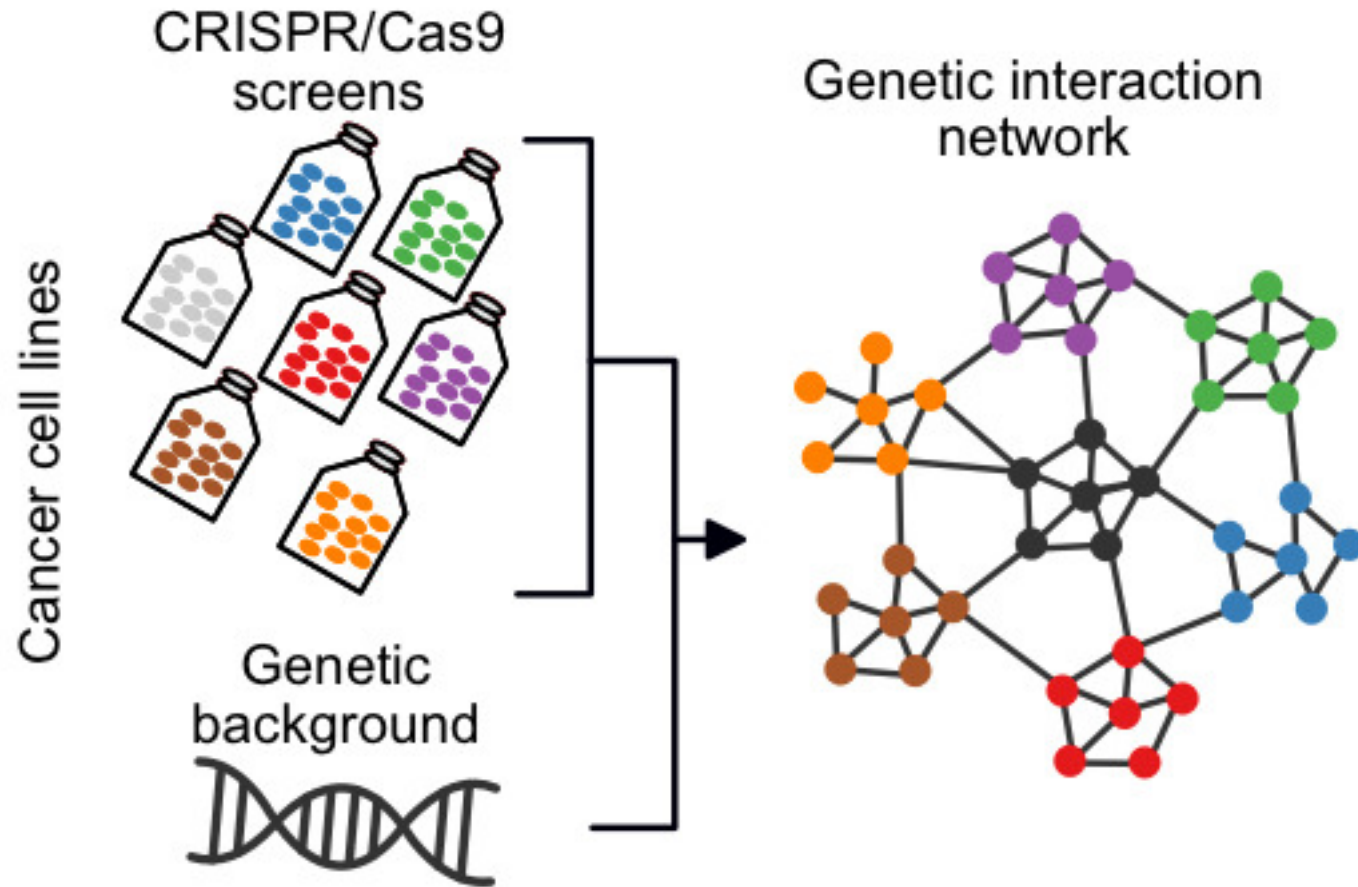


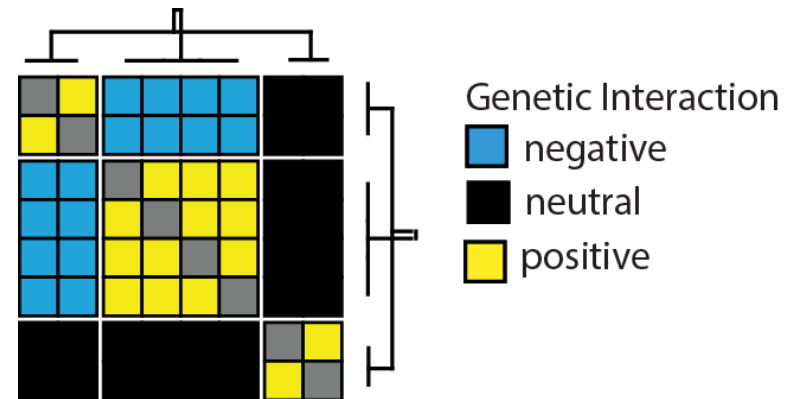
MINGLE



Asvin Lakkaraju
05.10.2020

Genetic Interaction maps

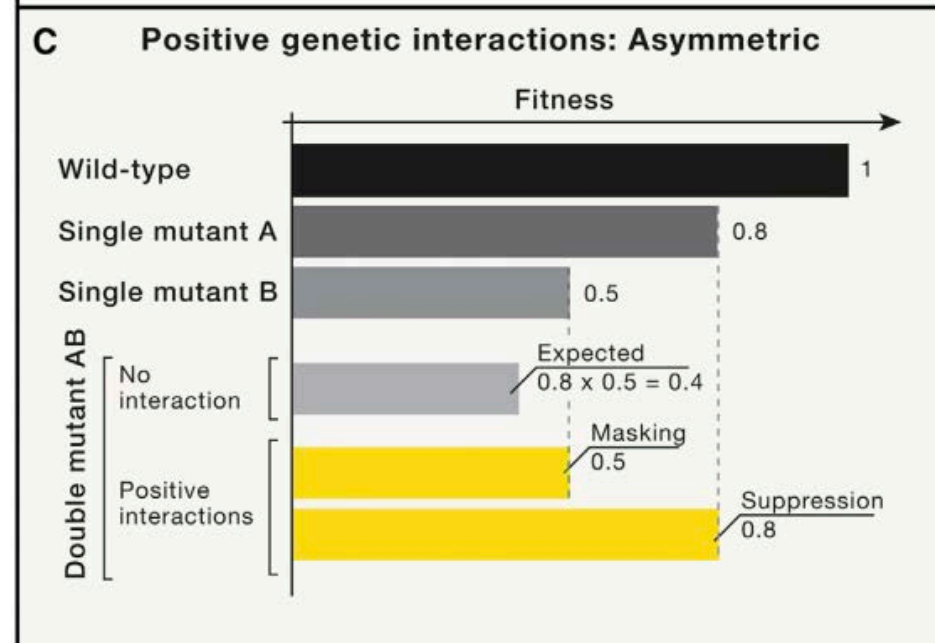
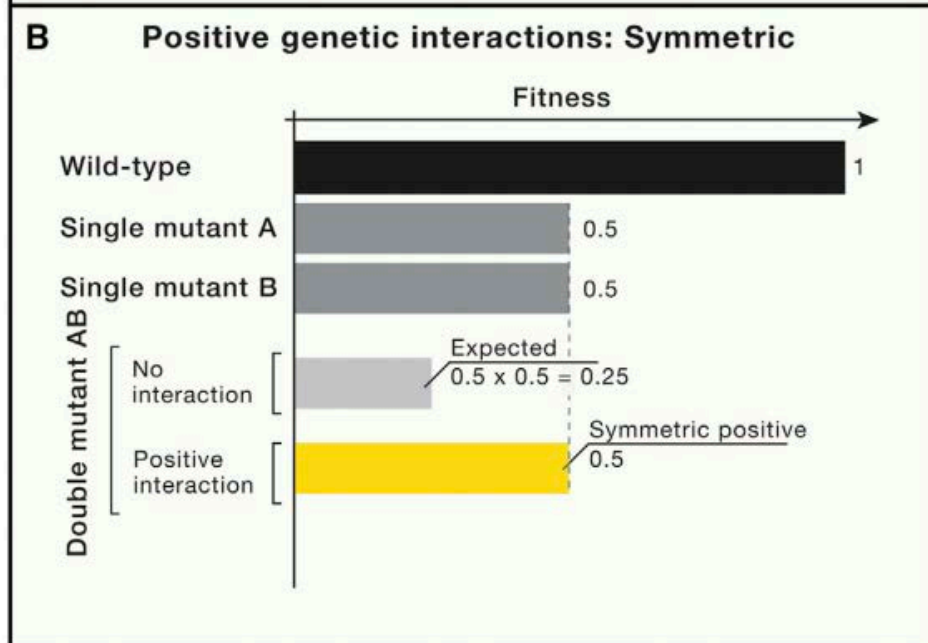
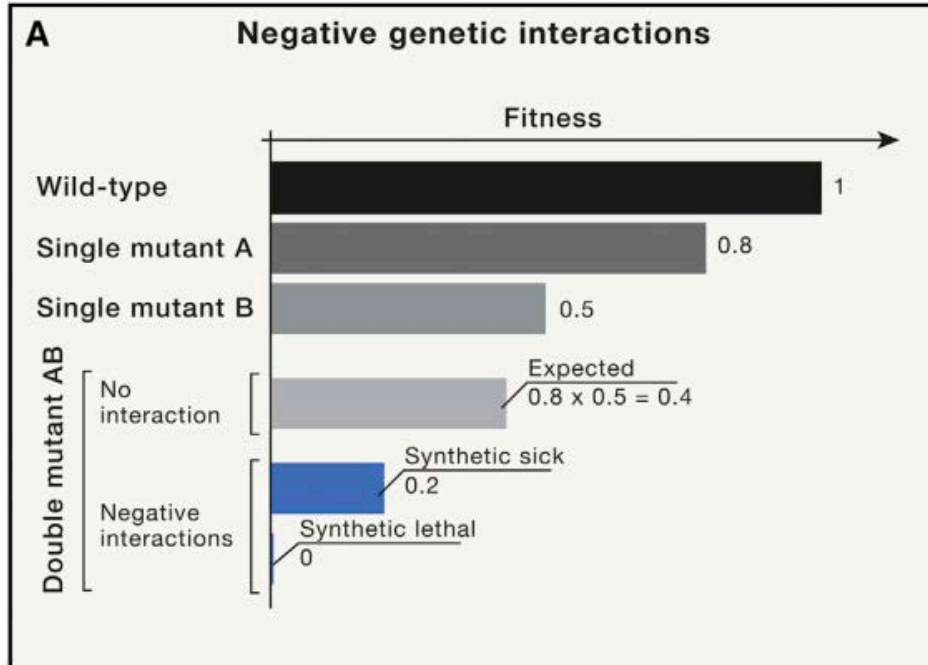
Genetic interaction (GI) mapping, pioneered in the early 2000s, is a powerful technique to systematically reveal functional relationships between genes, which often also reveal the presence of a physical interaction.



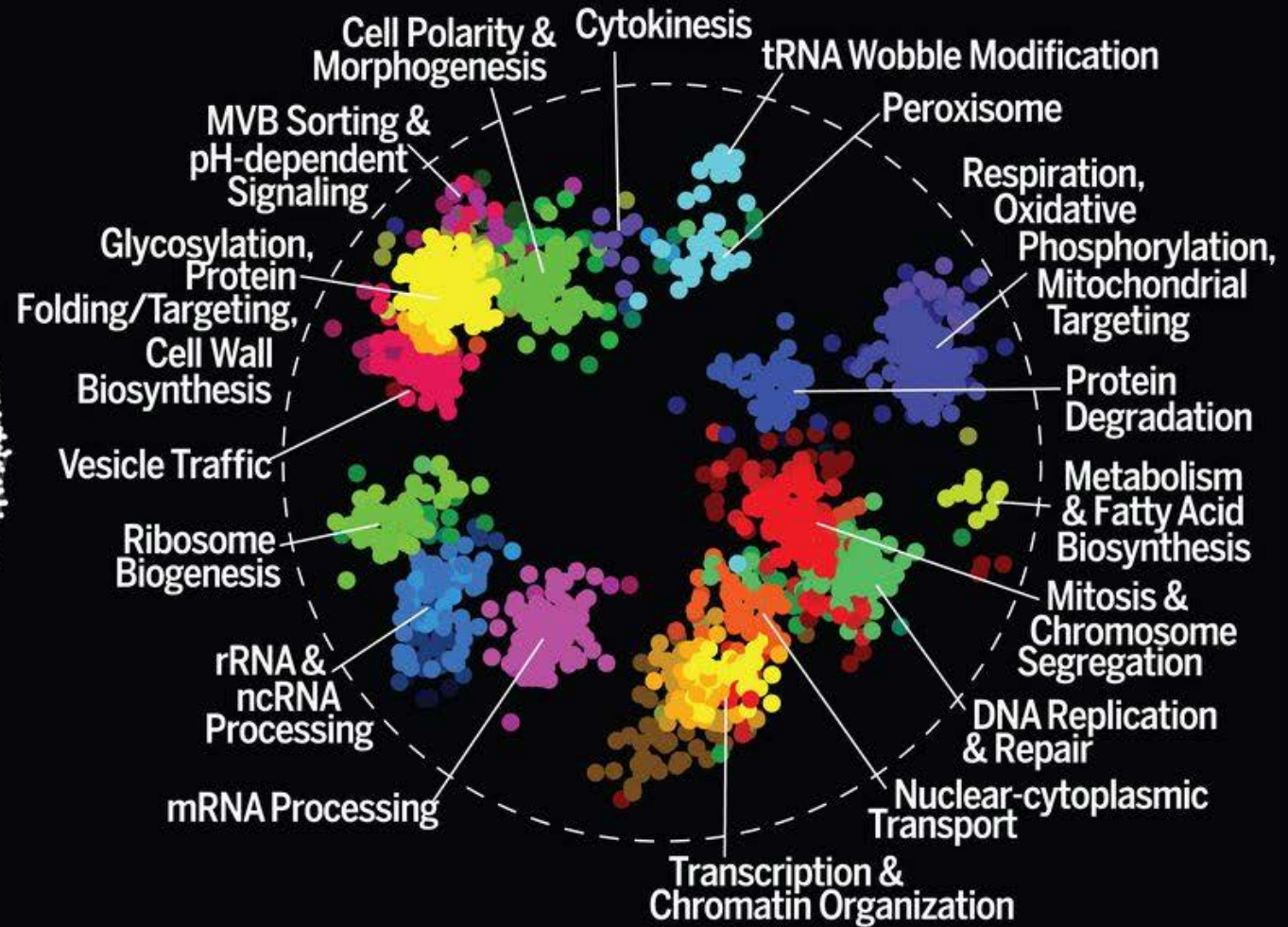
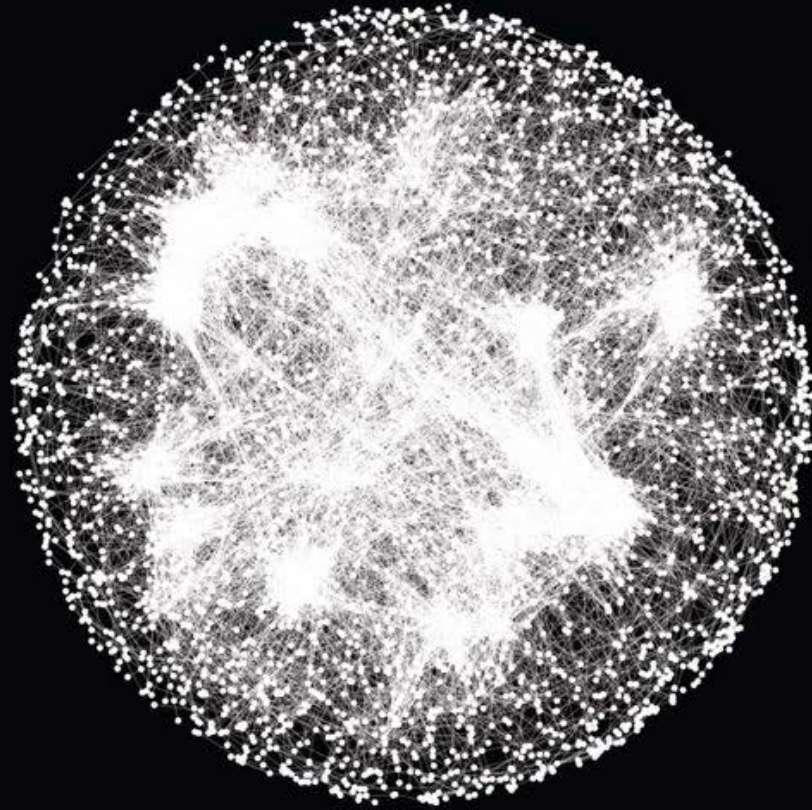
GI mapping involves the pairwise perturbation of genes (e.g. knockout, knockdown or overexpression) in order to elucidate how one gene modulates the phenotype of the other.

Typically, cell viability is used as the phenotypic readout, where GIs that increase cellular fitness are said to be “positive” and GIs that decrease cellular fitness are said to be “negative”.

Genetic Interaction maps: At a glance

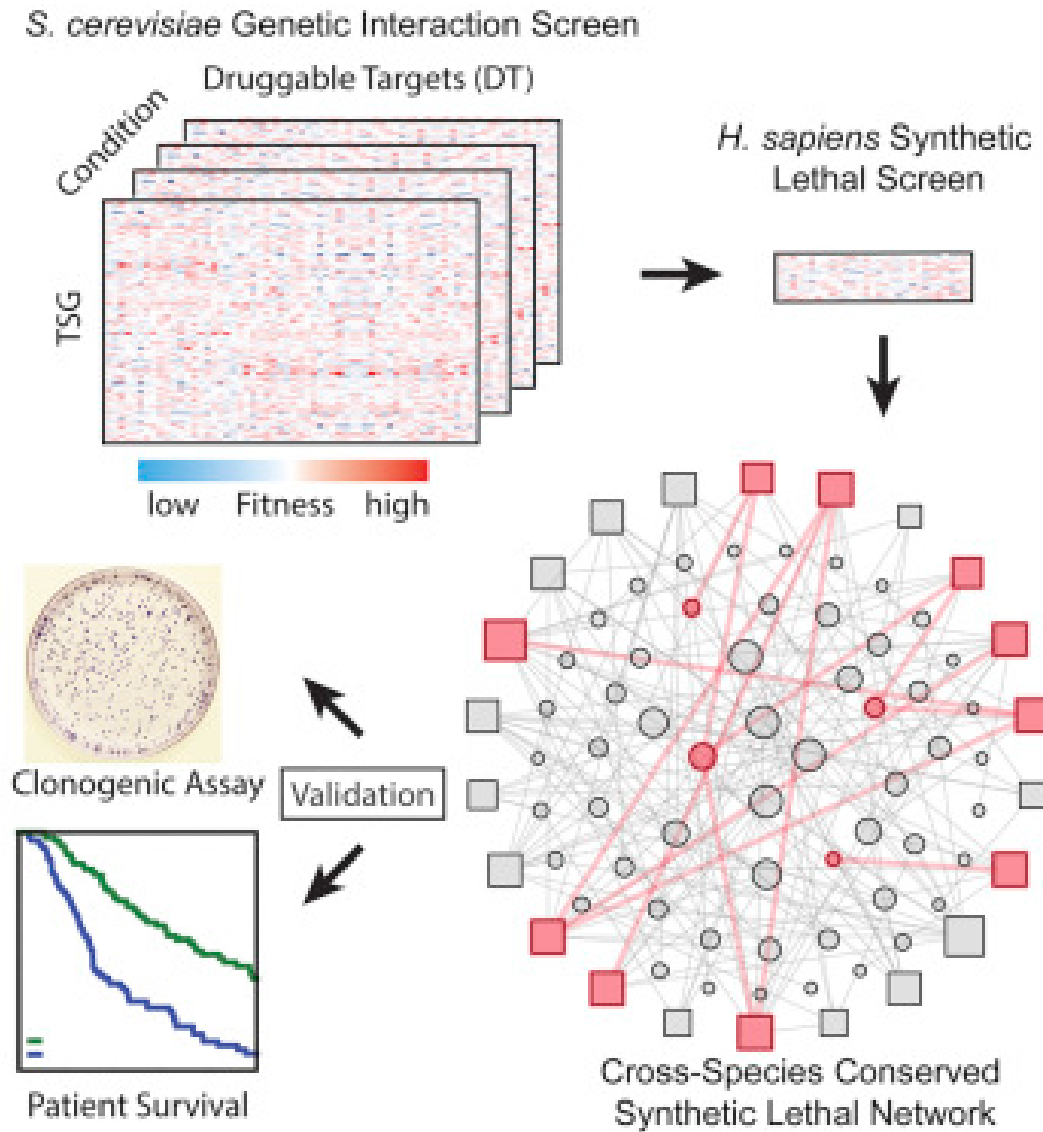


Genetic Interaction maps in Yeast

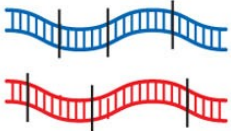
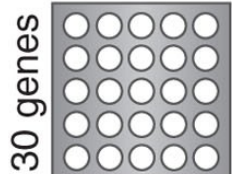

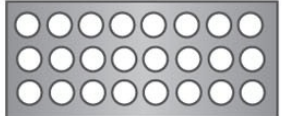
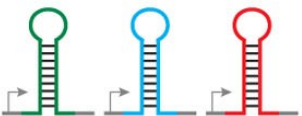
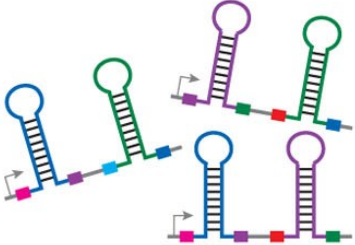



23 million double mutants, identifying about 550,000 negative and about 350,000 positive genetic interactions.

Genetic Interaction maps in mammalian cells

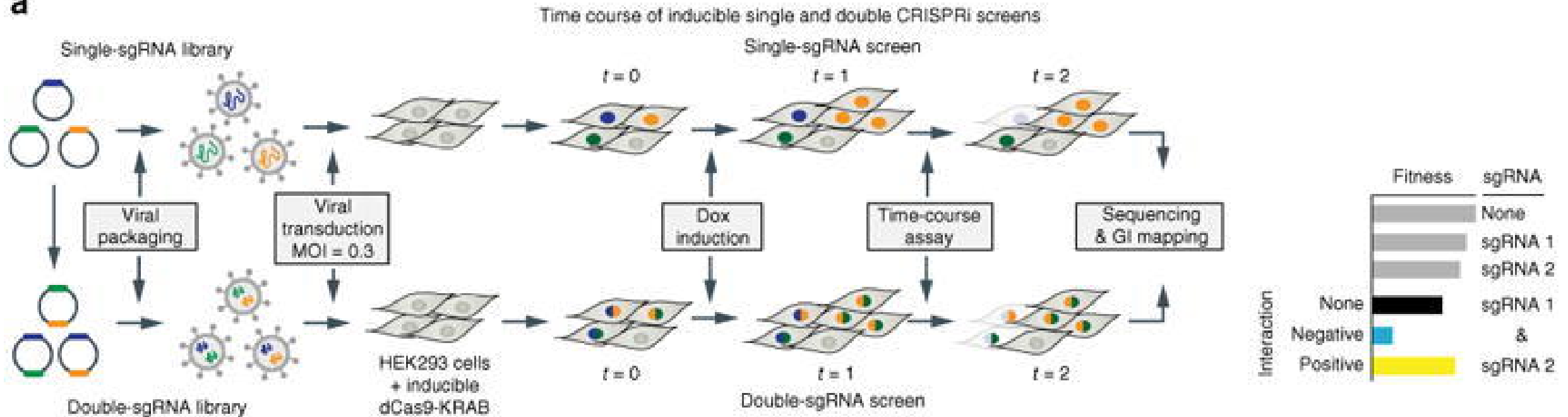


Genetic interaction maps in Mammalian cells: Attempts

	Select genes	Generate library	Perform experiment	Assay	Pros	Cons
Roguev <i>et al.</i> ³	130 genes (chromatin modifiers)	 esiRNA	 130 genes 130 genes	Cell count	Reagent design	Reproducibility
Laufer <i>et al.</i> ²	323 genes (chromatin modifiers)	 siRNA	 20 genes 323 genes	High-content microscopy	Reproducibility; deep phenotyping may improve GI calls	Labor-intensive reagent design
Bassik <i>et al.</i> ¹	 60 genes (hits from primary screen)	 Dual shRNA	 Pooled library	Sequence depth	Reproducibility; pooled libraries and sequencing most scalable	Labor-intensive reagent design; paired hairpins limit scale

Mammalian Cells: CRISPR based screens

a



Mammalian cells: Complexities

1. High off target effects and low KD efficiency with RNAi and shRNA
2. Variability among the cell types chosen.
3. Problems with CRISPR methodologies so far: multiple plasmid transfection or cloning steps, large constructs and complex combinations of promoters, which limit its usage in genome-wide genetic screening due to high risk of losing library components during library construction.
4. The application of barcode to label multiple gRNAs has been found result in half of the mismatch of gRNA-barcodes due to lentiviral template switching

3 papers using different CRISPR based screens to map genetic interactions

Paper -1

[nature](#) > [nature biotechnology](#) > [articles](#) > [article](#)

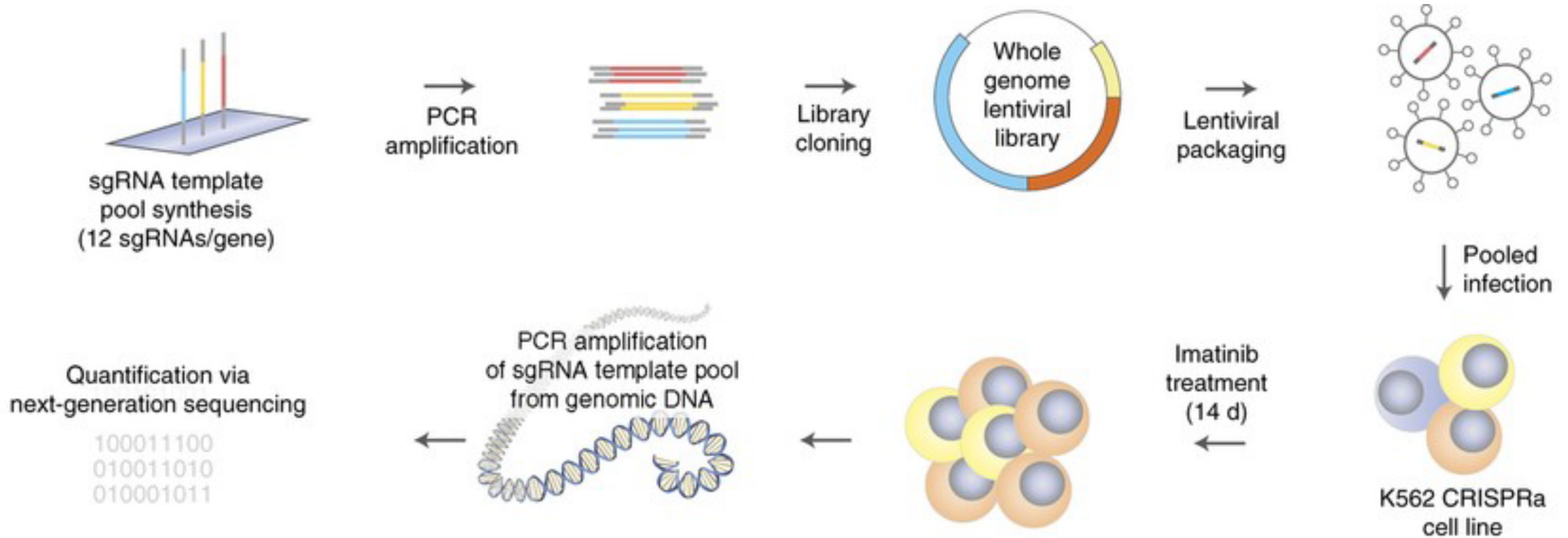
Published: 15 January 2018

Dual gene activation and knockout screen reveals directional dependencies in genetic networks

[Michael Boettcher](#), [Ruilin Tian](#), [James A Blau](#), [Evan Markegard](#), [Ryan T Wagner](#), [David Wu](#), [Xiulei Mo](#), [Anne Biton](#), [Noah Zaitlen](#), [Haian Fu](#), [Frank McCormick](#), [Martin Kampmann](#) & [Michael T McManus](#) 

CRISPRa screen: Identify genes involved in cancer signaling pathways

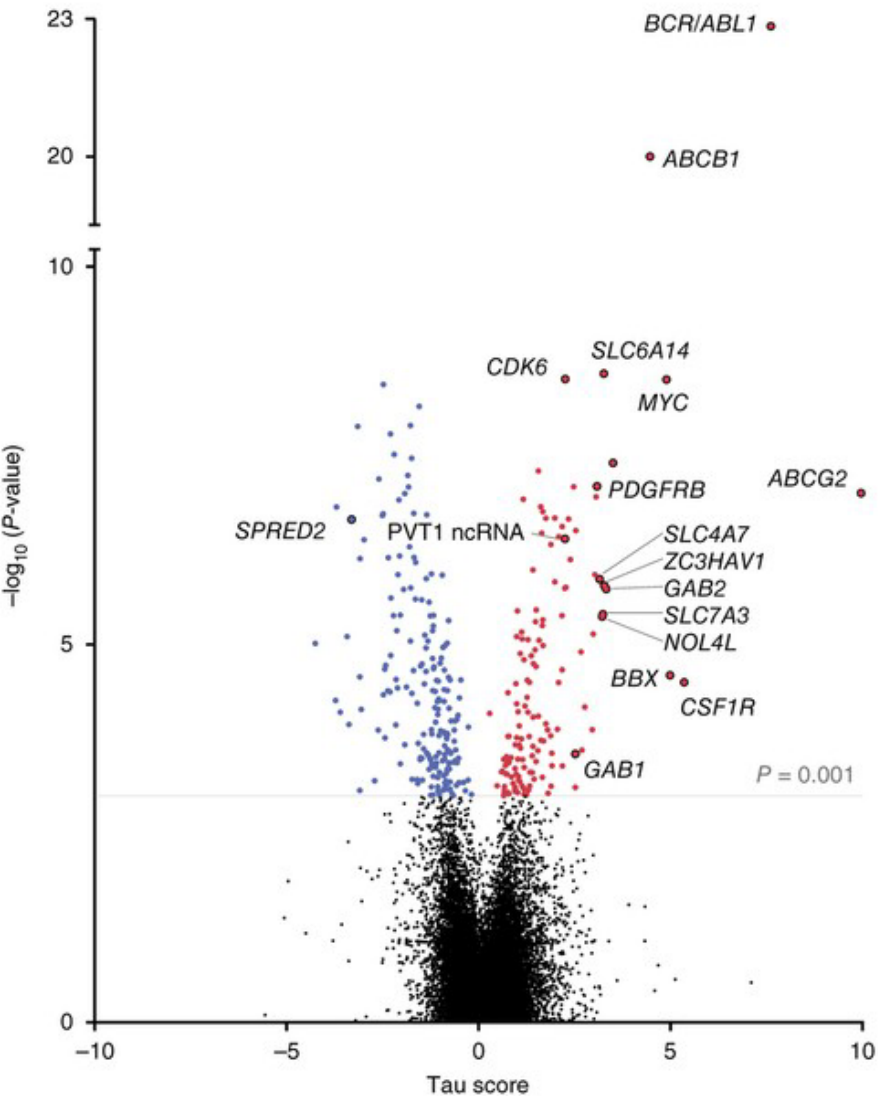
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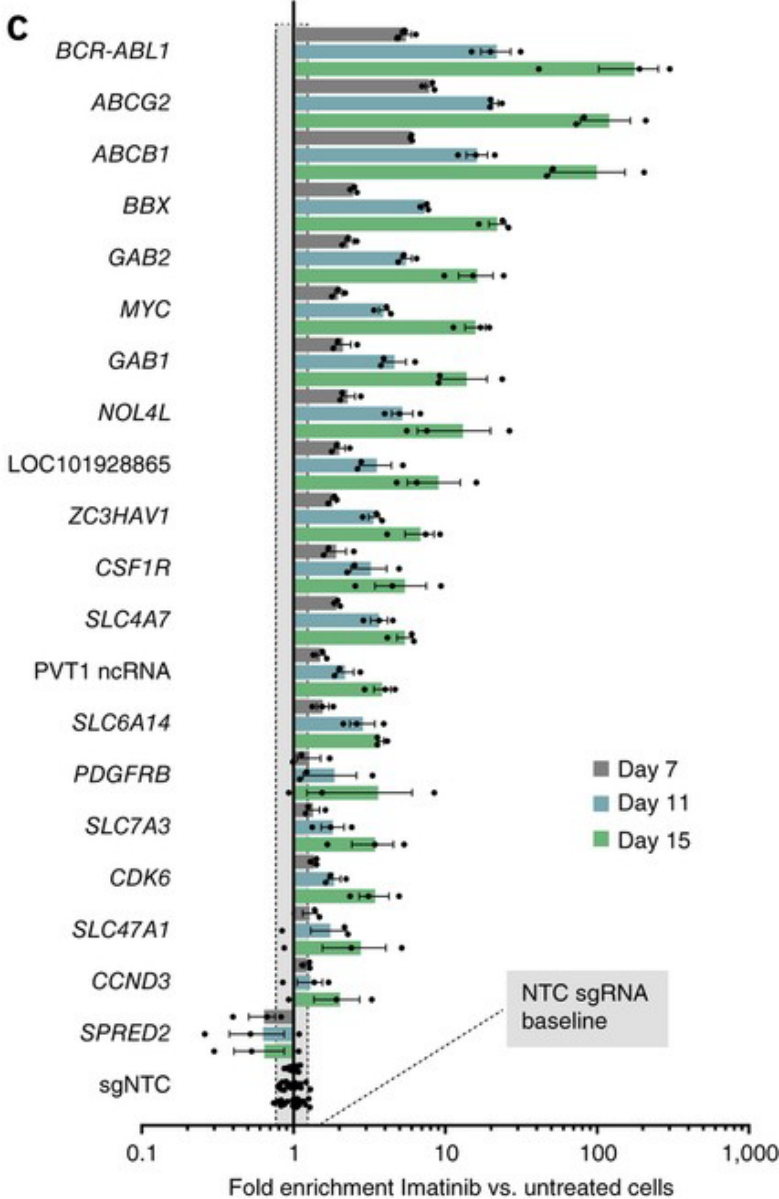
Imatinib is a small molecule kinase inhibitor used to treat certain types of cancer.

Genes altering sensitivity of cells to Imanitib

b

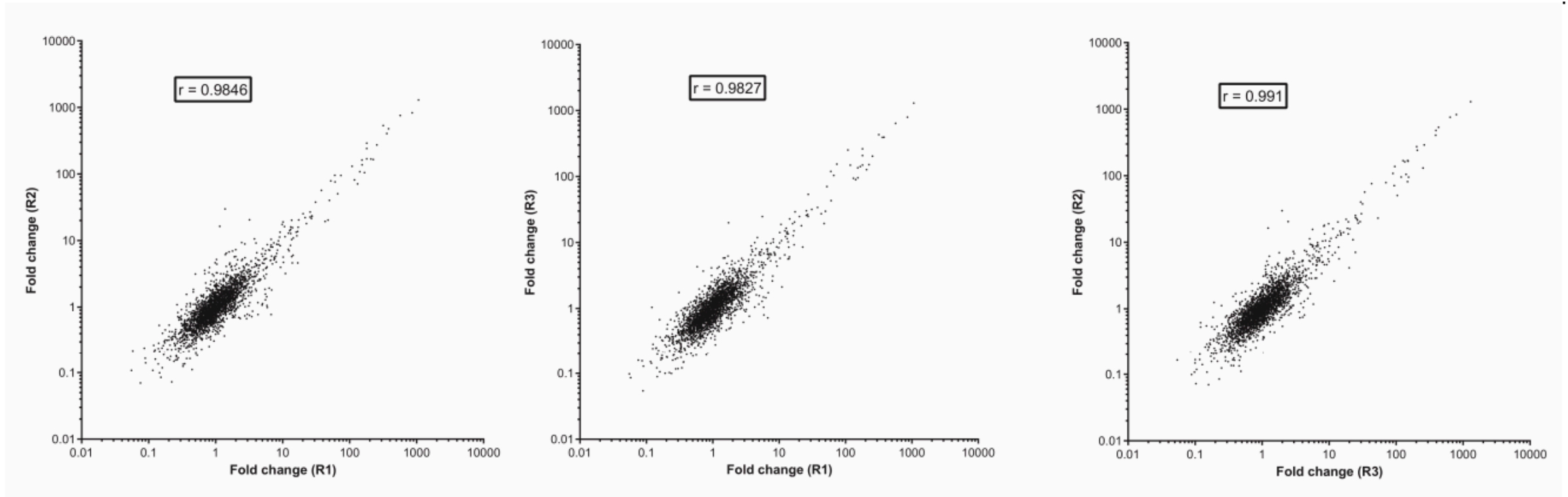


c

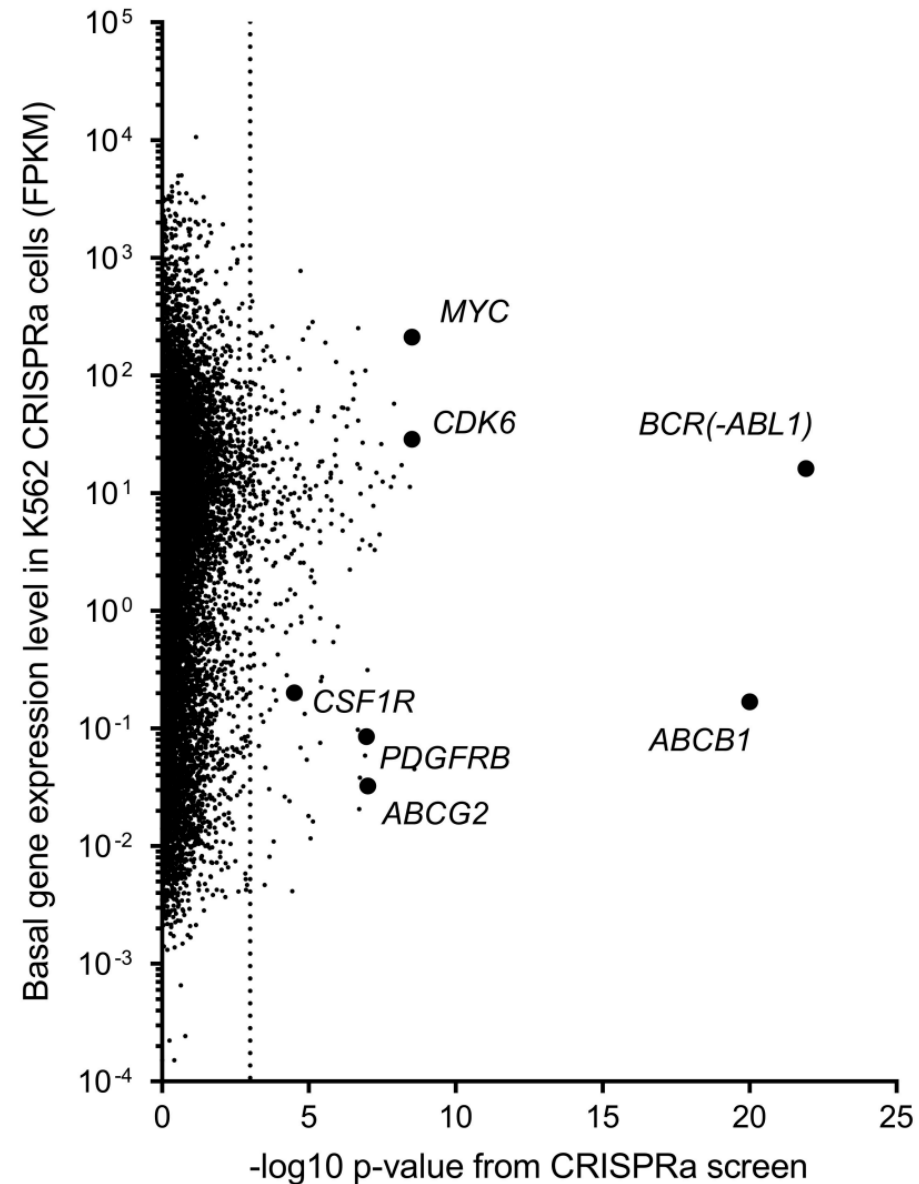


The activation of 332 genes significantly ($FDR < 0.05$, $P < 0.001$) altered the fitness of imatinib-treated K562 cells, with 57% (188 genes) causing significant depletion (blue) and 43% (144 genes) driving cell enrichment

Biological replicates of the screen



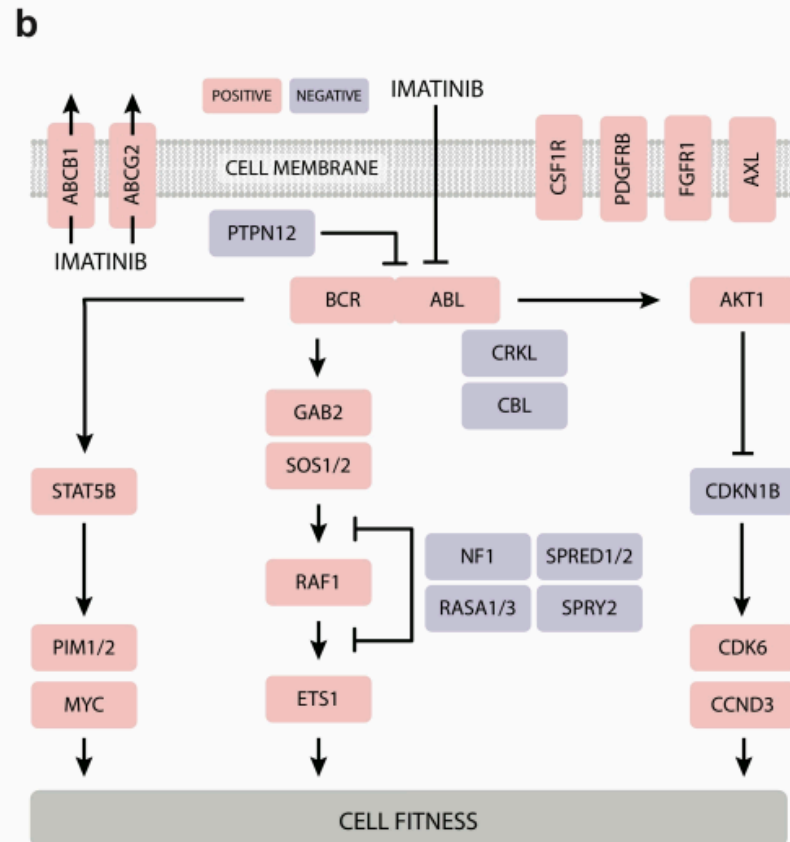
CRISPRa induced phenotypes are independent of endogenous gene expression levels



Pathway reconstruction from CRISPRa screen candidate genes

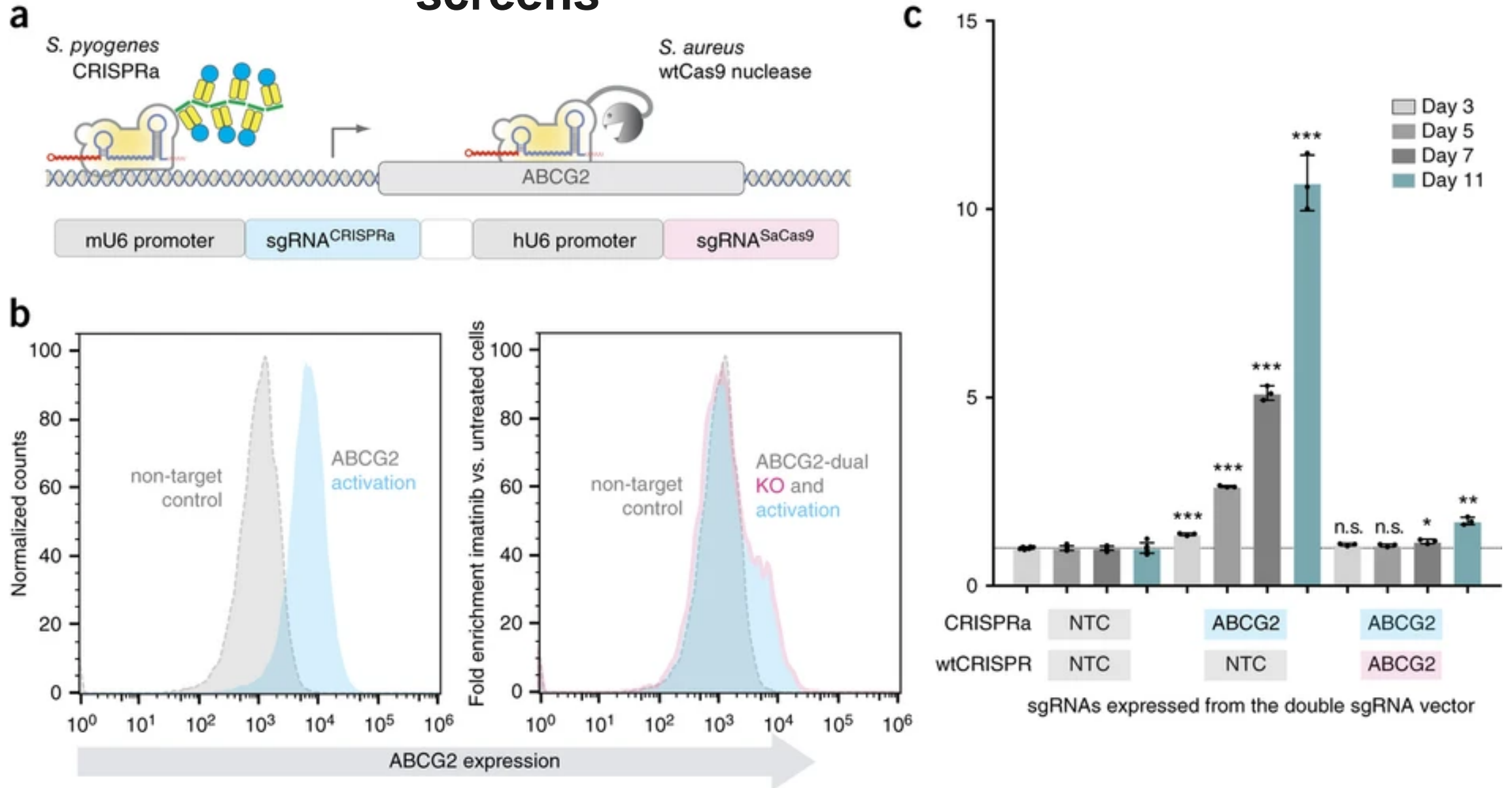
a

KEGG Pathway	Enrichment	p	FDR
Acute myeloid leukemia	11.54	6.6E-10	8.08E-07
Chronic myeloid leukemia	9.67	1.2E-09	1.45E-06
ErbB signaling pathway	8.00	1.3E-08	1.63E-05
Pathways in cancer	3.42	4.5E-08	5.49E-05
Transcriptional misregulation in cancer	5.03	1.9E-07	2.30E-04
MAPK signaling pathway	3.90	6.0E-07	7.45E-04



Subset of identified candidate genes mapped onto known signalling pathways (blue = negative, red = positive regulator of fitness)

Orthogonal CRISPR screens

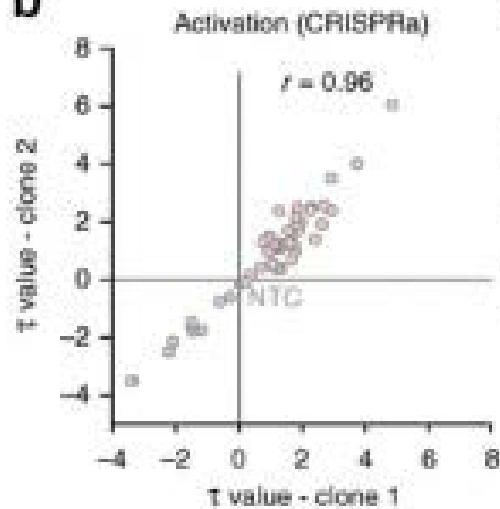


Orthogonal CRISPR screens can quantify directional genetic interactions.

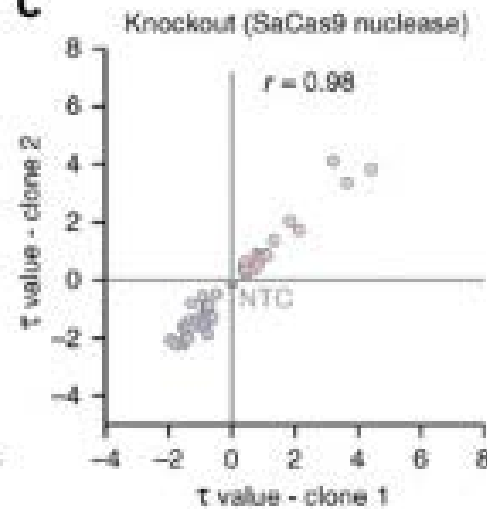
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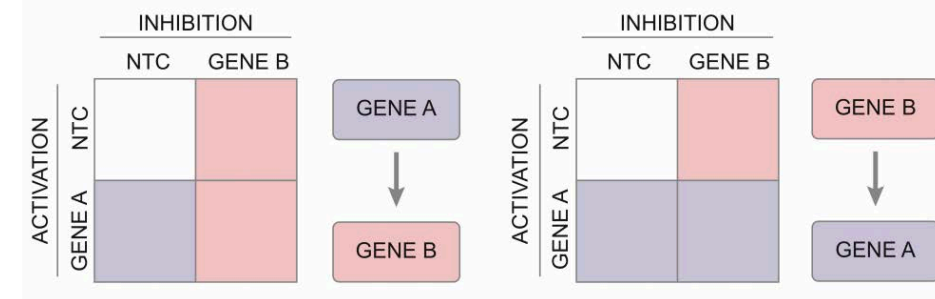
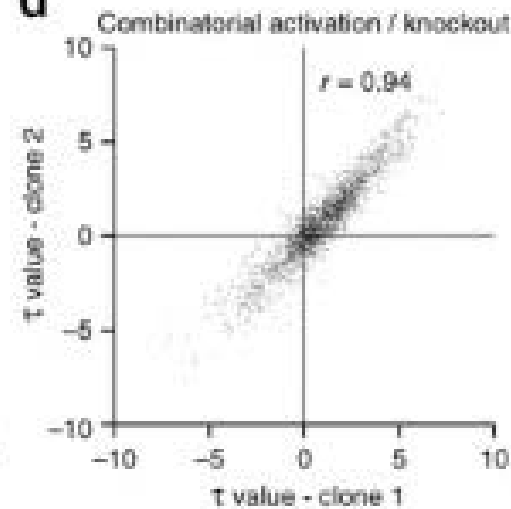
b



c



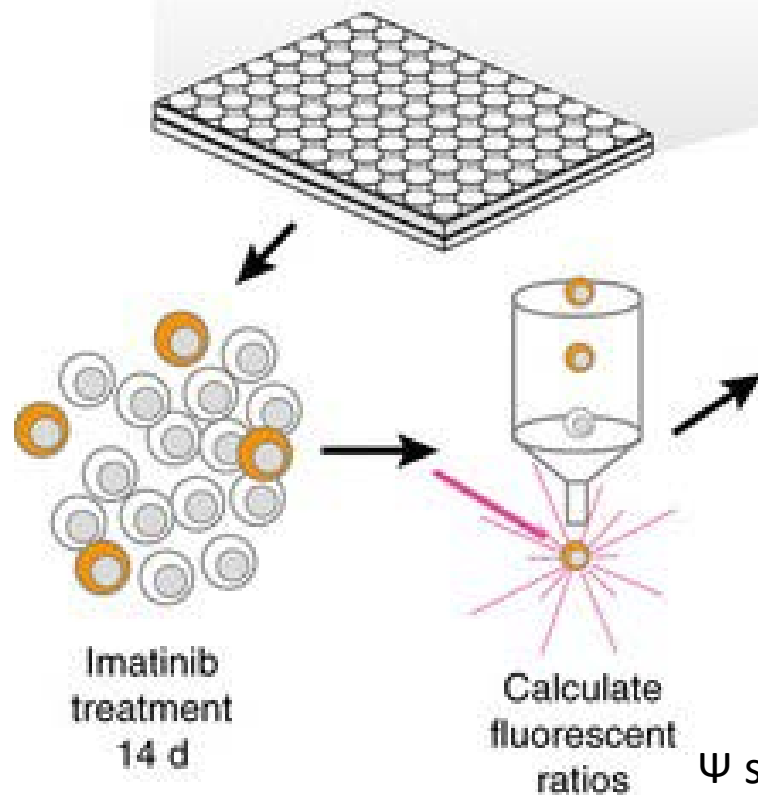
d



Orthogonal CRISPR screen: Workflow

a

sgRNA ^{activation}		sgRNA ^{knockout}		tau values (τ)
mU6	Gene A	hU6	NTC	Single activation (act)
mU6	NTC	hU6	Gene B	Single knockout (KO)
mU6	Gene A	hU6	Gene B	Combined activation and knockout (act+KO)



Calculate GI values

$$GI_v = (\tau_{act+ko}) - (\tau_{act} + \tau_{ko})$$

Calculate Ψ values

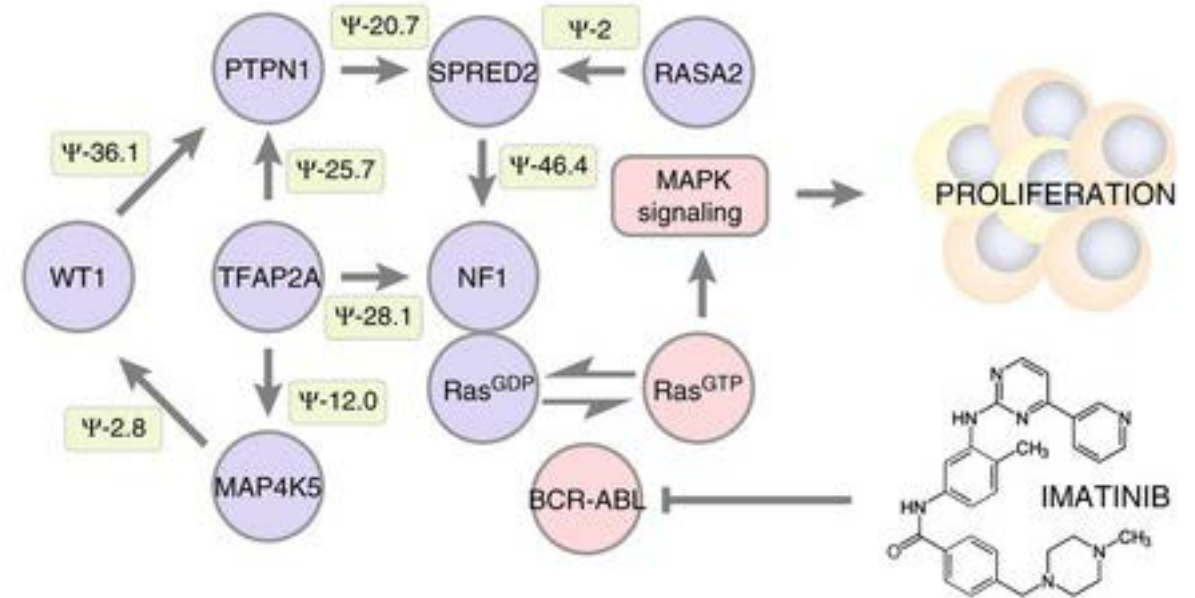
$$\Psi_v = \tau_{act} \times \tau_{ko} \times GI_v^2$$

Ψ scores define interactions in which directionality could be inferred

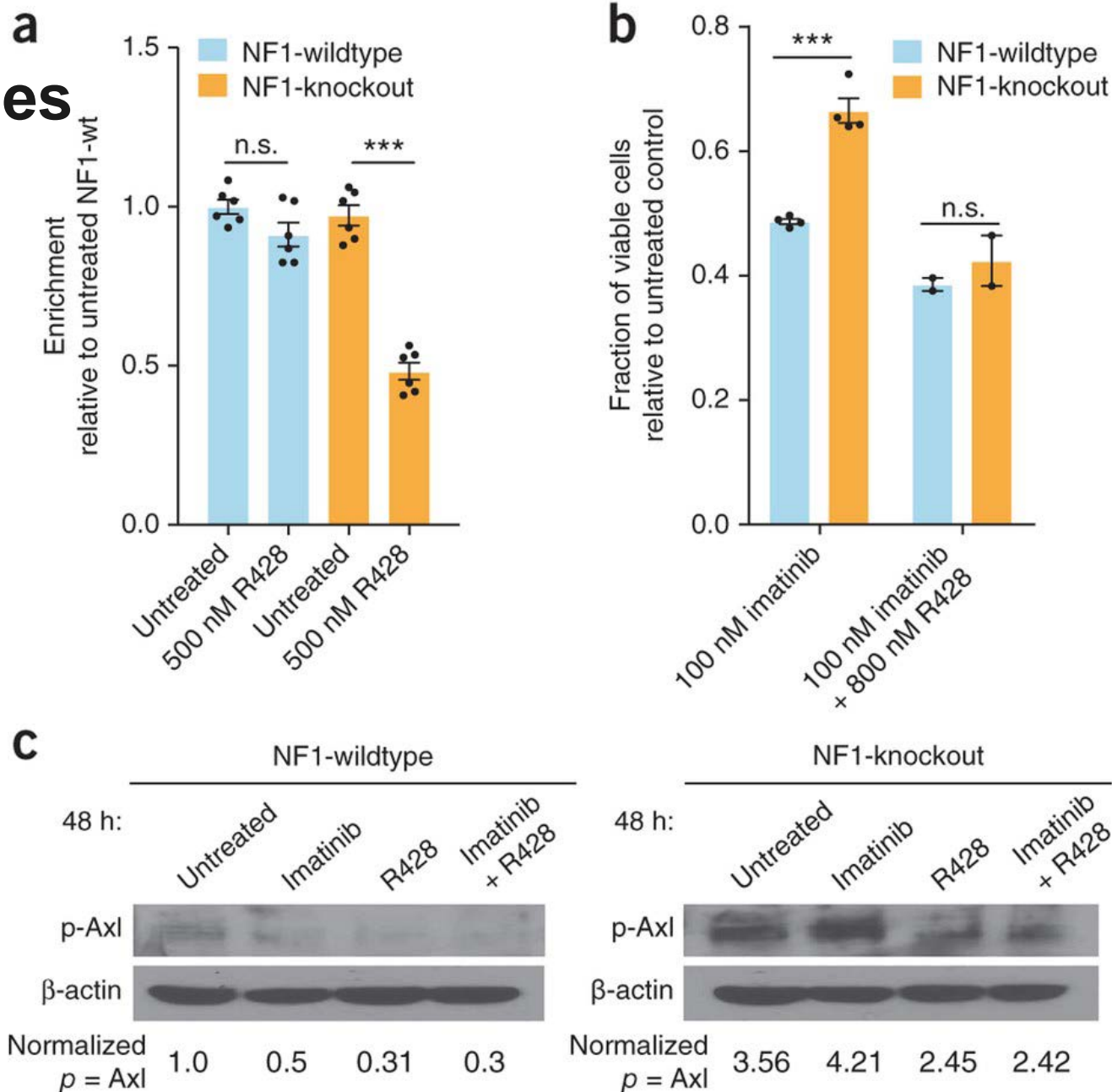
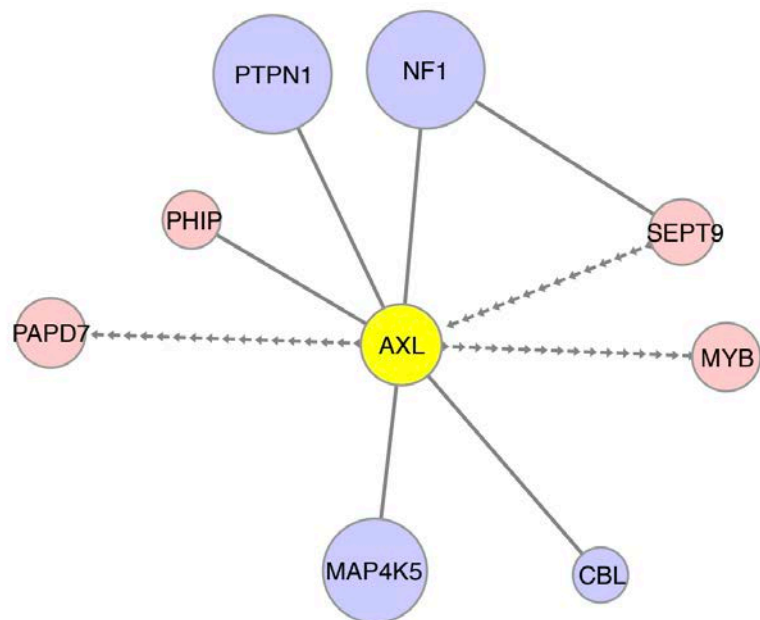
Validation of a directional Ras-centric genetic subnetwork



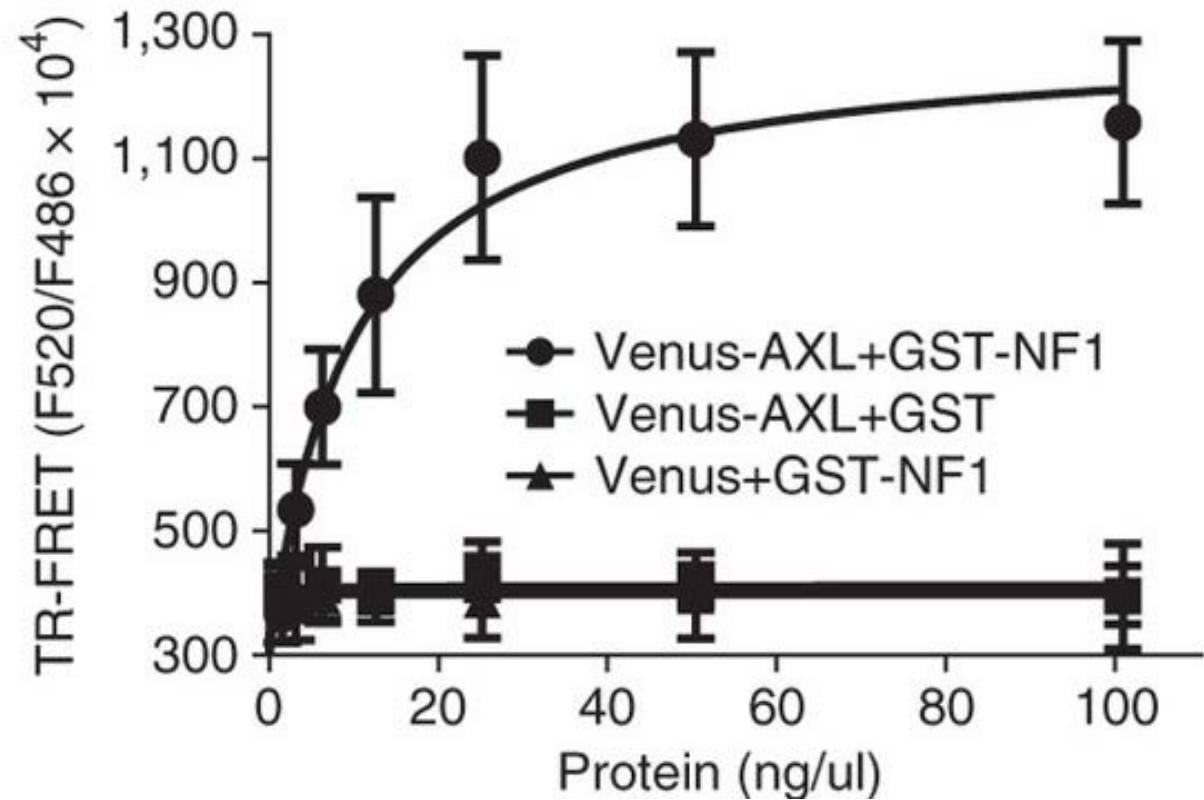
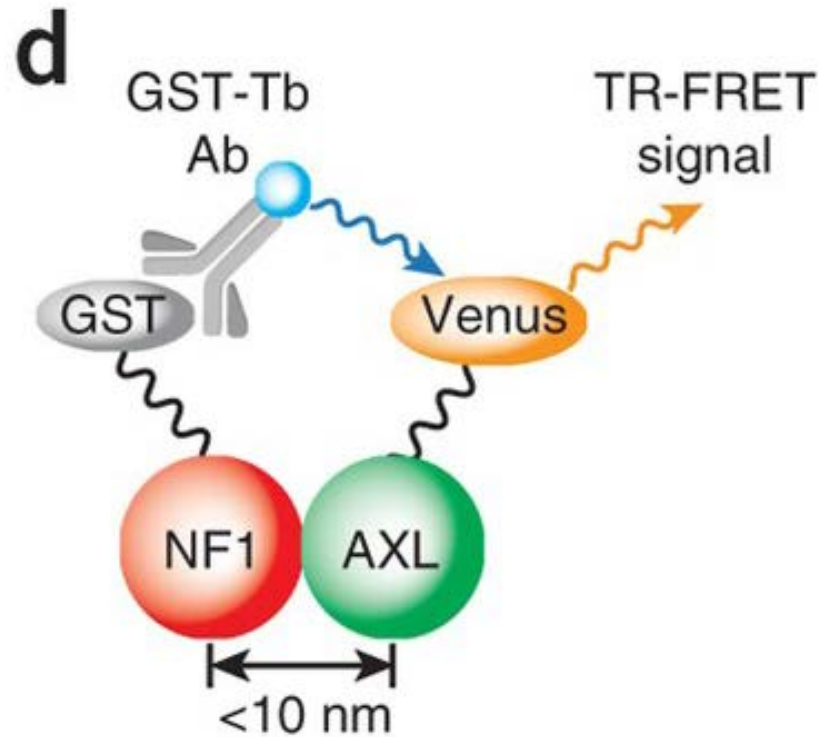
c



Exploiting genetic dependencies for cancer therapy.



Physical interaction between NF1 and AXL




Conclusions

1. Establishment of methods / tools to perform orthogonal screens.
2. Such screens help in identifying directionality for genetic interactions
3. Loss and gain of functions can be quantified from same cells

Paper -2

Article | [Open Access](#) | Published: 06 February 2020

Genetic screens in isogenic mammalian cell lines without single cell cloning

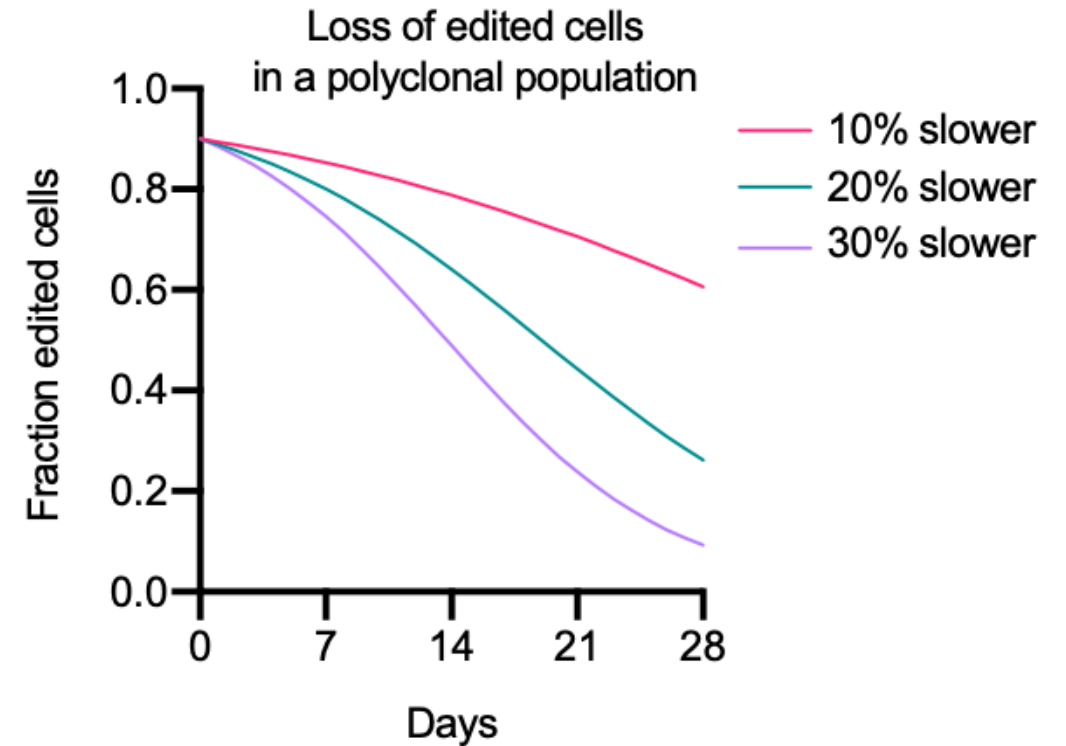
Peter C. DeWeirdt, Annabel K. Sangree, Ruth E. Hanna, Kendall R. Sanson, Mudra Hegde, Christine Strand, Nicole S. Persky & John G. Doench 

Nature Communications **11**, Article number: 752 (2020) | [Cite this article](#)

5877 Accesses | **4** Citations | **21** Altmetric | [Metrics](#)

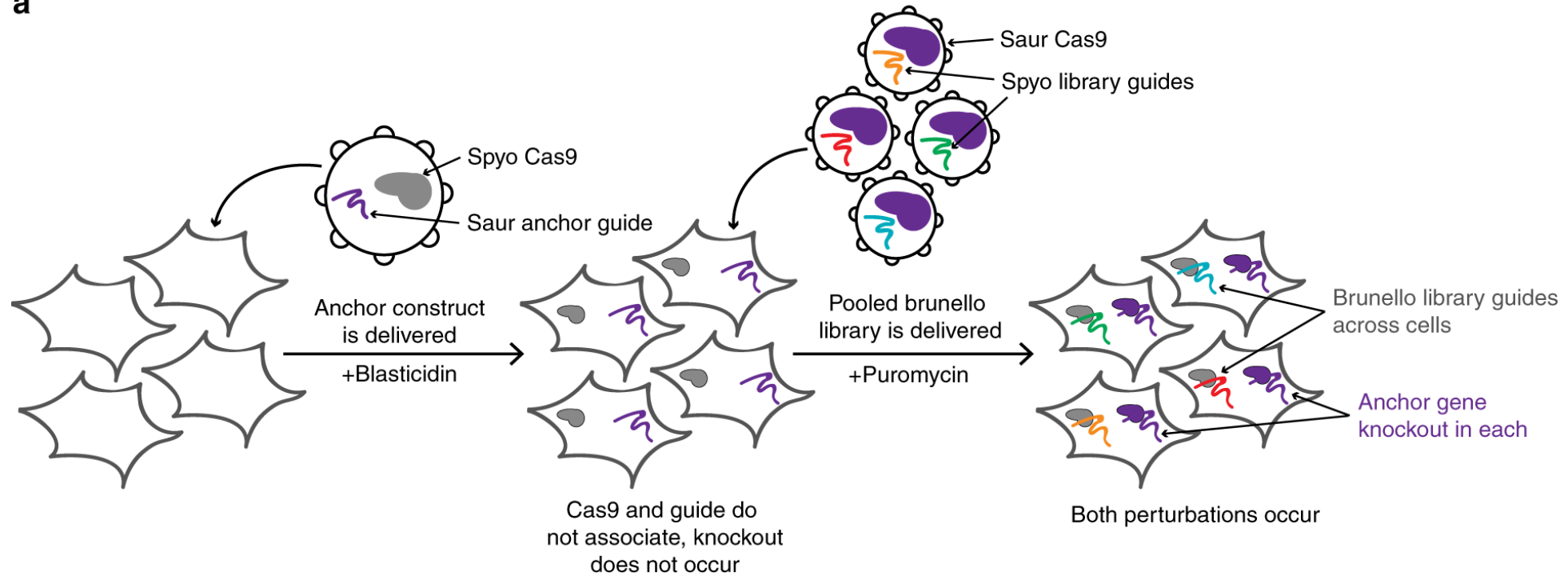
Problems with synthetic lethality screens

1. Selection pressure against ko cells
2. Cells harboring antibiotic resistance but not gRNA

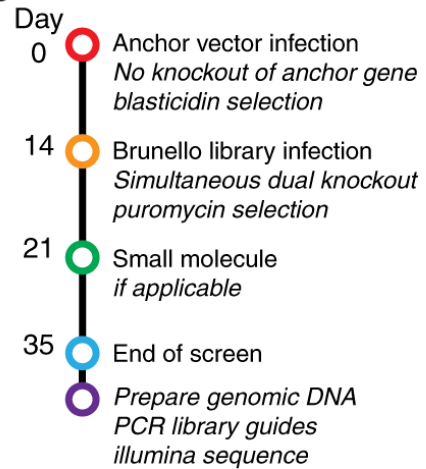


Anchor screen: Methodology

a

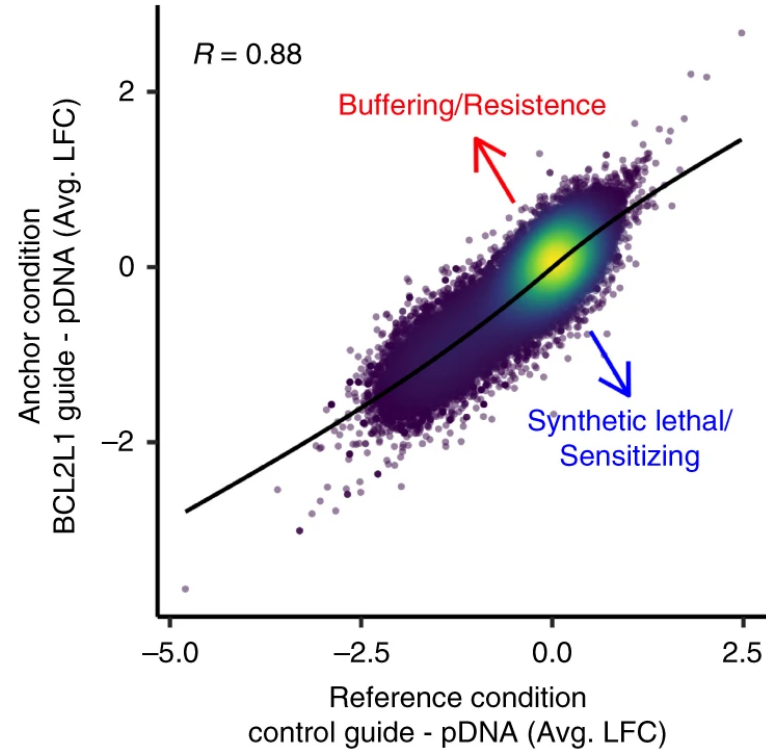


b

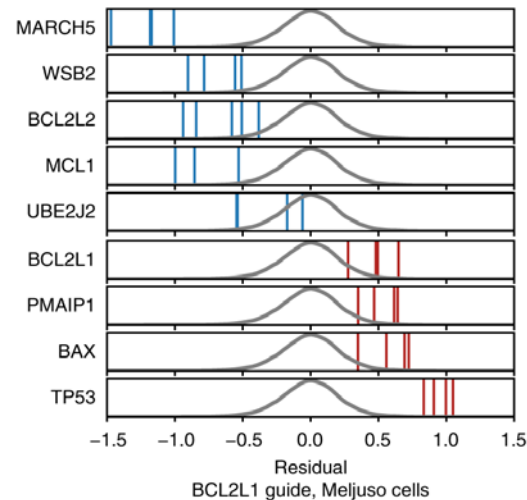
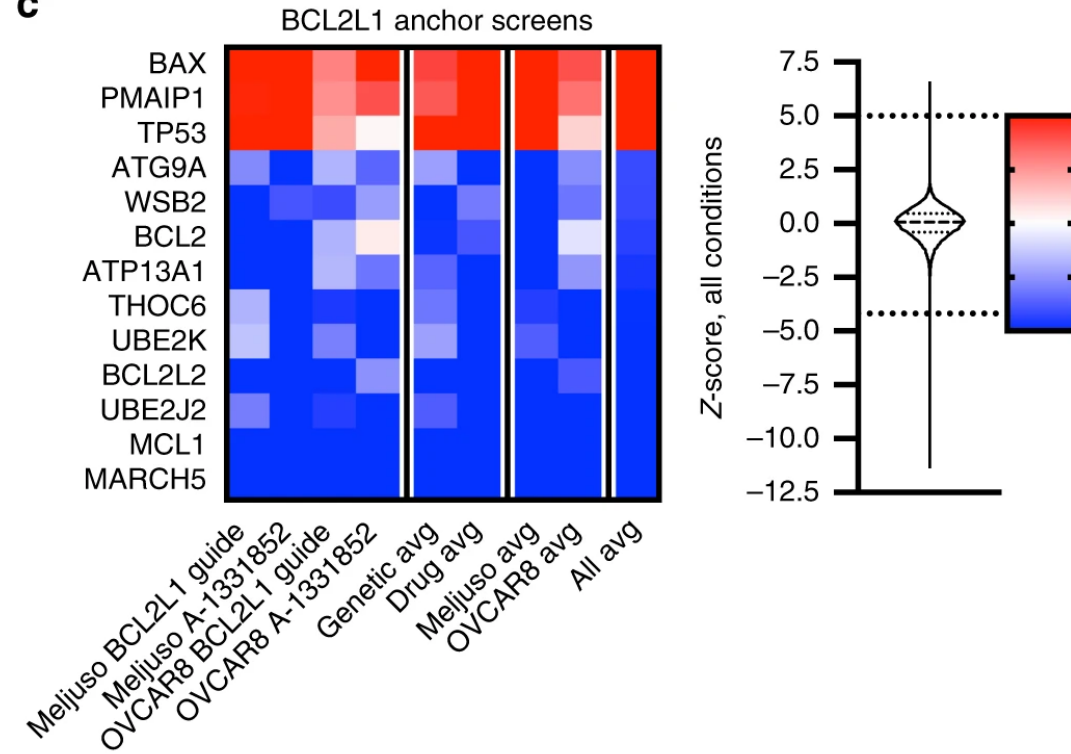


Anchor screen for BCL2L1. Synthetic lethality

a

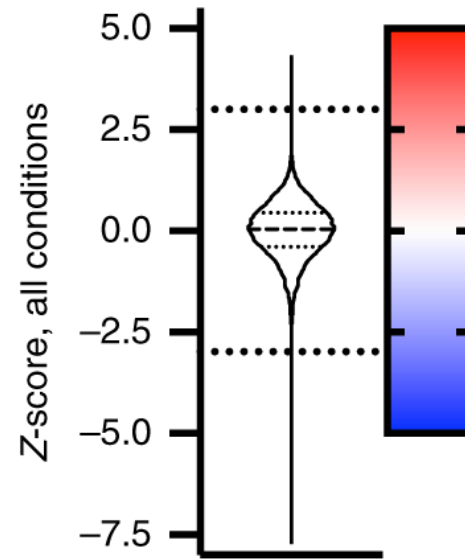
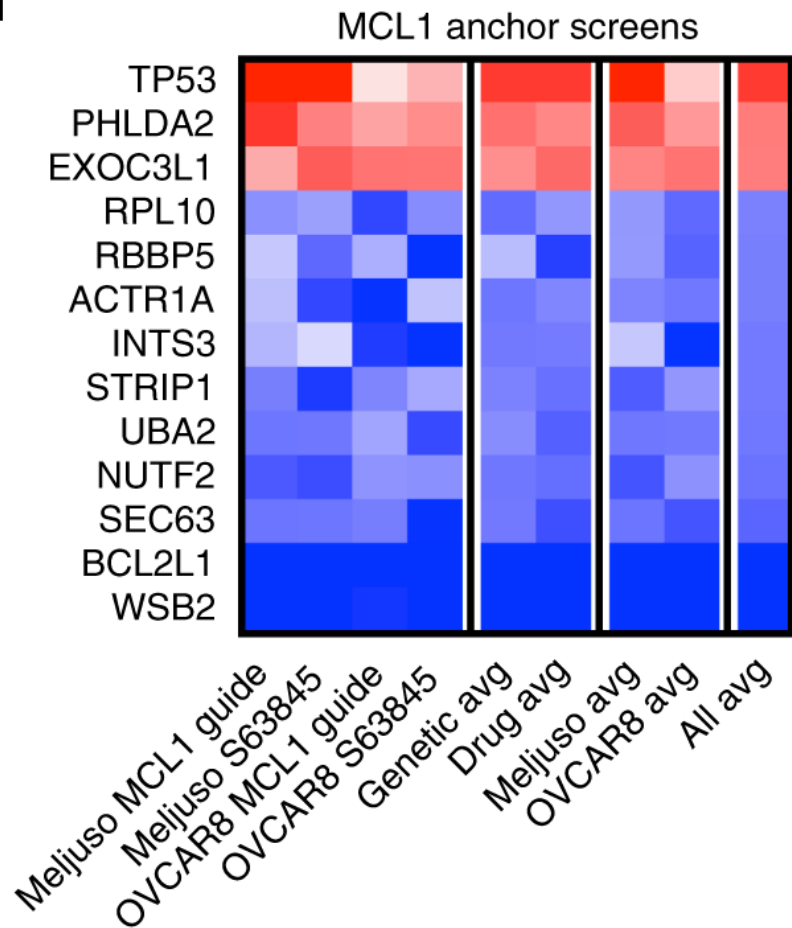


c

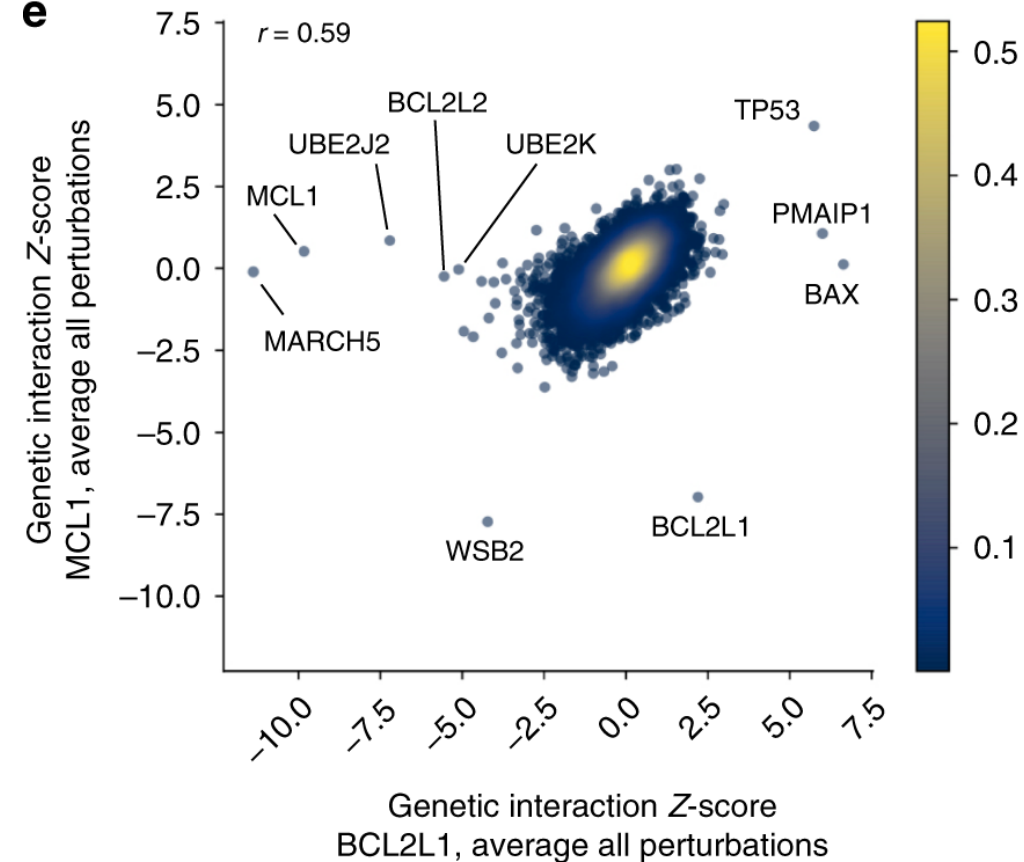


Anchor screen for MCL1. Synthetic lethality

d

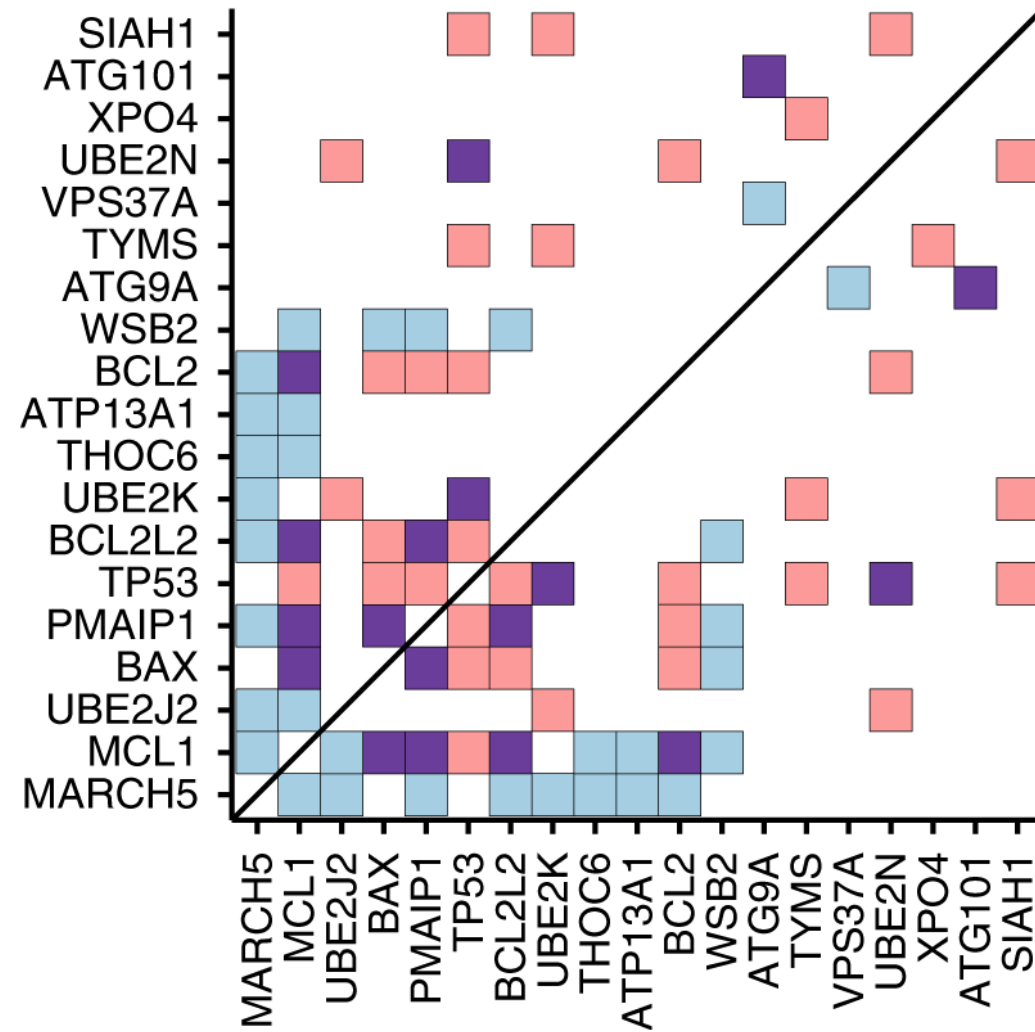


e

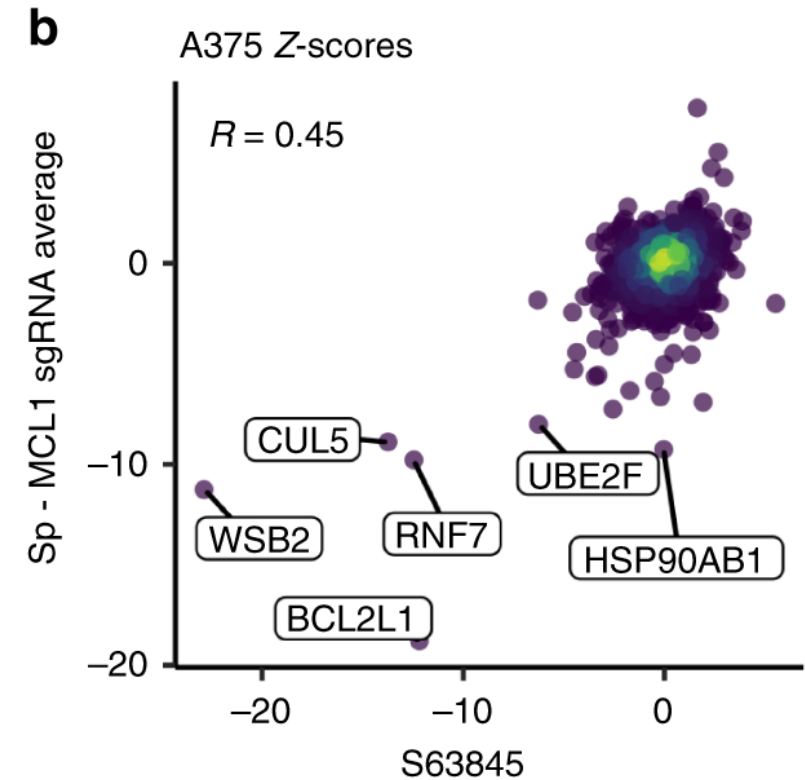
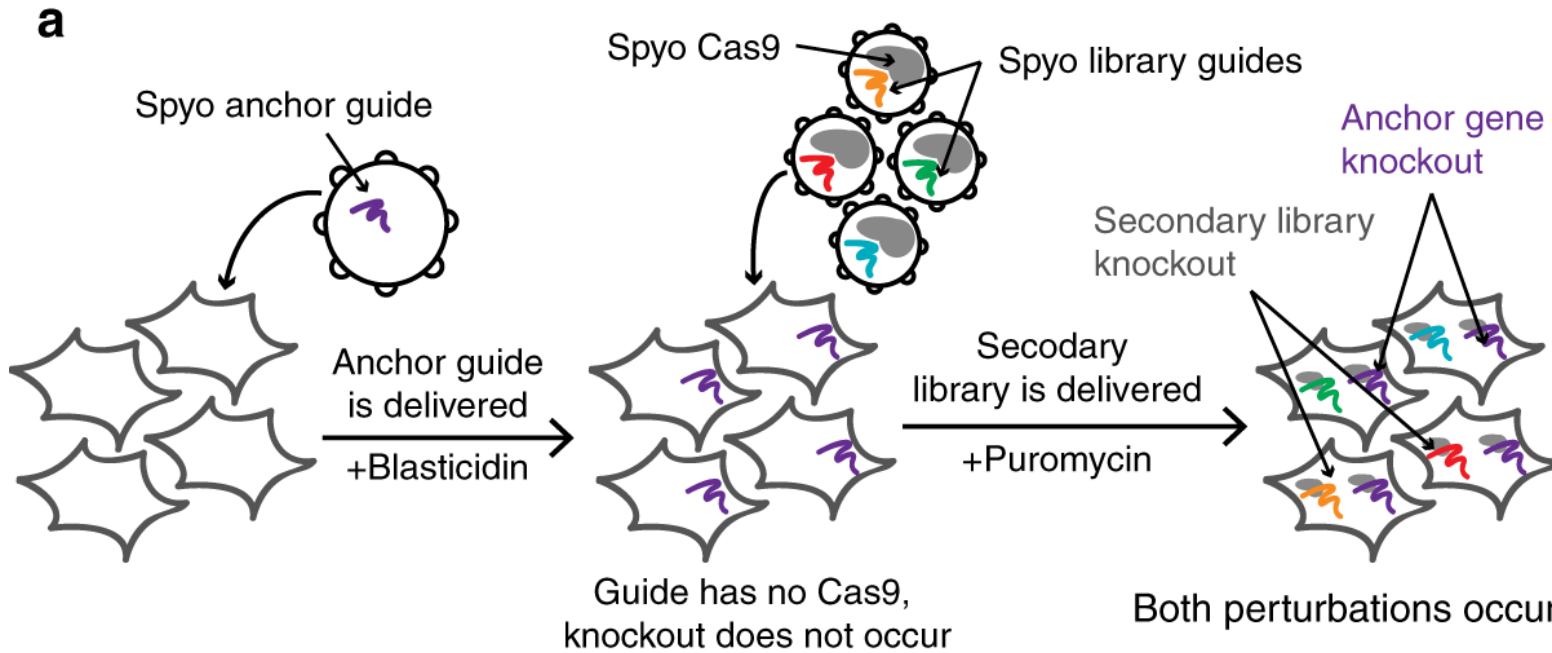


C

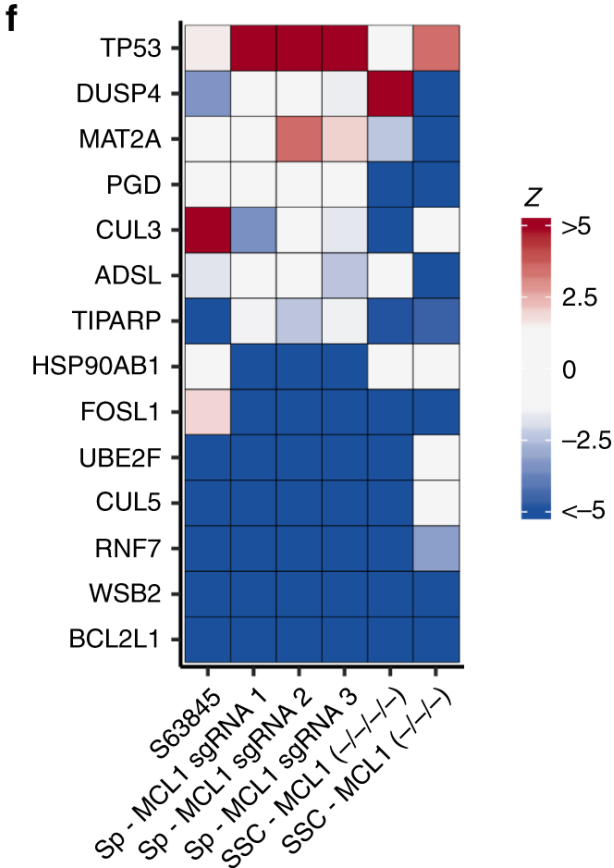
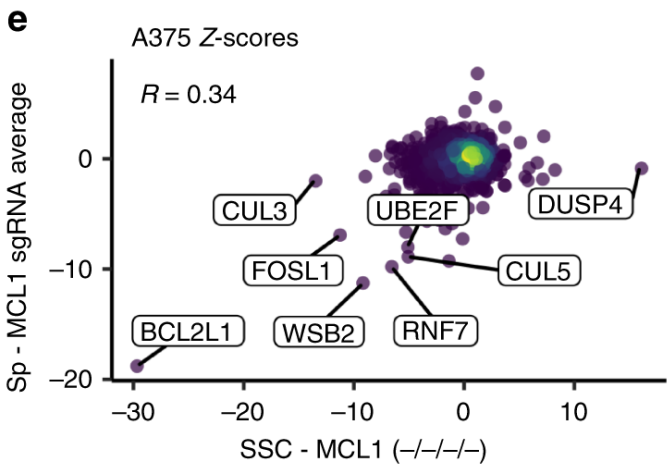
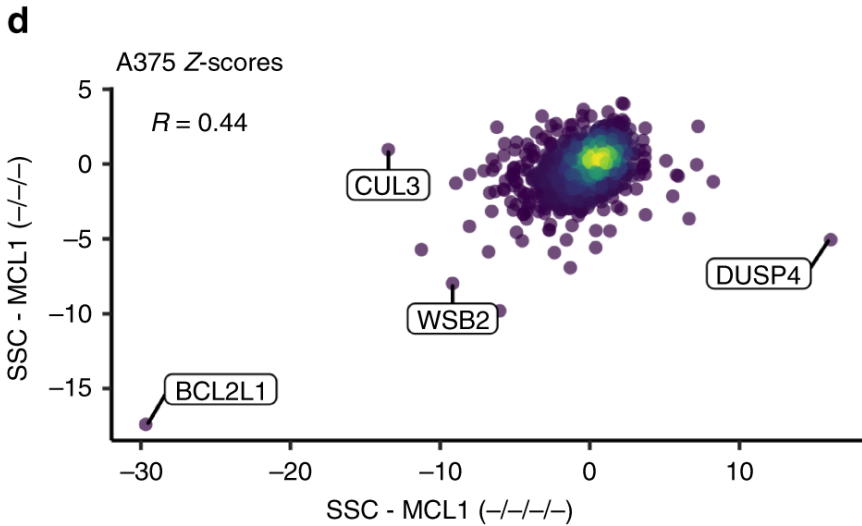
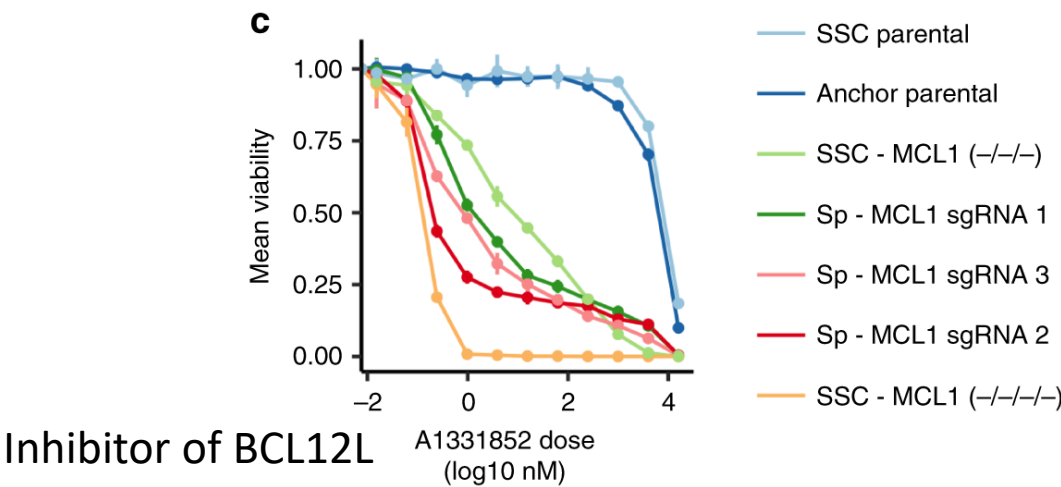
Category  Both  Co-essentiality  String



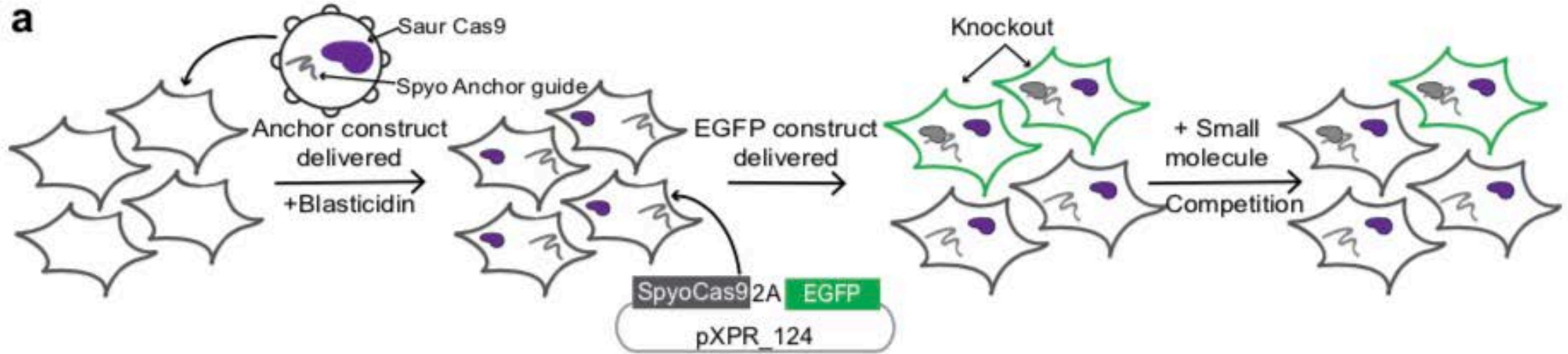
MCL1 secondary screens comparing alternative screening approaches.

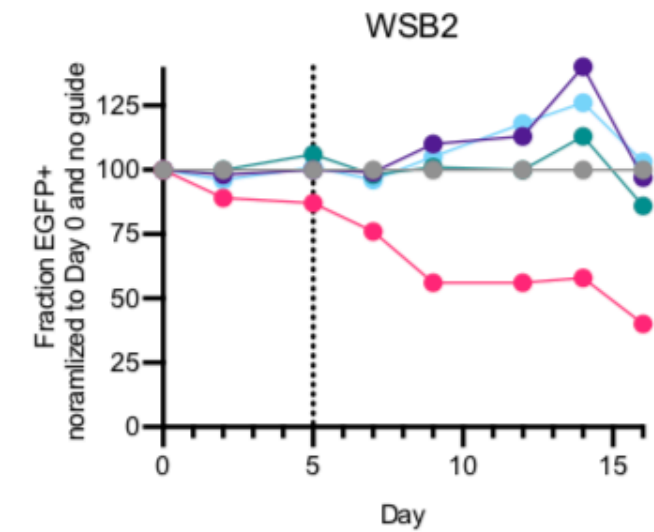
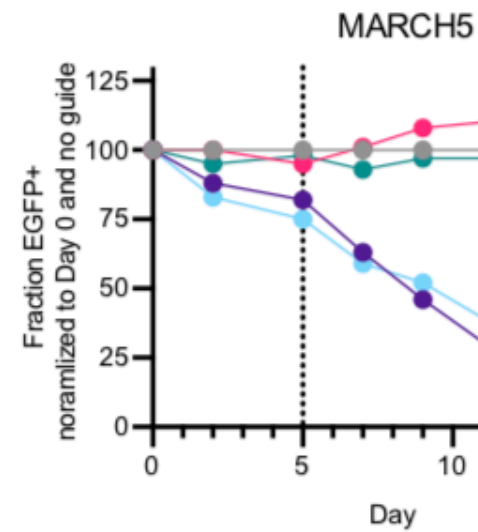
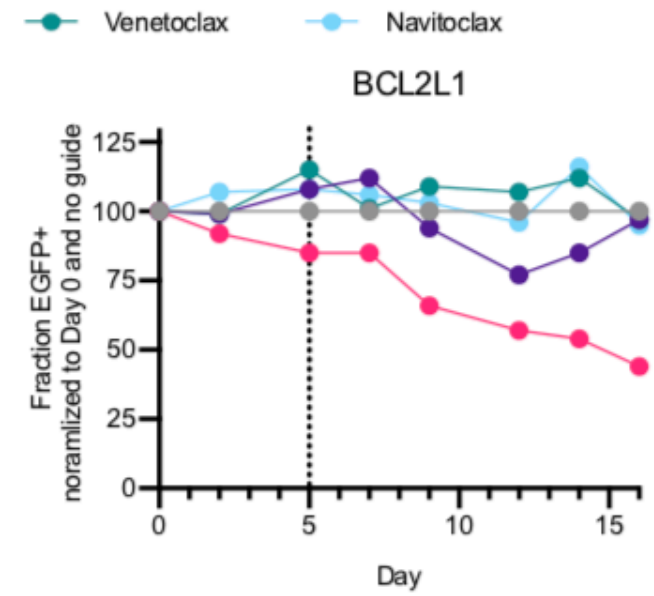
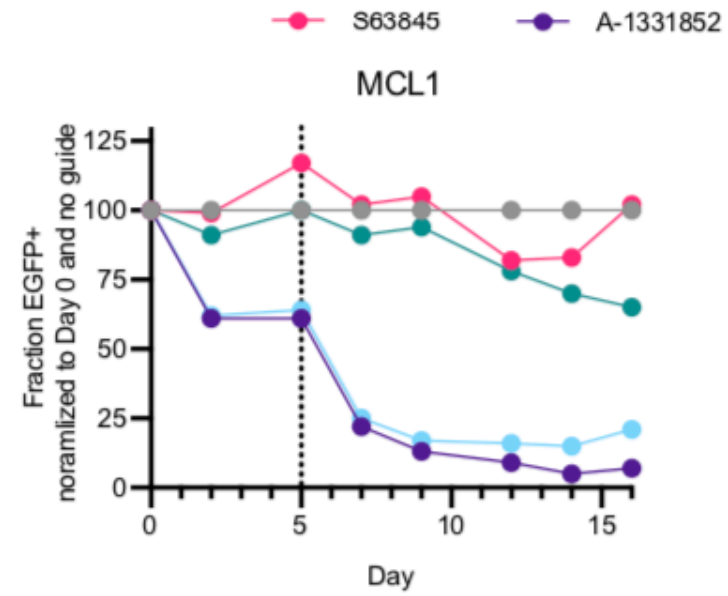


Validation



Validation of the data using a competition assay

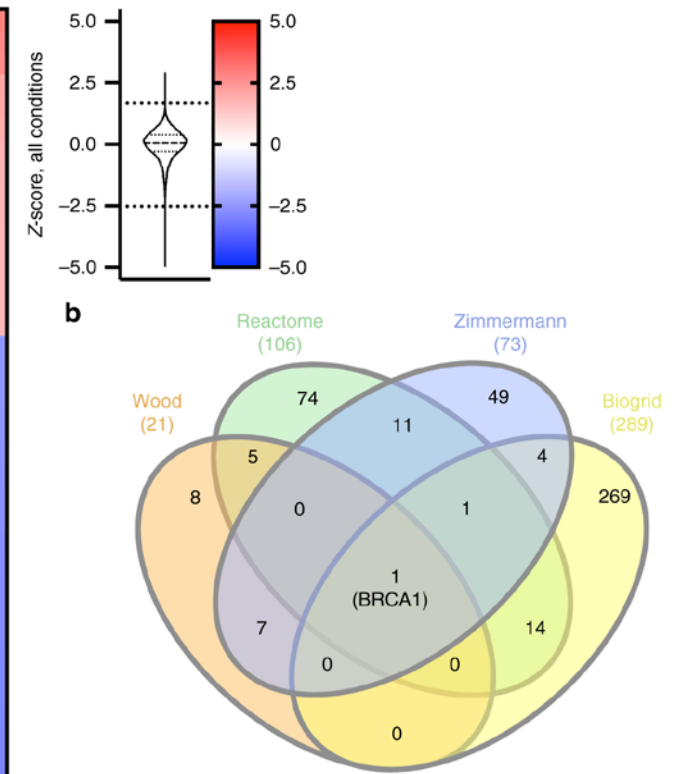
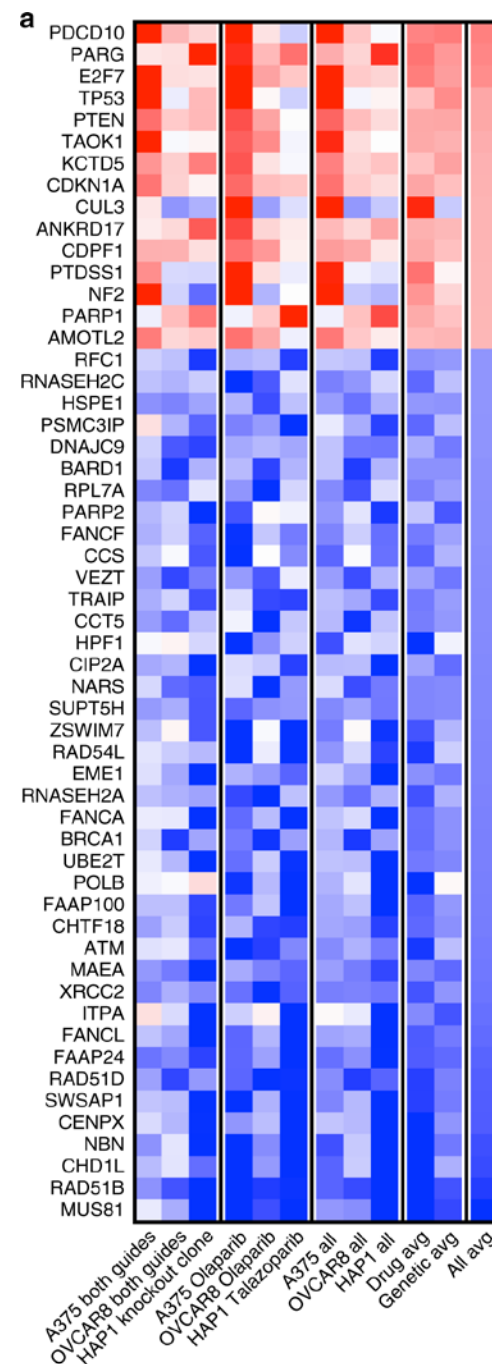


b

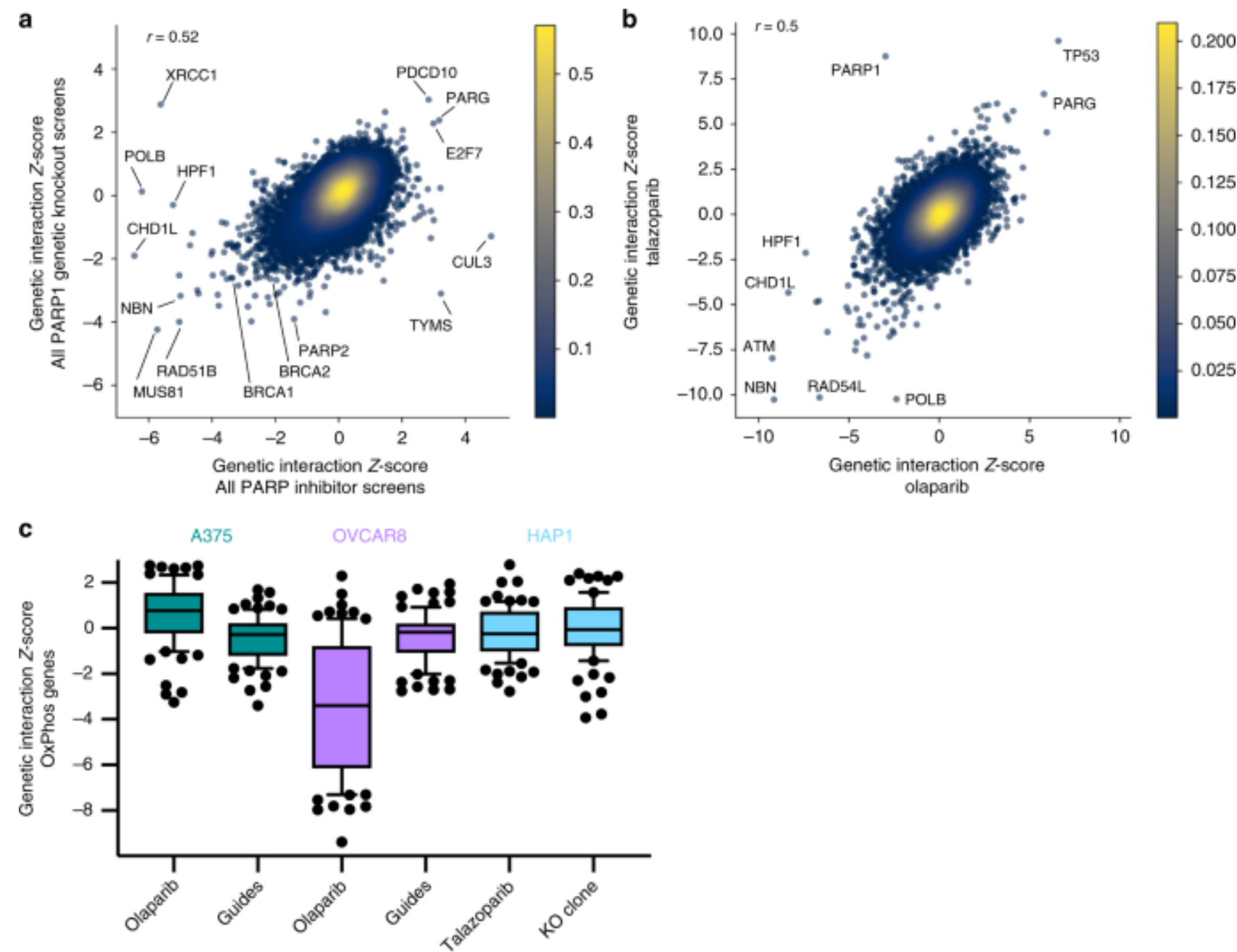
Anchor screen with DNA damage response gene *PARP1*

Screen was performed in multiple cell lines.

Also performed with two inhibitors of PARP1



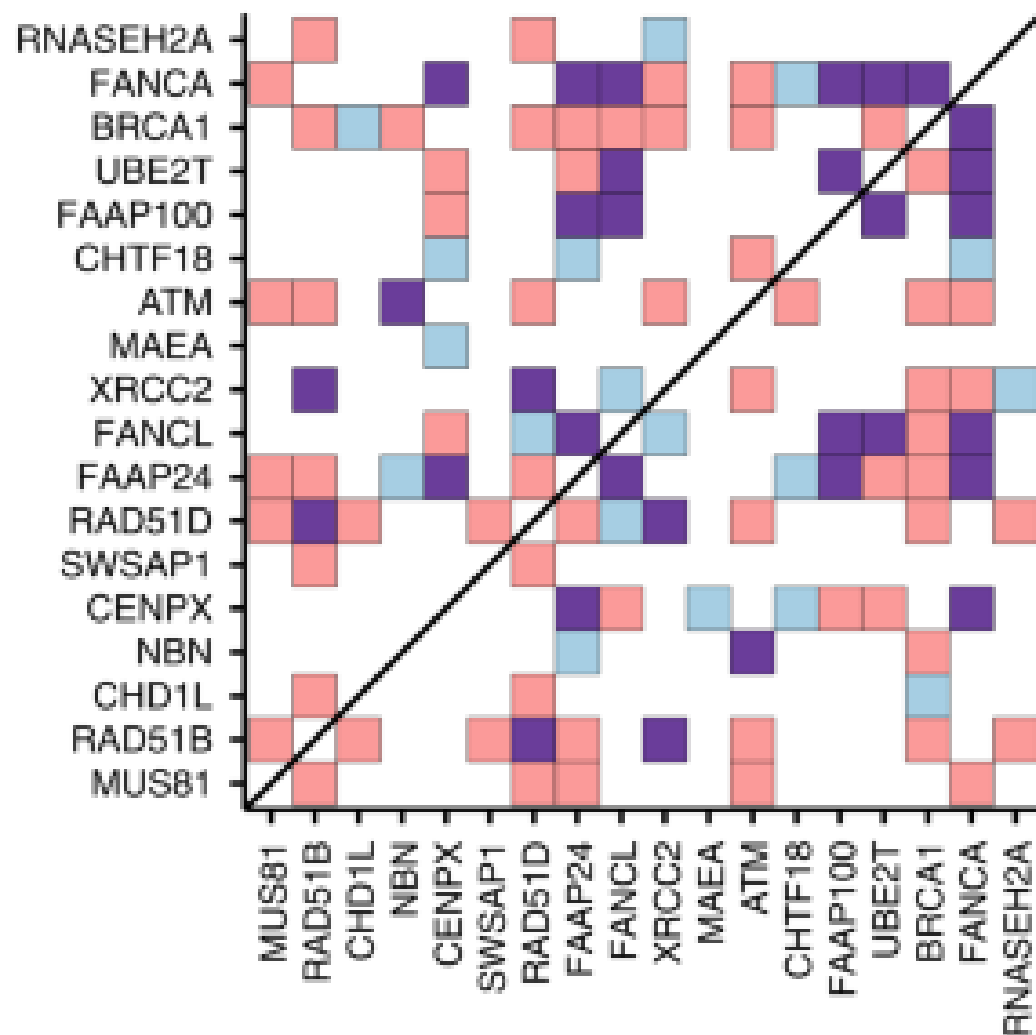
GI Score: Residuals of all the guides/
SD of all the guides



Creating networks from the data obtained

a

Category  Both  Co-essentiality  String



Conclusions

1. A smart way to perform synthetic lethality screens overcoming the need for an inducible system.
2. Coessentiality data can be used to generate genetic interaction maps.

Questions:

1. Stability of the anchor guides?
2. Can we use patient derived cell lines?

Paper -3

Cell Reports

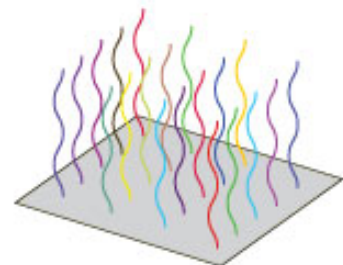


Article

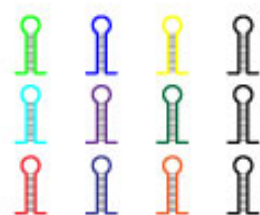
A Three-Way Combinatorial CRISPR Screen for Analyzing Interactions among Druggable Targets

Peng Zhou,¹ Becky K.C. Chan,¹ Yuk Kei Wan,¹ Chaya T.L. Yuen,¹ Gigi C.G. Choi,¹ Xinran Li,² Cindy S.W. Tong,¹ Sophia S.W. Zhong,¹ Jieran Sun,¹ Yufan Bao,^{3,4} Silvia Y.L. Mak,³ Maggie Z.Y. Chow,³ Jien Vei Khaw,¹ Suet Yi Leung,^{5,6,7} Zongli Zheng,^{3,4,8} Lydia W.T. Cheung,² Kaeling Tan,^{9,10} Koon Ho Wong,^{9,11} H.Y. Edwin Chan,^{12,13} and Alan S.L. Wong^{1,14,15,*}

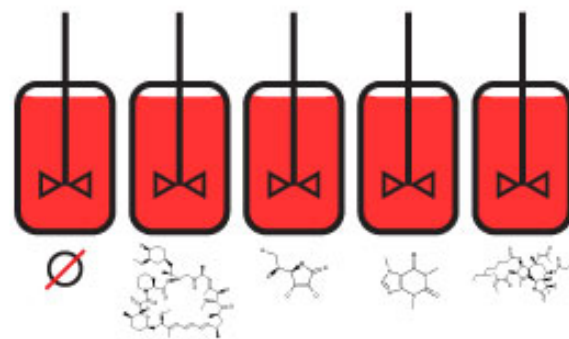
Oligonucleotide Library
Synthesis



Infect cells
with high-coverage
sh/sgRNA library



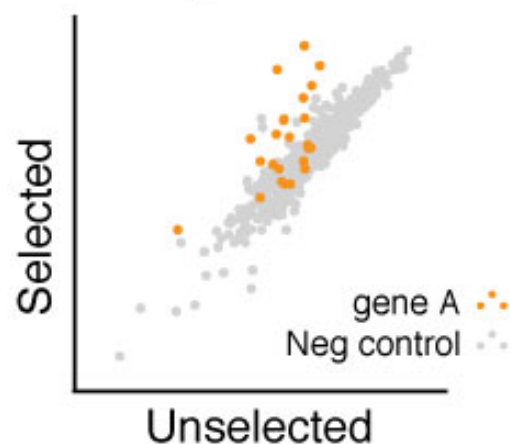
Pooled screening



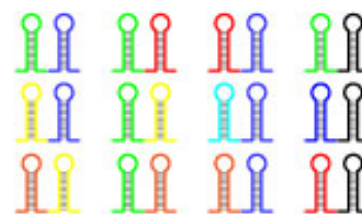
Count sh/sgRNAs
by deep sequencing



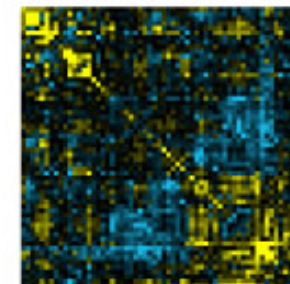
Primary Hit Identification



- Select Active sh/sgRNAs
- Ligate to generate double library



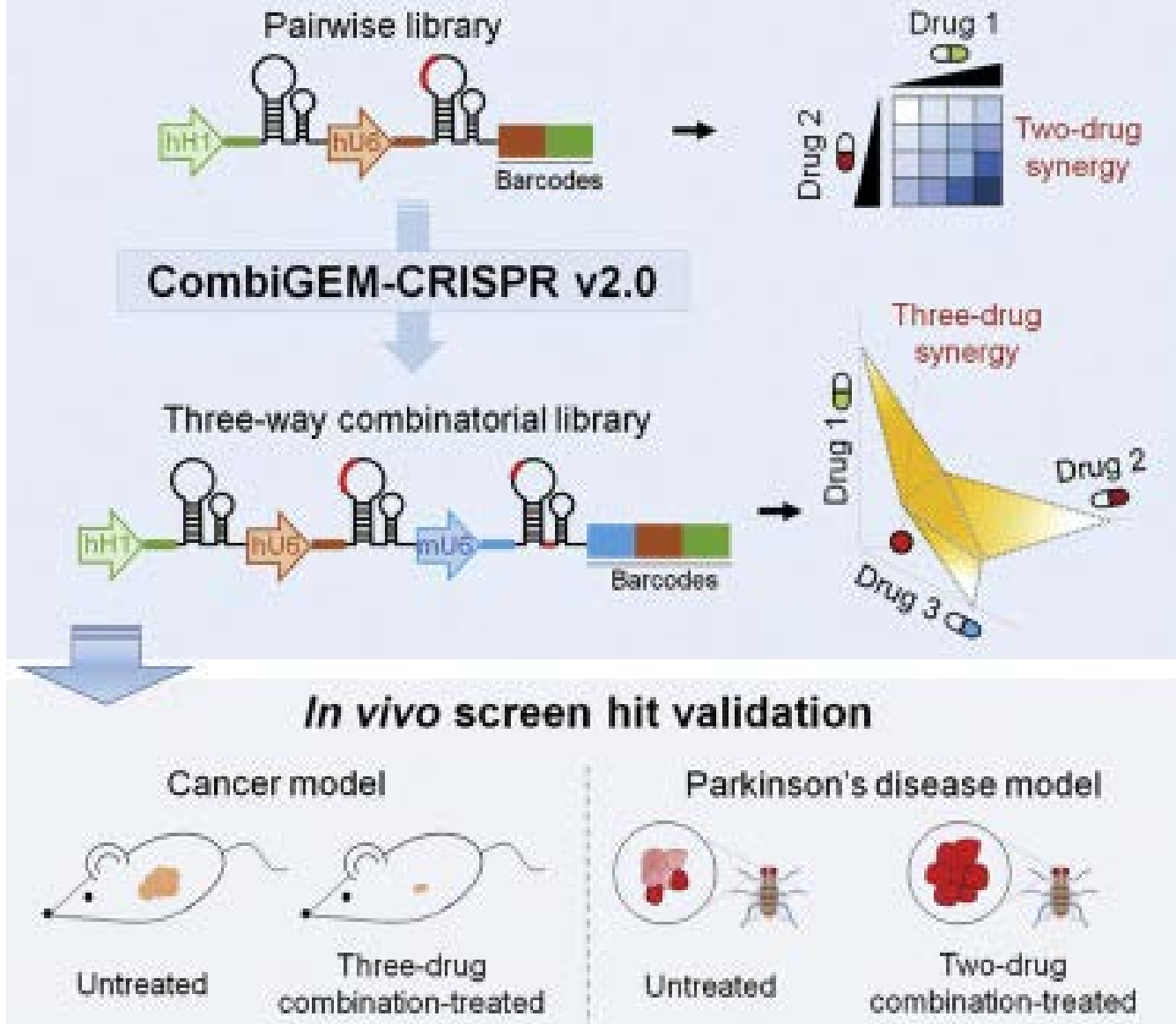
Genetic
Interaction Map



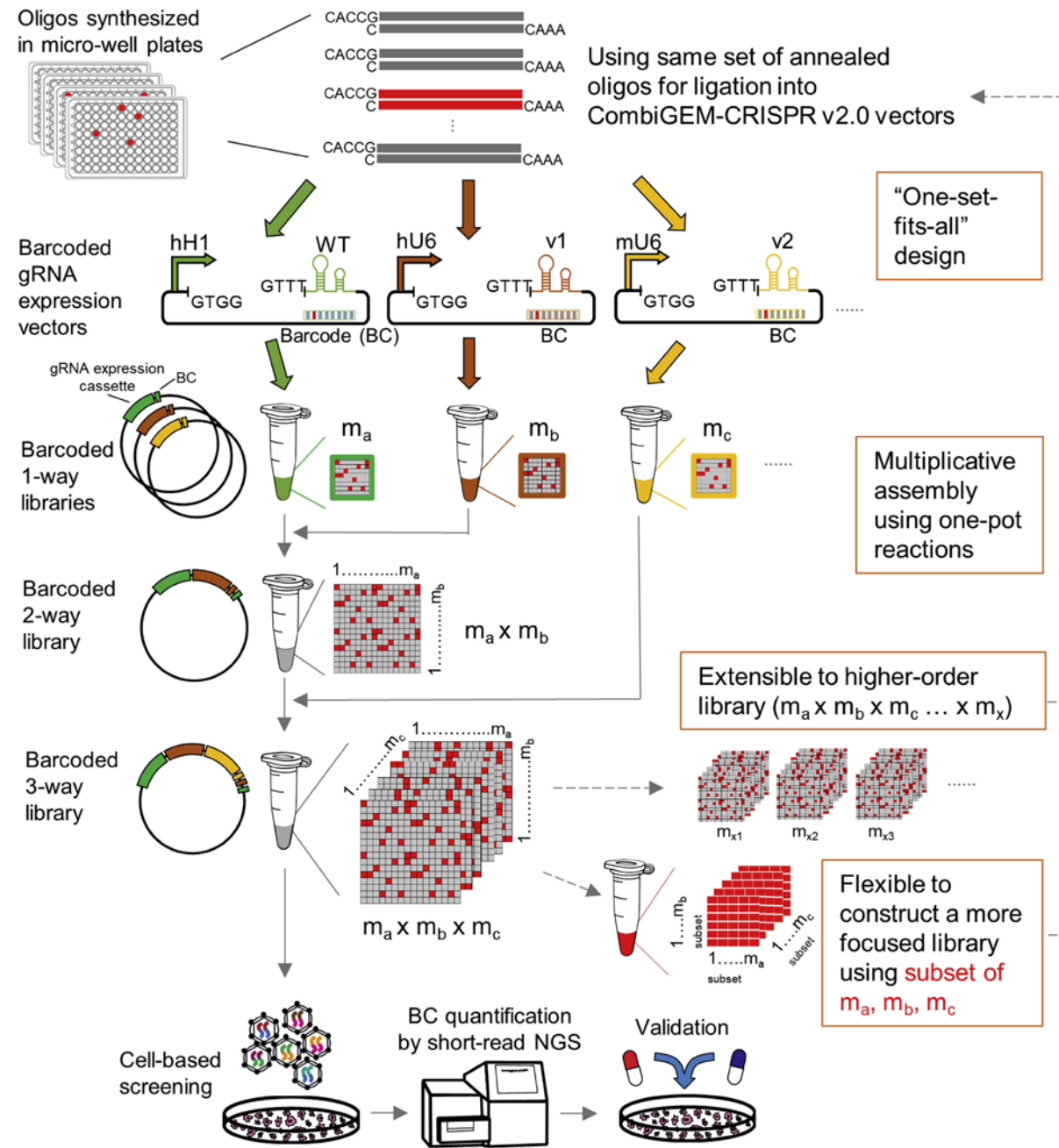
Buffering
Synergistic

A vertical color scale bar with a gradient from blue at the bottom to yellow at the top, with a black band in the middle.

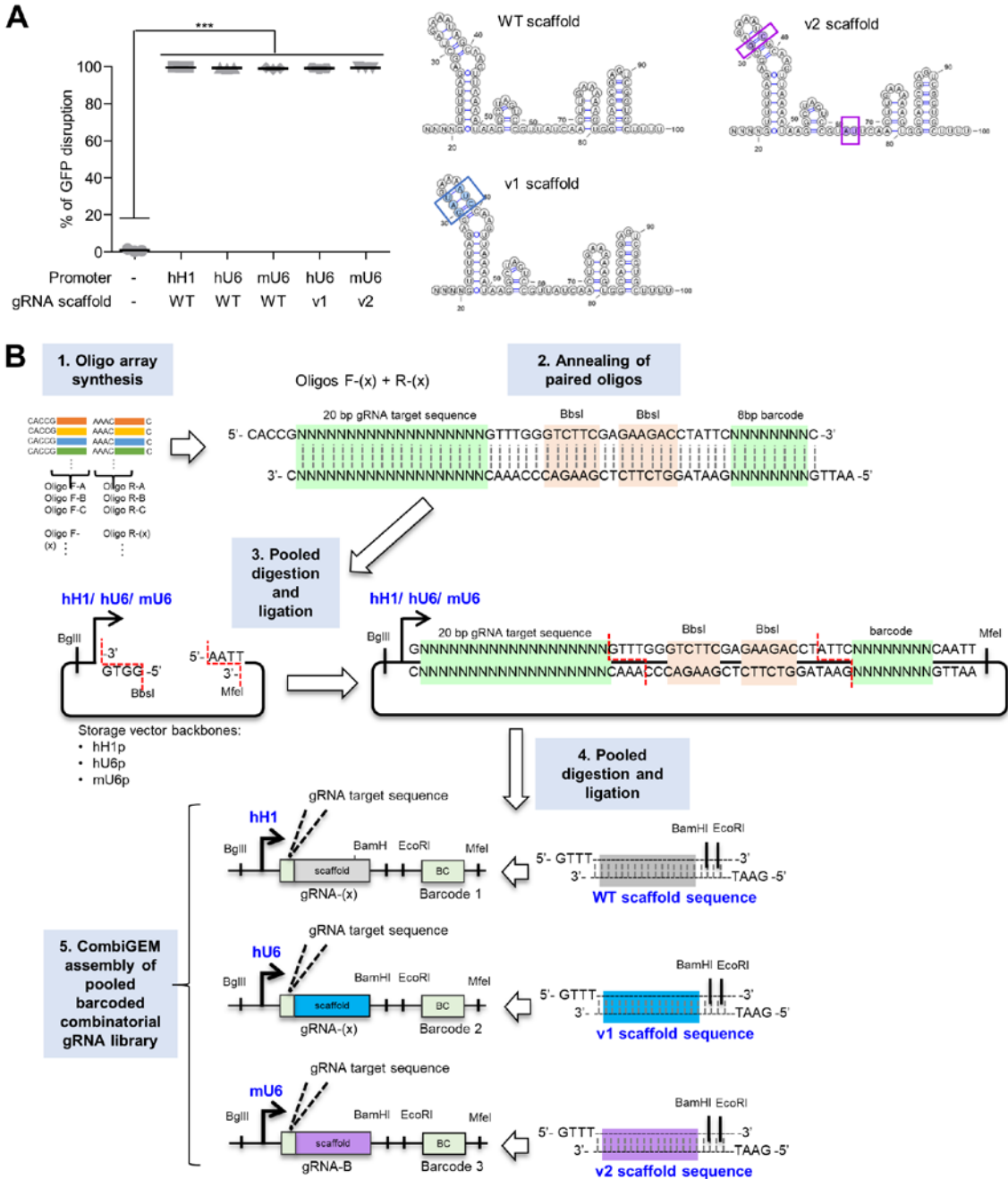
Extensible assembly of gRNA combinations for CRISPR screens to discover drug synergy



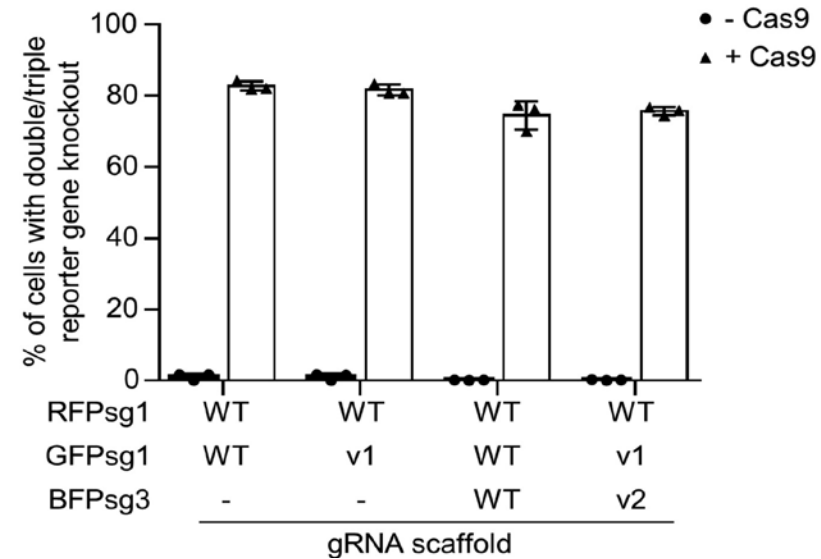
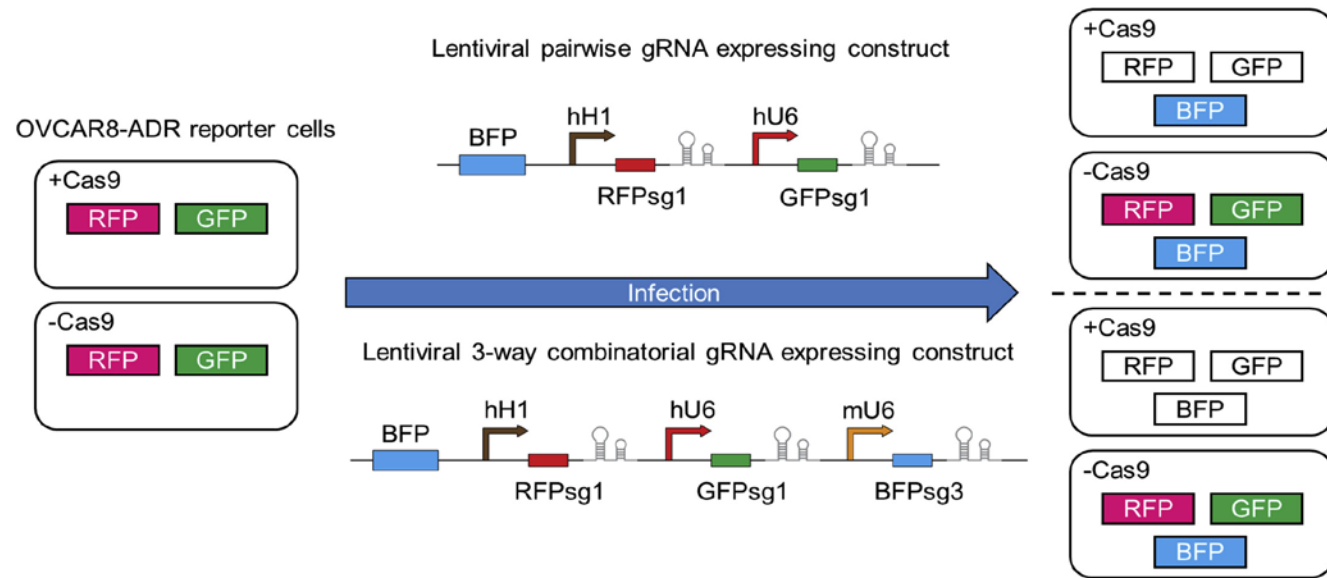
Screen Outlay



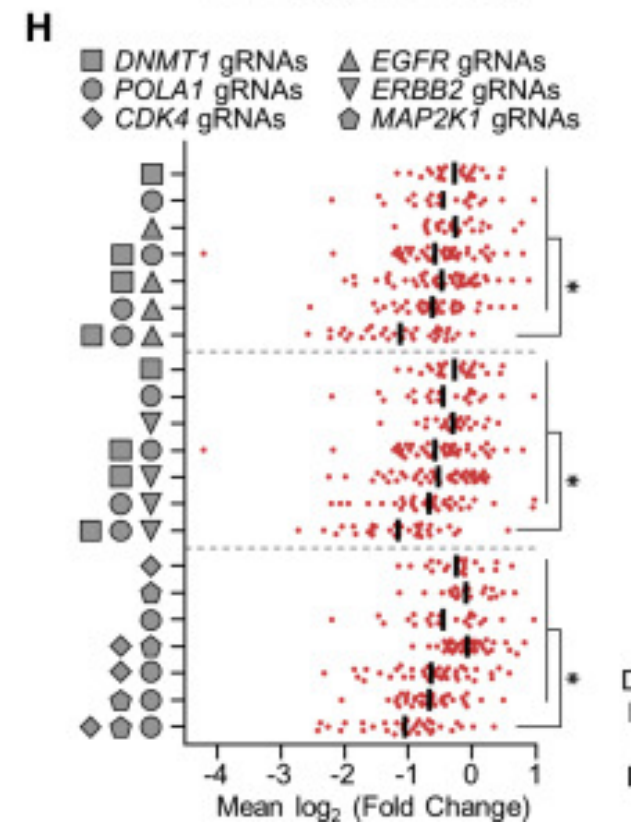
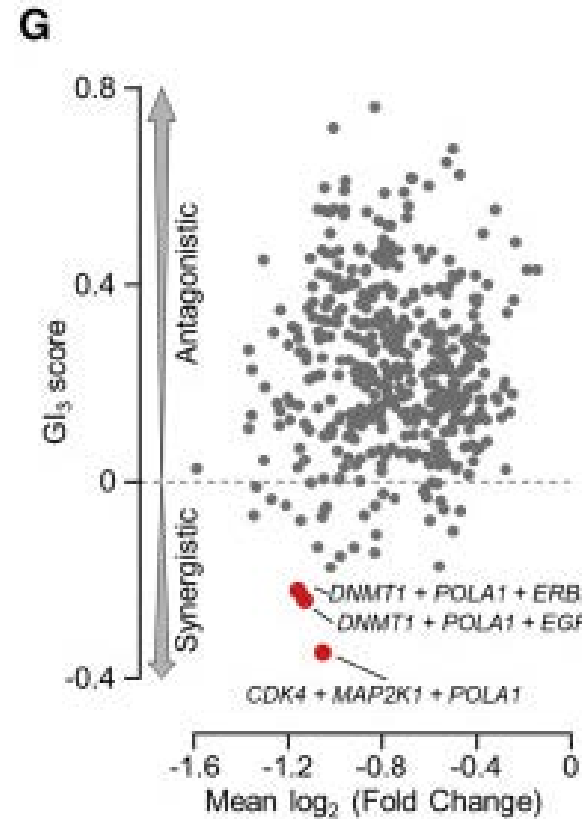
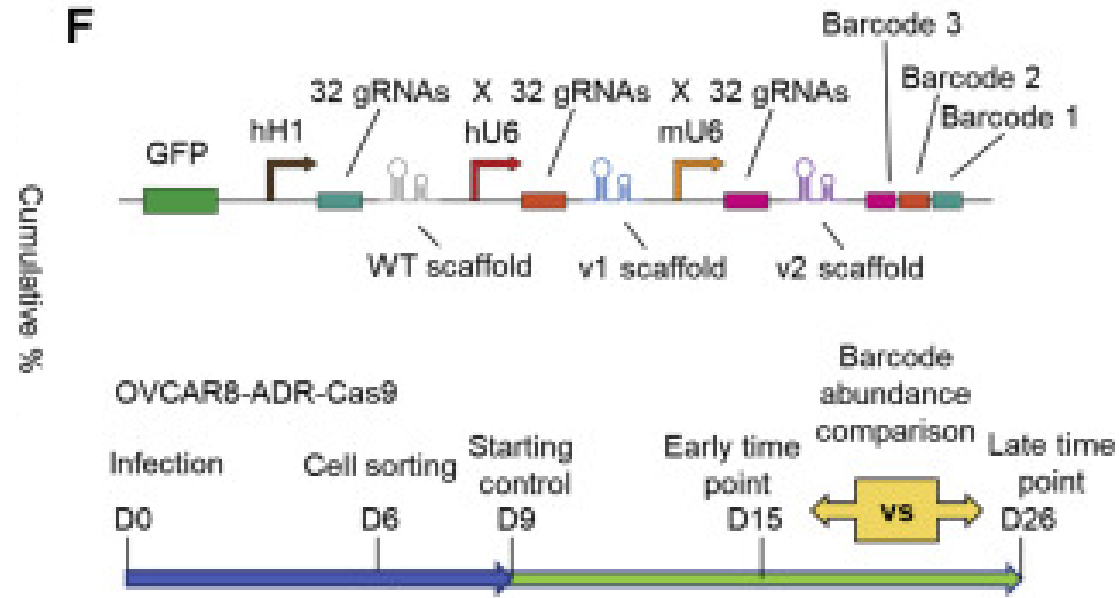
Cloning strategy



Proof of principle: Multiple gene knock out



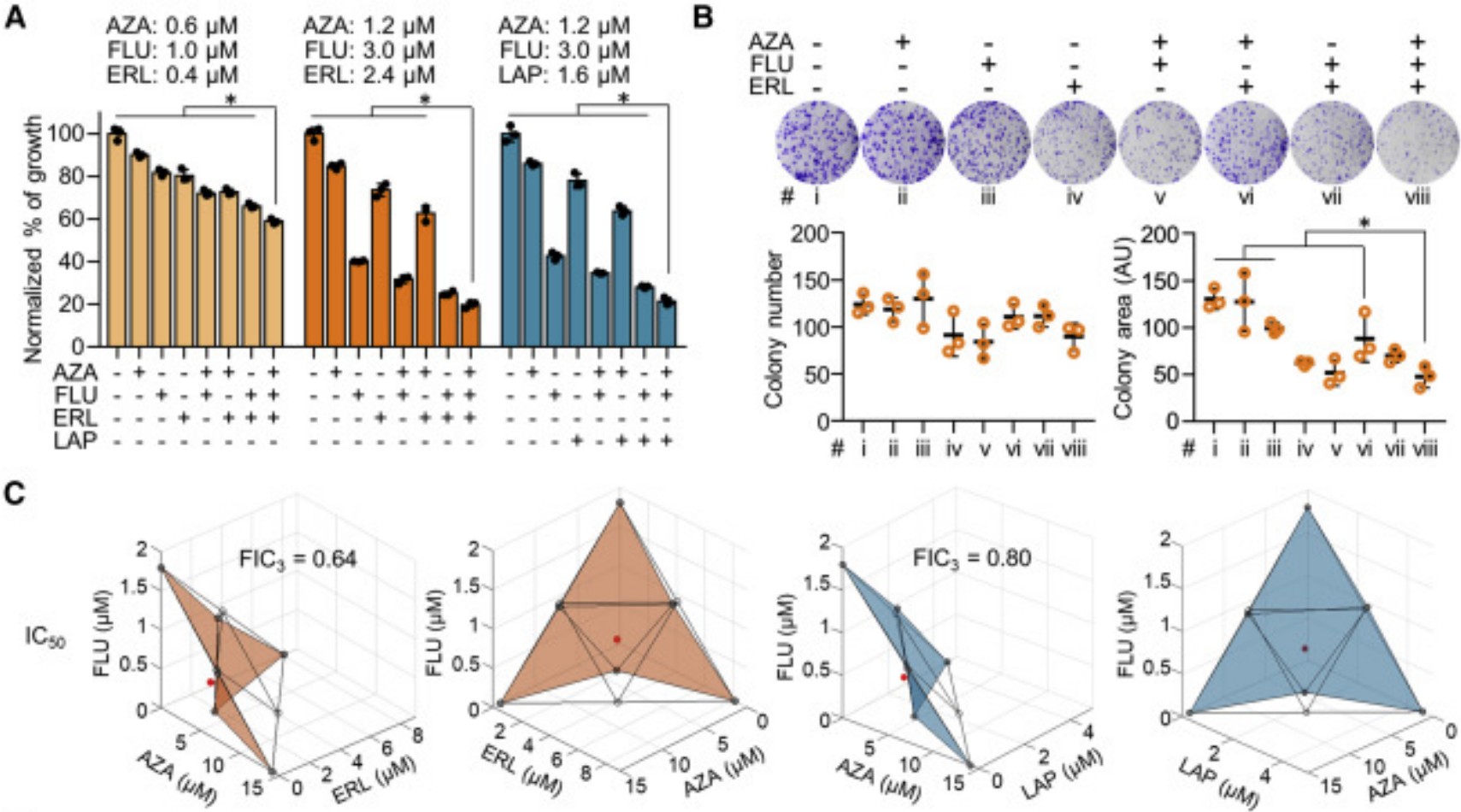
Triple gene knock out-Ovarian cancer



15 druggable genes were chosen

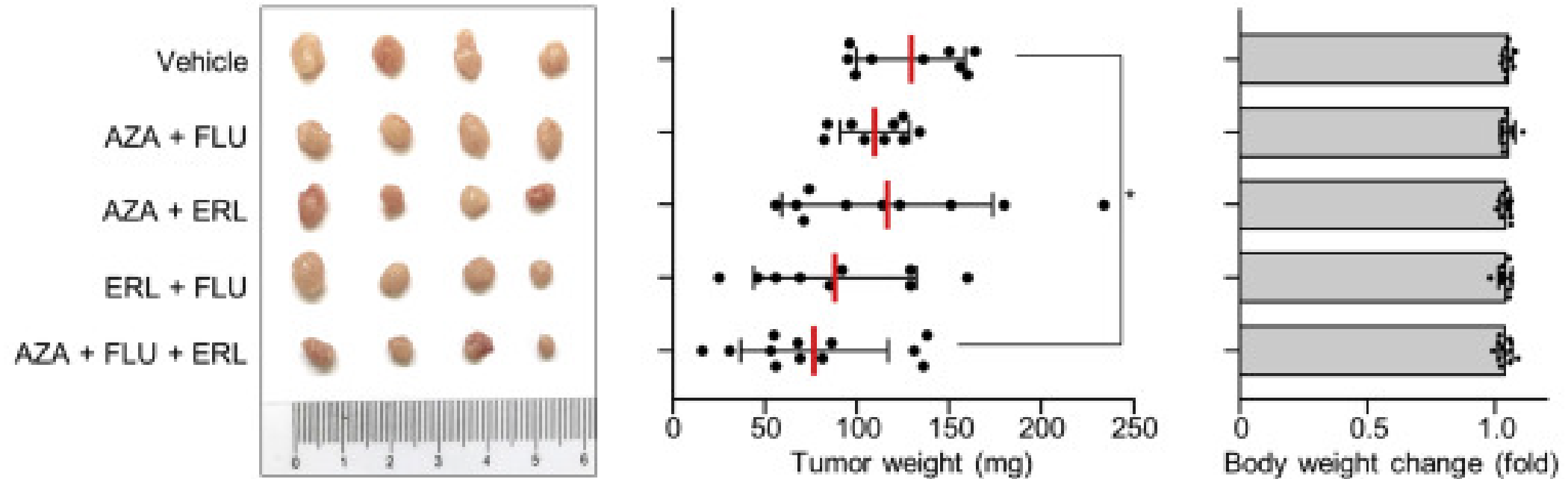
32x32x32gRNA were used

Validation of the hits in the screens using pharmacological inhibitors



New combination therapy of drugs alters the tumor size in vivo

D



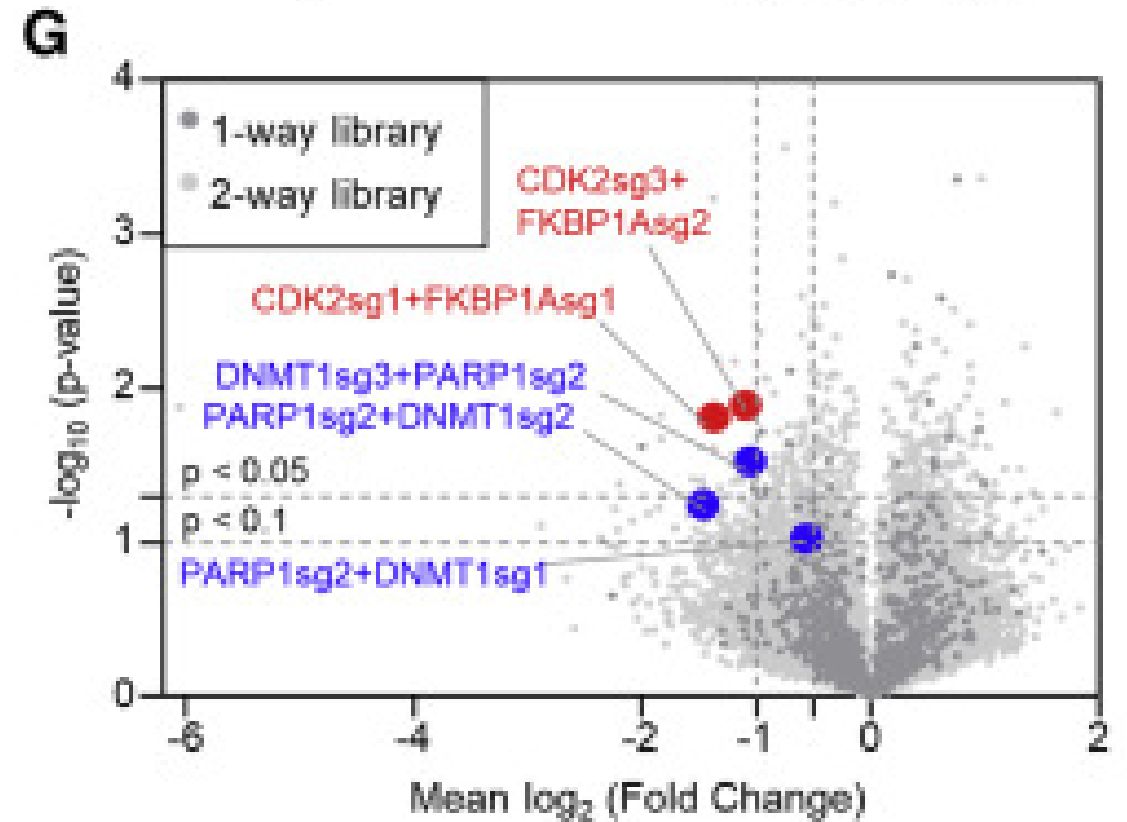
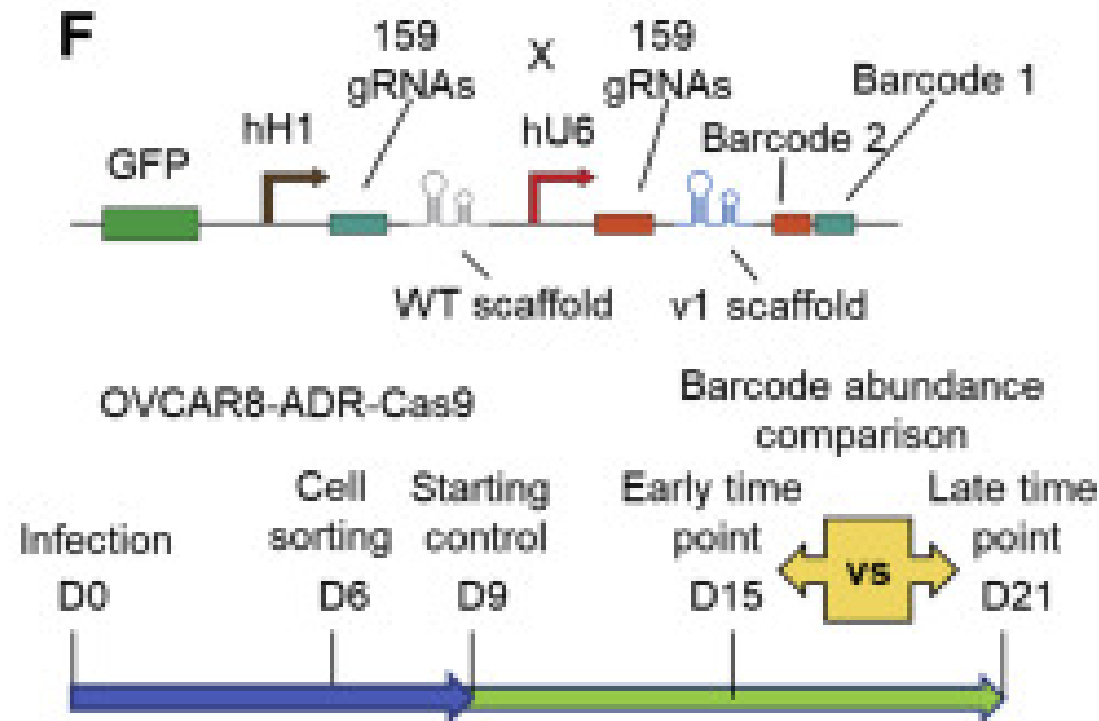
Double gene knock out-Ovarian cancer

10/29/2016 10:00:00 AM

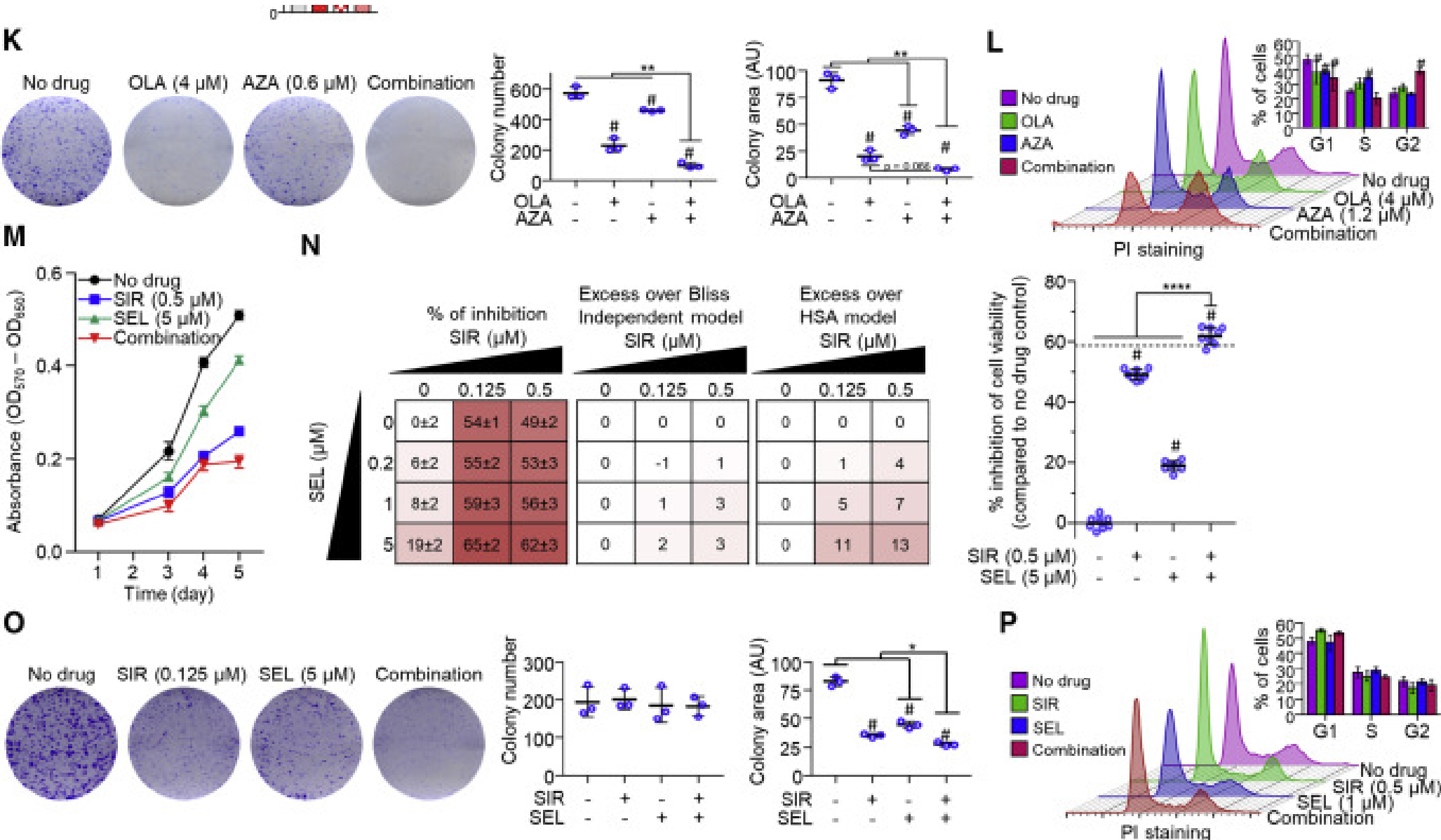
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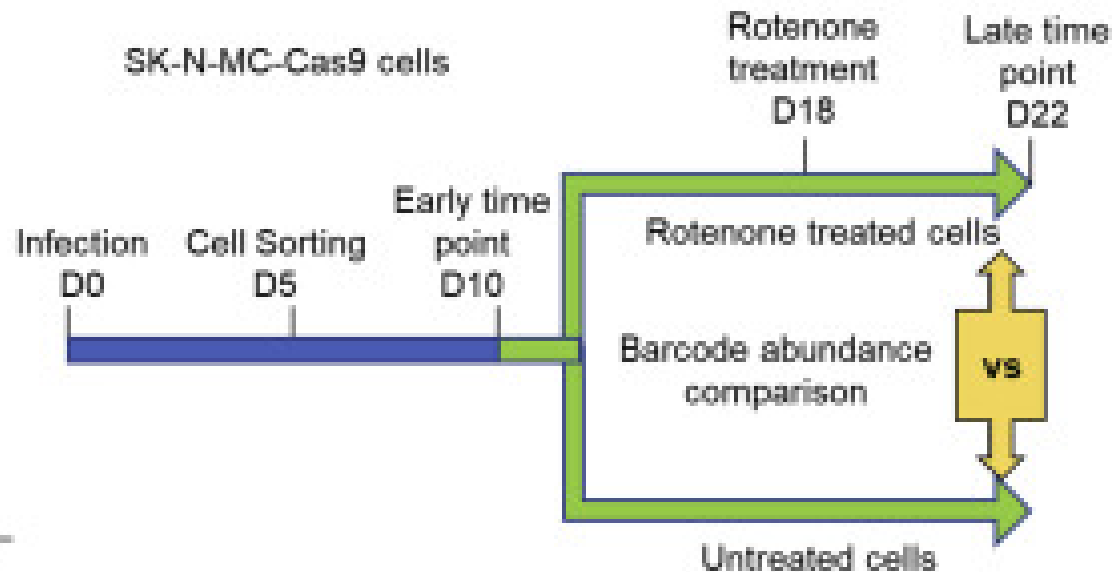


Validation of the hits in the screens using pharmacological inhibitors

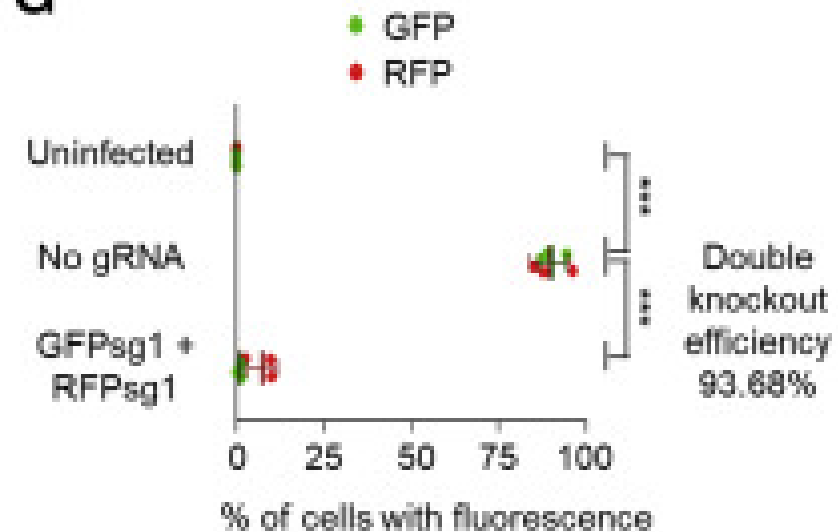


Dual-Gene Knockout Screen Identifies a Drug Combination that Enhances Protection against PD Toxicity

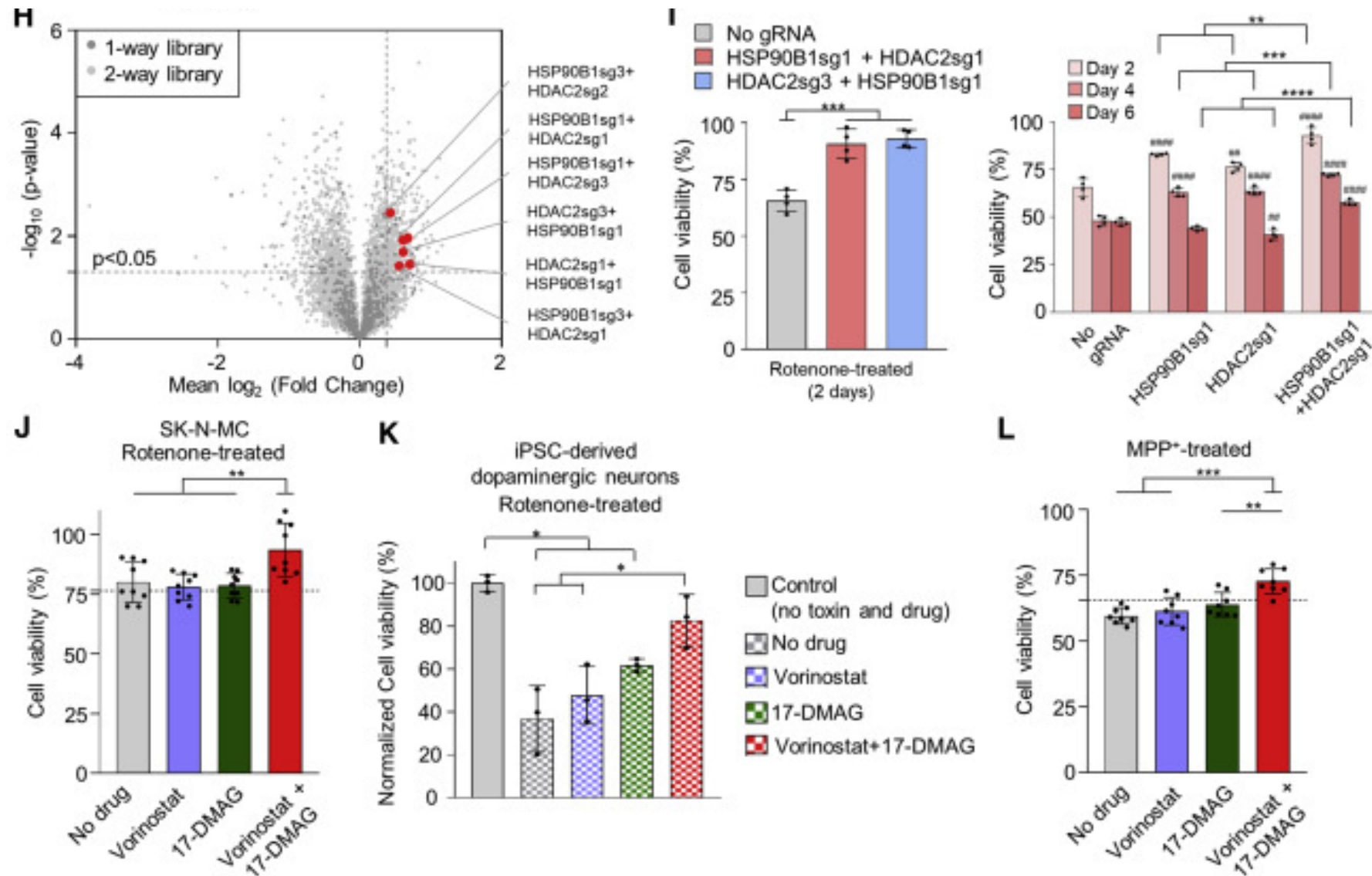
F



G



Validation of the Hits



Conclusions

1. A nice strategy to identify novel combination of genes to identify new therapeutic targets.
2. Platform can be combined with CRISPRi to mimic drugs.
3. Platform can be combined with single cell RNA seq.

Questions:

1. Patient derived cell lines?
2. Efficiency of the guide RNAs

