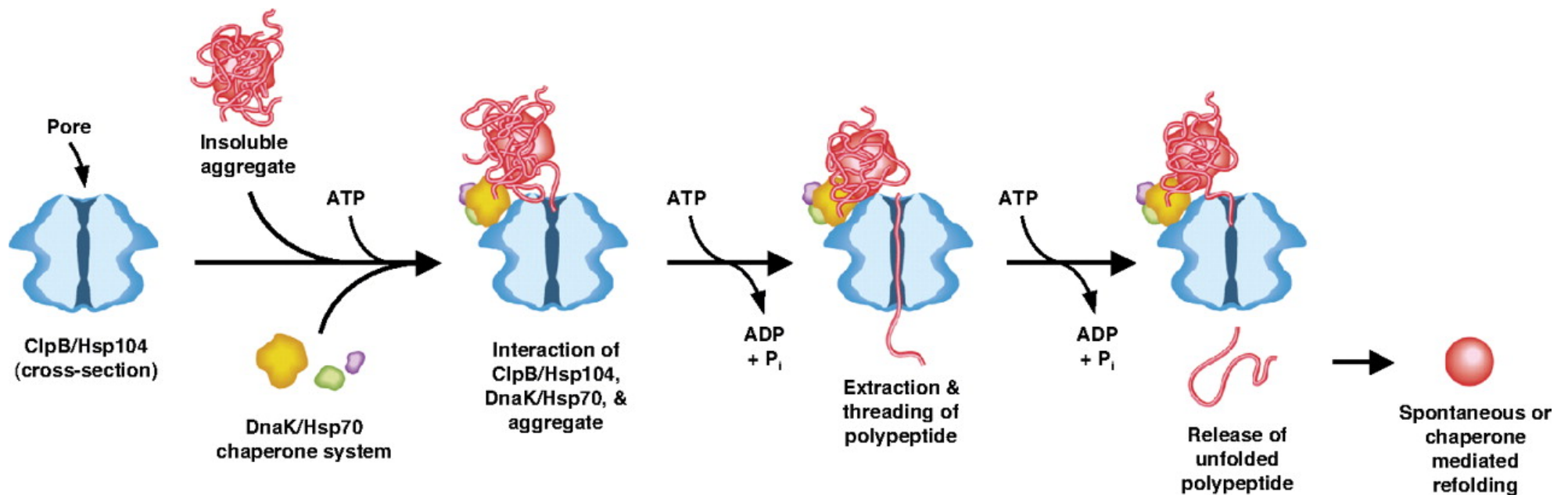
Hsp104 disaggregase

molecular chaperone, heat-shock protein

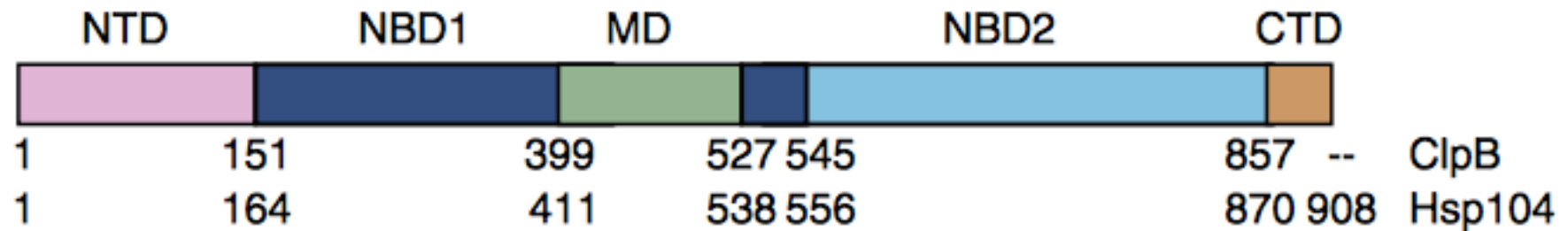
cooperation with Hsp70 in unfolding and rescuing aggregated protein

-> active translocation of polypeptide substrates through central channel

dose-dependent effects on yeast prion propagation

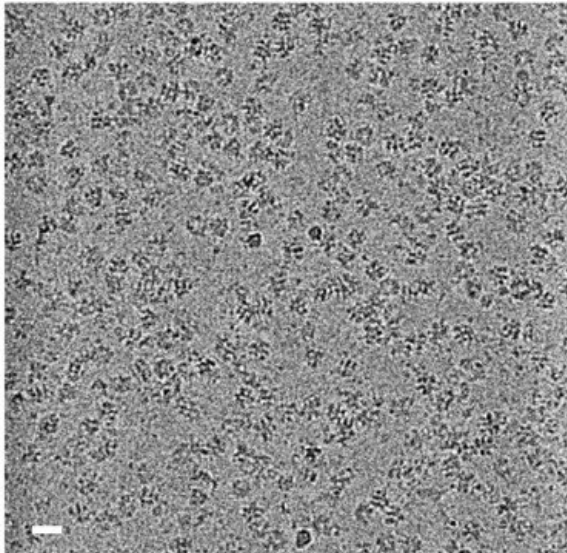


Hsp104 domain arrangement

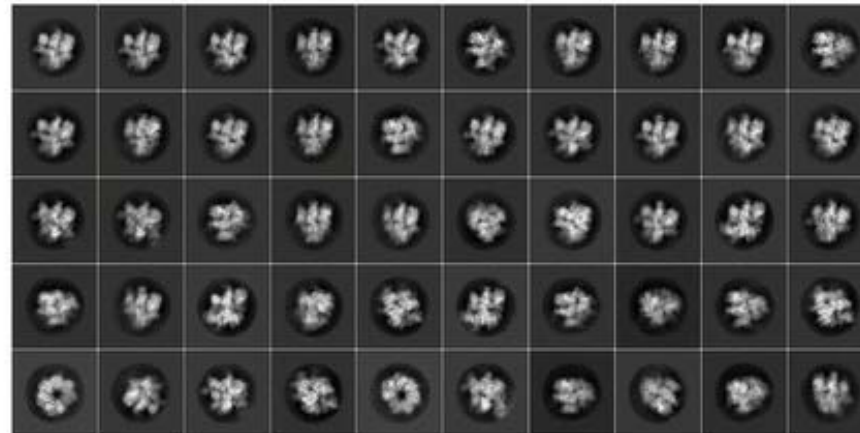


NTD	N-terminal domain	substrate engagement
NBD	nucleotide-binding domain	ATPase-binding domain, power translocation
MD	middle domain	disaggregation, interaction with Hsp70
CTD	C-terminal domain	required for hexamerisation

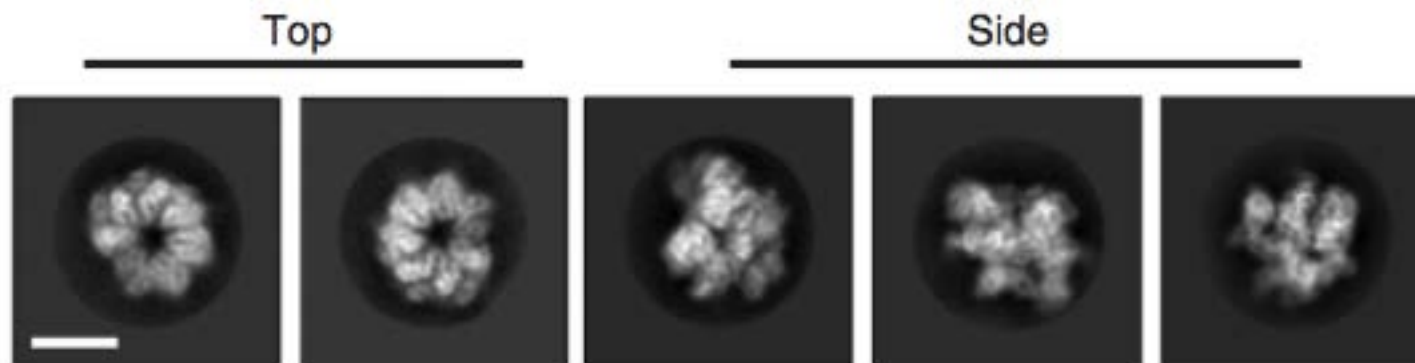
Hsp104: data collection and processing



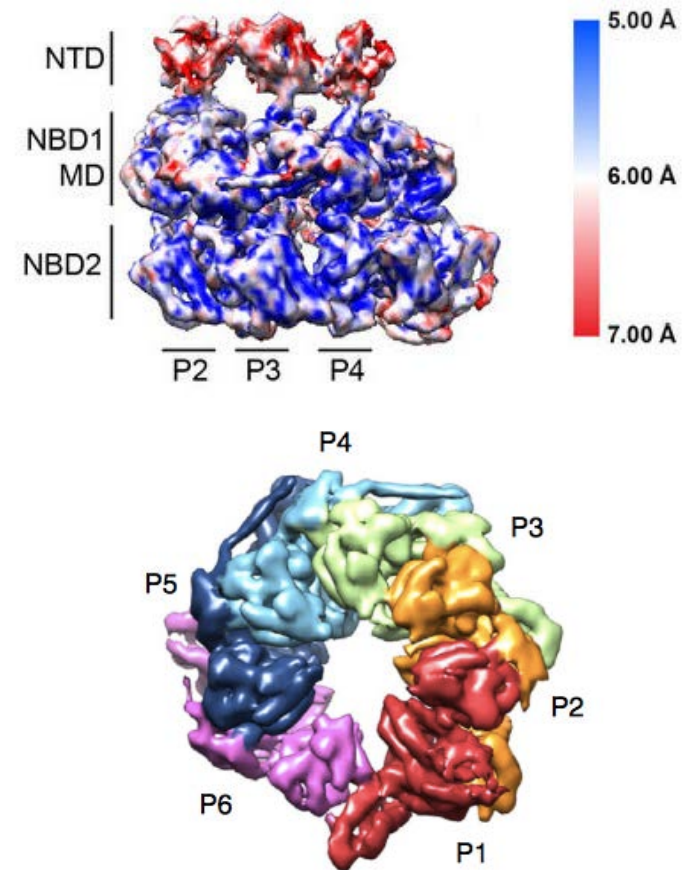
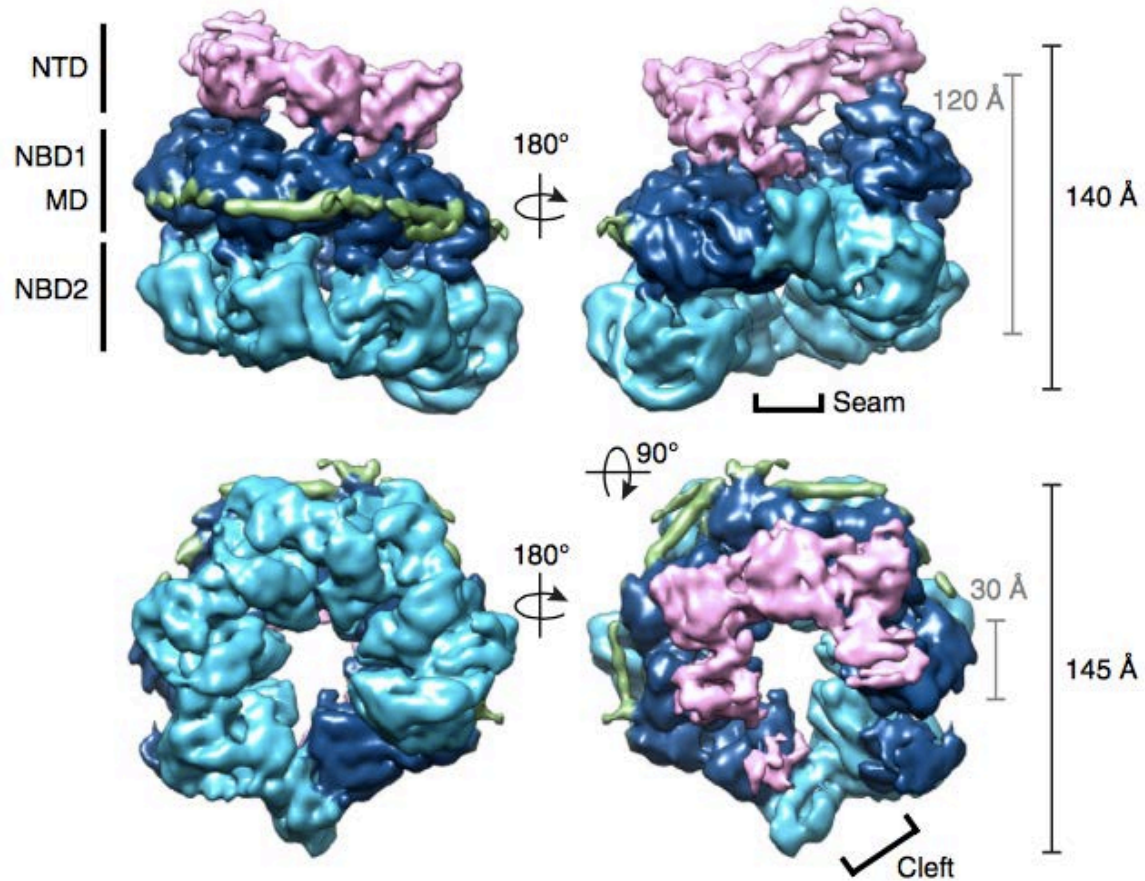
2D projection



classified and averaged particles

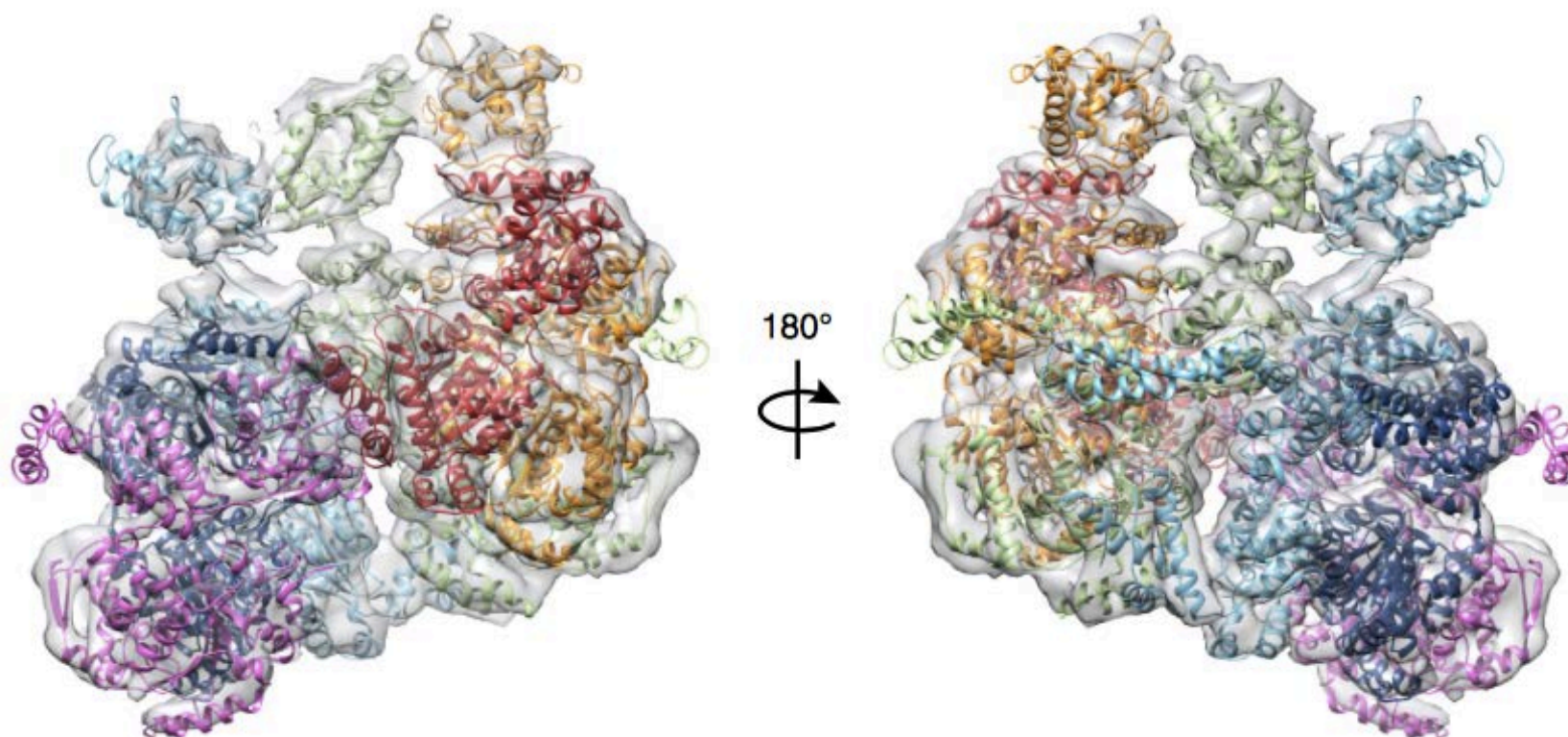


Hsp104: data processing

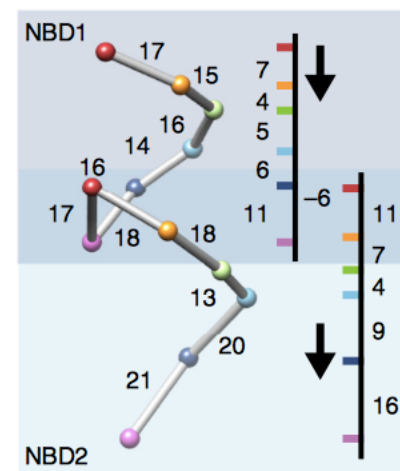
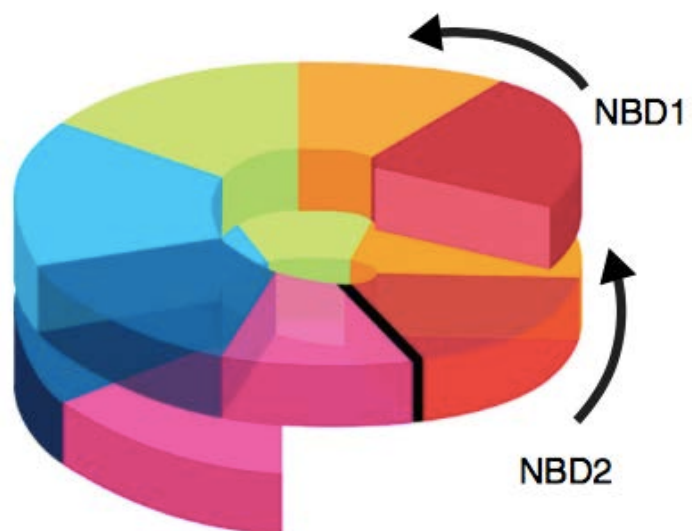
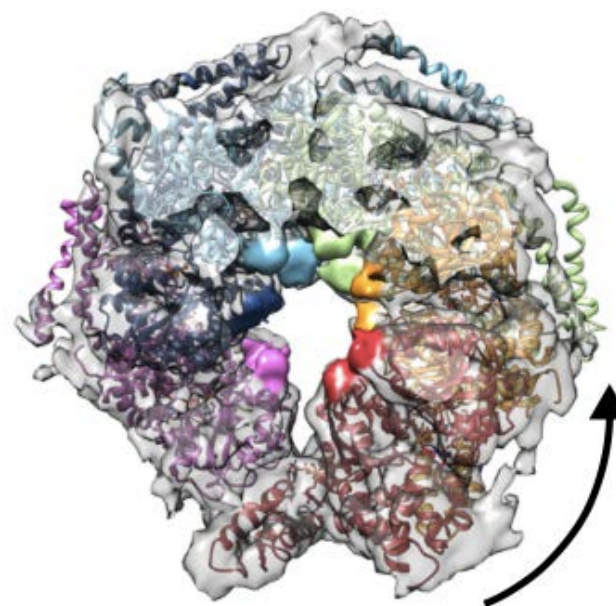
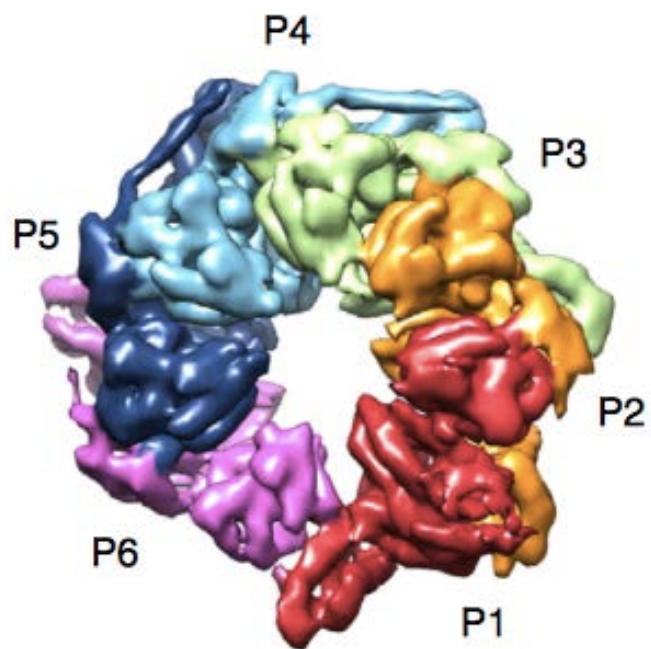


3D maps

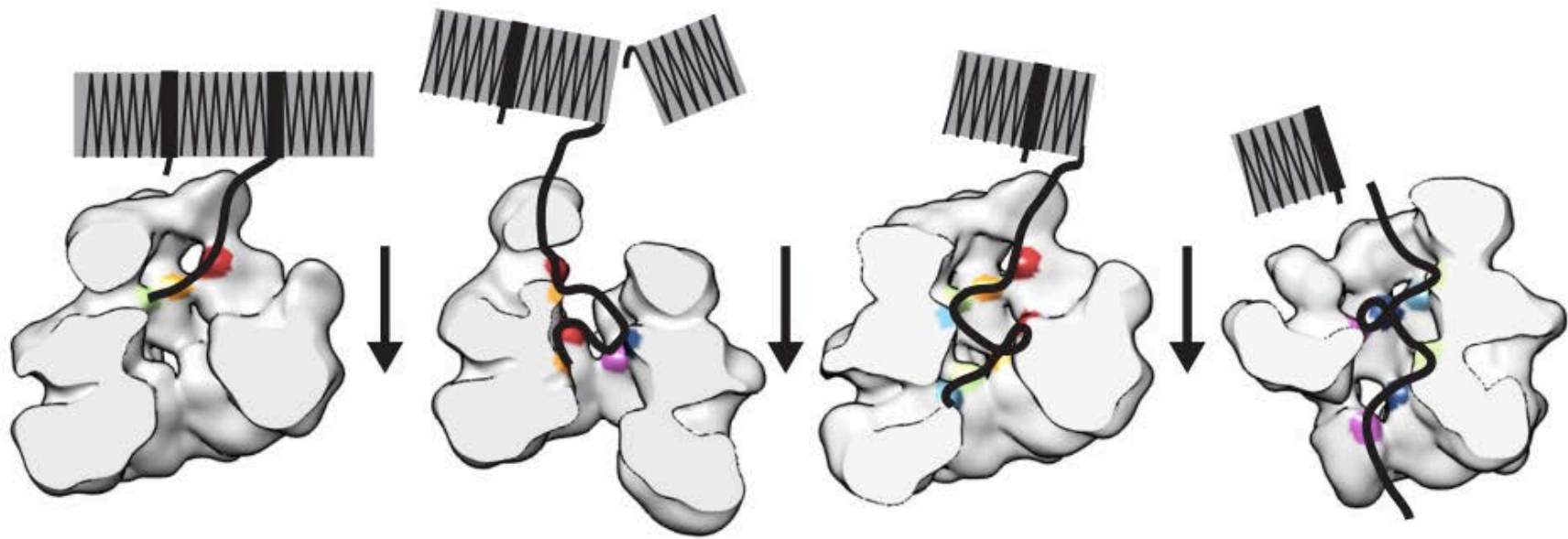
Hsp104: data processing



spiral architecture



mechanism of cooperative disaggregation



cooperative disaggregation

conclusions and outlook

- + near-atomic resolution data
- + native and hydrated structure (no contact surfaces)
- + capturing of different functional states / conformational transitions
- + flexible regions do not impede structure determination

- low signal-to-noise ratio -> many images required
- time consuming and delicate sample preparation
- operation in a high vacuum
- large, expensive facilities

outlook

optimisation and automation of sample preparation

further improvements of direct electron detectors

software development

time-resolved studies

