

***getting into it***

—

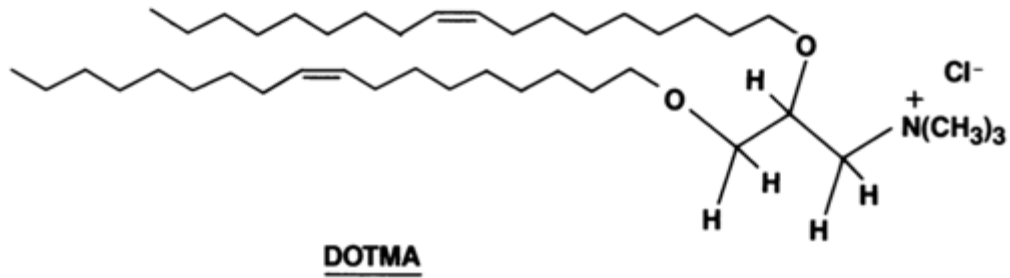
**new technologies for intracellular  
cargo delivery**

**TECHNICAL JOURNAL CLUB 19TH MAY 2015**

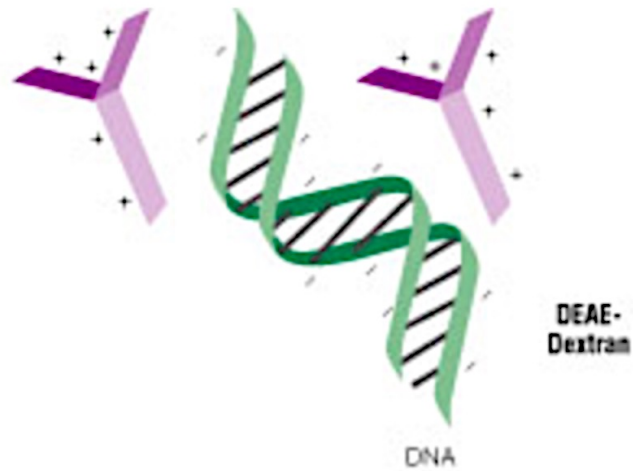
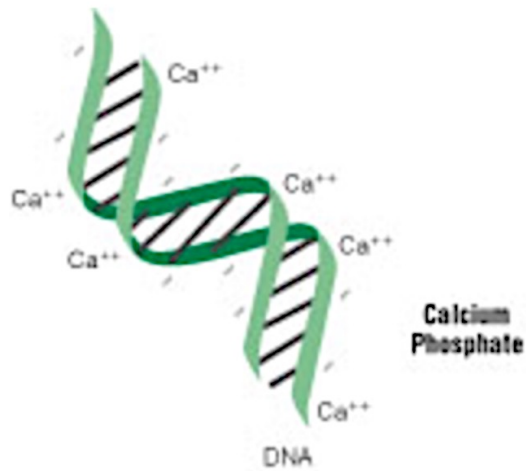
**KARL FRONTZEK**

**INSTITUTE OF NEUROPATHOLOGY**

# Intracellular delivery



L  
i  
p

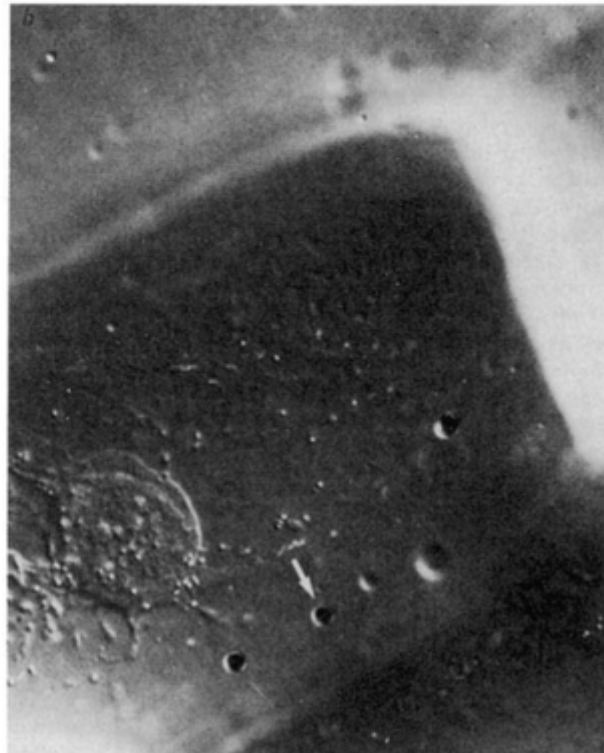
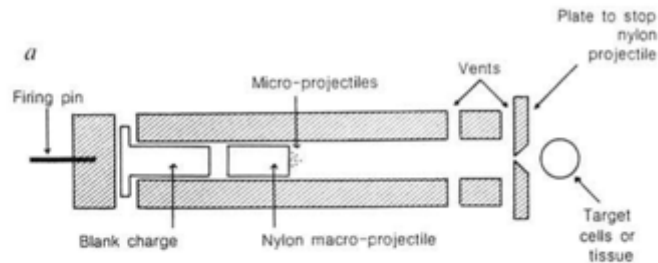


f  
s  
h  
n  
t  
O

# Intracellular delivery

NATURE VOL. 327 7 MAY 1987

LETTERS TO NATURE



npg © 1998 Nature Publishing Group <http://www.nature.com/naturebiotechnology>

RESEARCH

## Gene transfer into muscle by electroporation in vivo

Hiroyuki Aihara<sup>1,2</sup> and Jun-ichi Miyazaki<sup>1\*</sup>

### Brief Communications

*Nature* **418**, 290-291 (18 July 2002) | doi:10.1038/418290a

### Cell biology: Targeted transfection by femtosecond laser

Uday K. Tirlapur & Karsten König

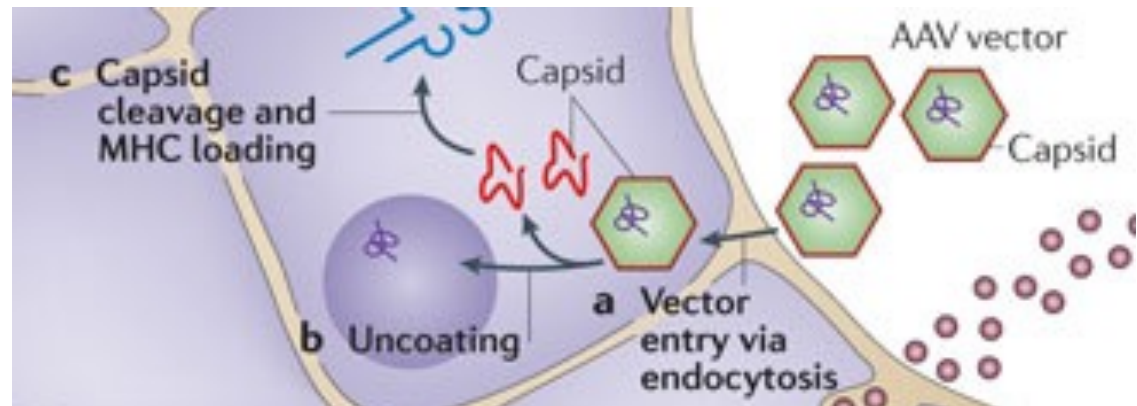
## Gene guns

TM Klein et al., *Nature* 1987

# Intracellular delivery



**Lentiviral packaging**  
[www.clontech.com](http://www.clontech.com)



**Adeno-associated virus packaging**  
Mingozi F & High KA, Nat Rev Genetics 2011

# Intracellular delivery of native proteins/DNA

*„in vitro“ surgery*

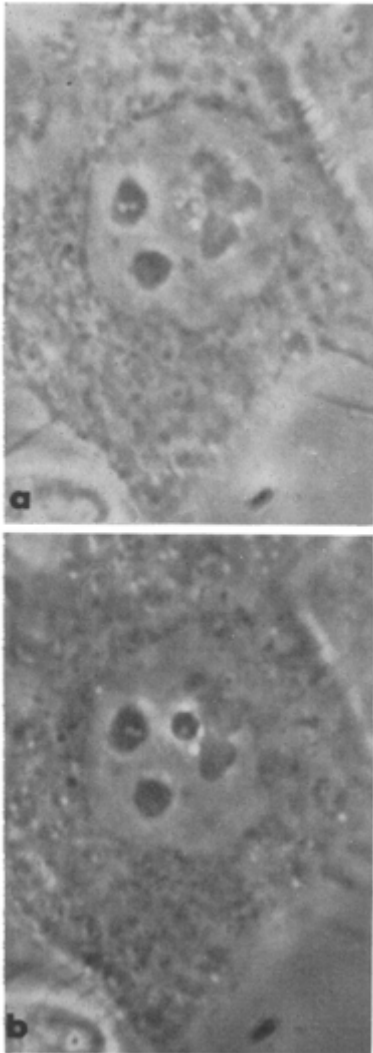
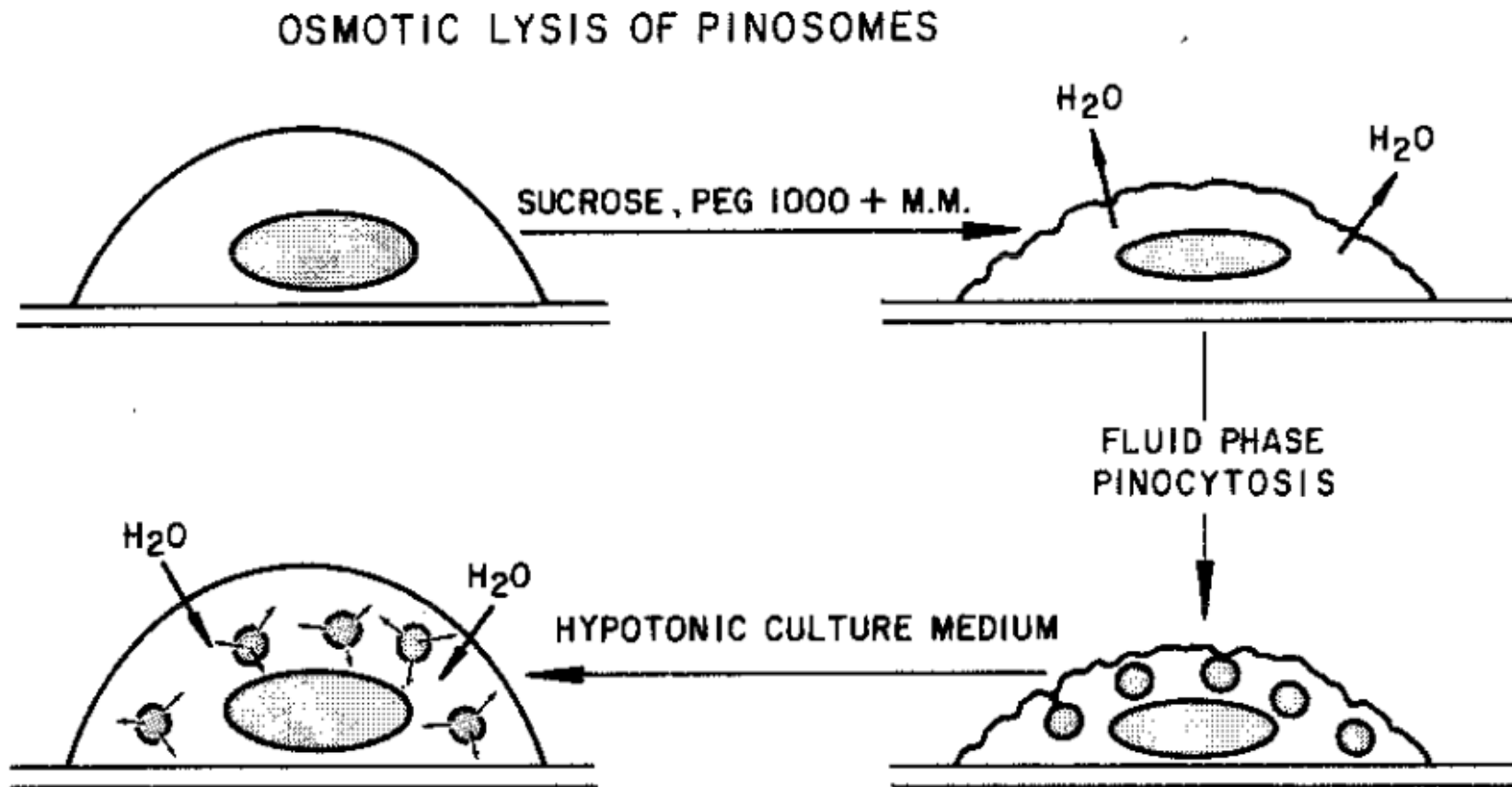


FIG. 1.

Diacumos EG et al., PNAS 1970

# Intracellular delivery of native compounds

*„osmotic lysis of pinocytic vesicles“*

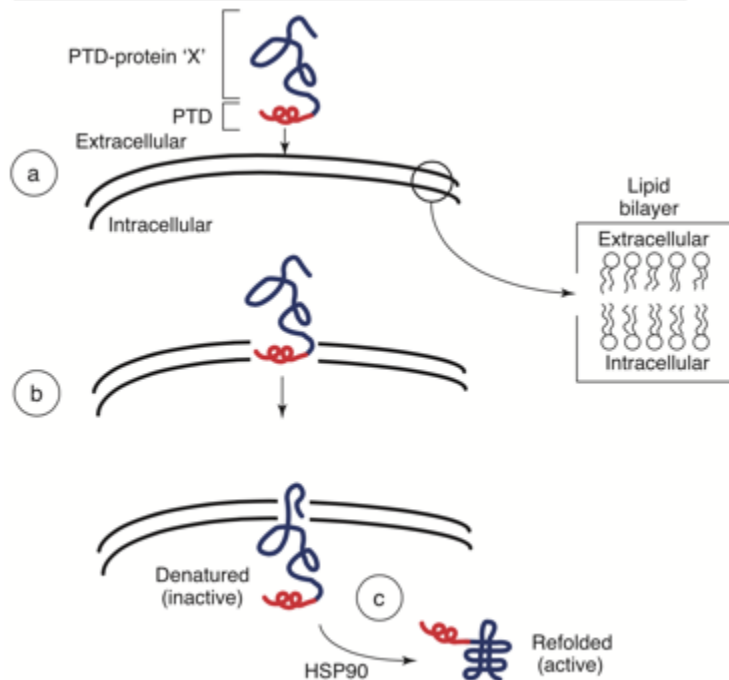


# Intracellular delivery with fusion proteins carrying „protein-transduction domains“

TABLE 1 – AMINO ACID SEQUENCE OF CHARACTERIZED PTDs

PTD	Amino acid sequence
HIV-1 TAT	Tyr-Gly-Arg-Lys-Lys-Arg-Arg-Gln-Arg-Arg-Arg <sup>a</sup>
HSV VP22	Asp-Ala-Ala-Thr-Ala-Thr-Arg-Gly-Arg-Ser-Ala-Ala-Ser-Arg-Pro-Thr-Glu-Arg-Pro-Arg-Ala-Pro-Ala-Arg-Ser-Ala-Ser-Arg-Pro-Arg-Arg-Pro-Val-Glu
Antp	Arg-Gln-Iso-Lys-Iso-Trp-Phe-Gln-Asn-Arg-Arg-Met-Lys-Trp-Lys-Lys

<sup>a</sup>Gly-Arg-Lys-Lys-Arg-Arg is a potential nuclear localization sequence.  
Abbreviation: PTD, protein-transduction domain.



trends in Cell Biology

## Correspondence

*Nature Biotechnology* **19**, 713 (2001)  
doi:10.1038/90741

## Is VP22 nuclear homing an artifact?

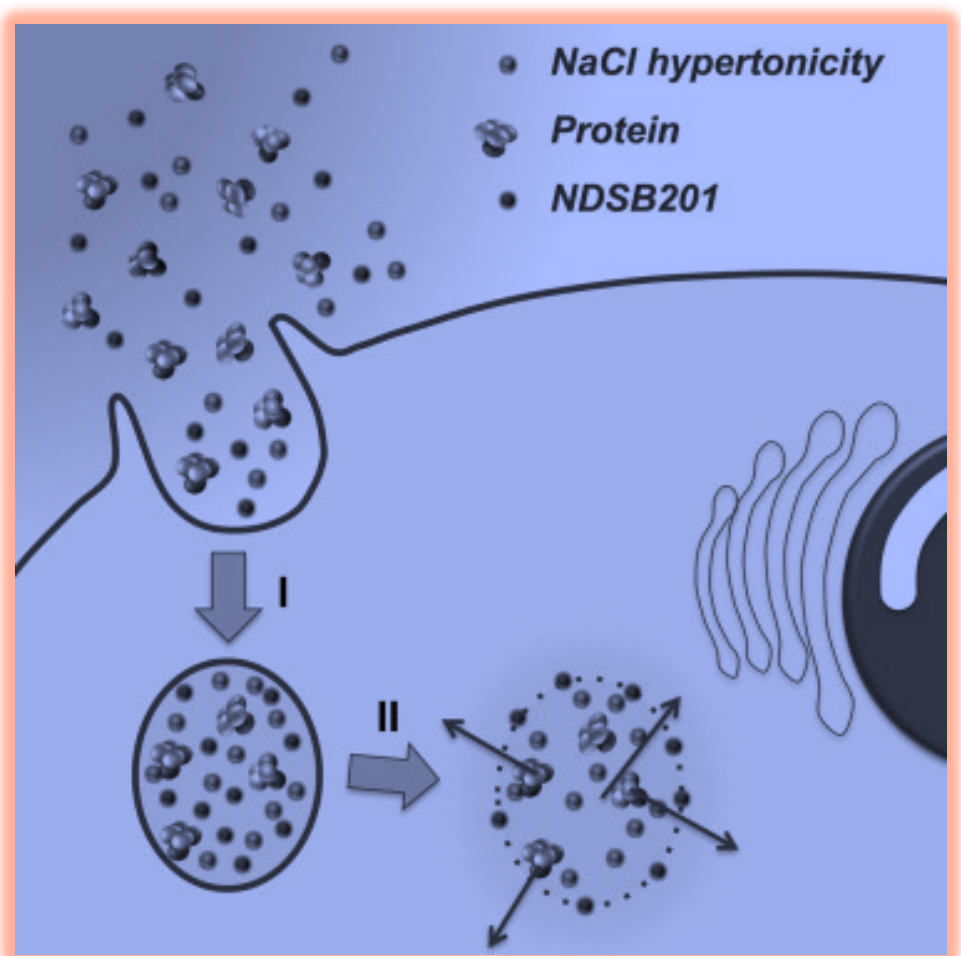
Mathias Lundberg<sup>1</sup> & Magnus Johansson<sup>2</sup>

1. Department of Medical Biochemistry and Biophysics, Karolinska Institute, S-17177 Stockholm, Sweden
2. Division of Clinical Virology F68, Karolinska Institute, Huddinge University Hospital, S-14186 Stockholm, Sweden  
(e-mail: [magnus.johansson@mbb.ki.se](mailto:magnus.johansson@mbb.ki.se)).

**„ITOP“**

**INDUCED TRANSDUCTION BY  
OSMOCYTOSIS AND PROPANEBETAINE**

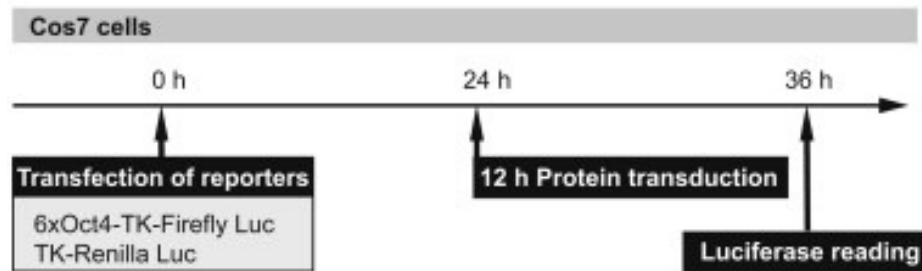
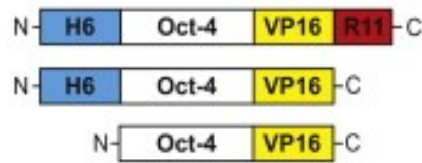
**D'ASTOLFO ET AL., CELL 2015**





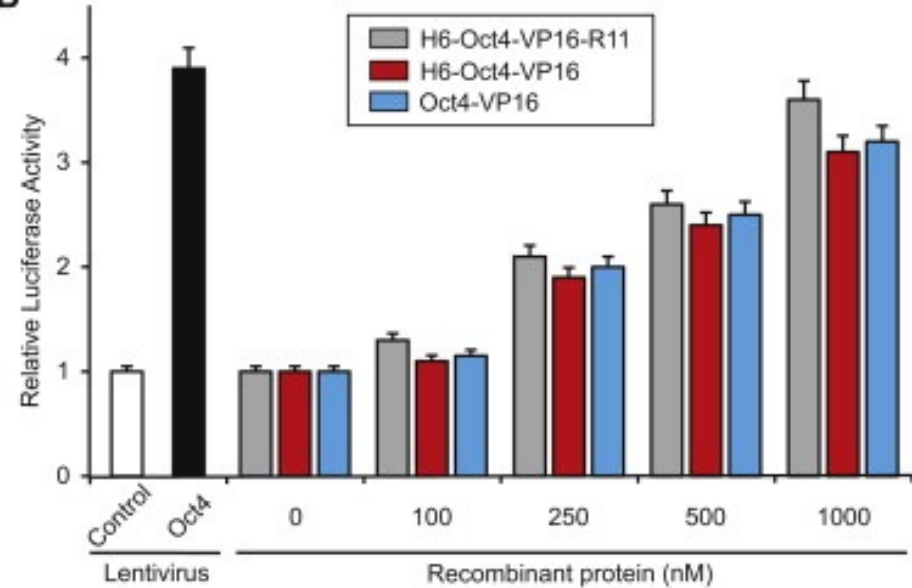
# Transduction of native protein independent of a „cell-penetrating peptide“

A



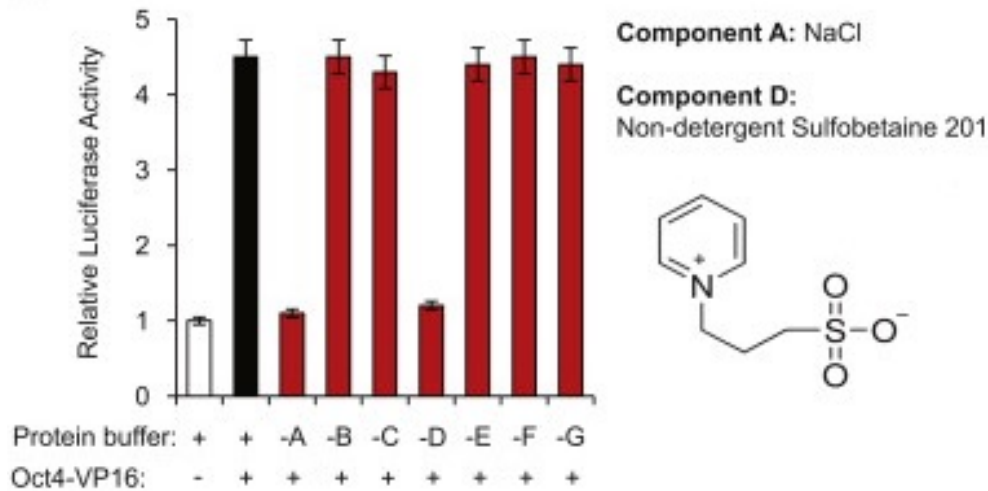
H6 = 6xHis Tag  
R11 = poly-arginine CPP

B

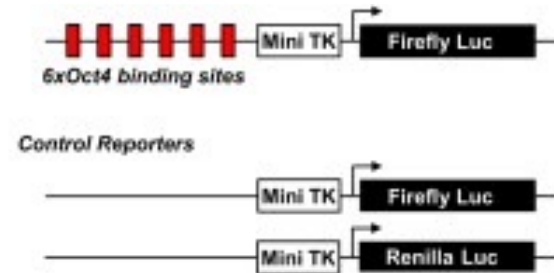


# Omitting NaCl and NDSB-201 from the transduction buffer prevents efficient Oct4 protein delivery

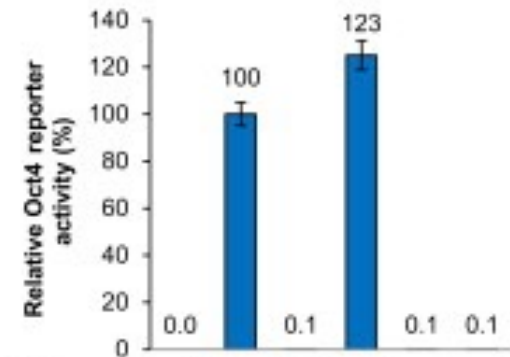
C



A



B



NDSB-201: non-detergent Sulfobetaine 201

A: 1M NaCl

B: 50 mM NaH<sub>2</sub>PO<sub>4</sub>

C: 50 mM Tris-HCl

D: 250 mM NDSB-201

E: 100 μM 2-mercaptoethanol

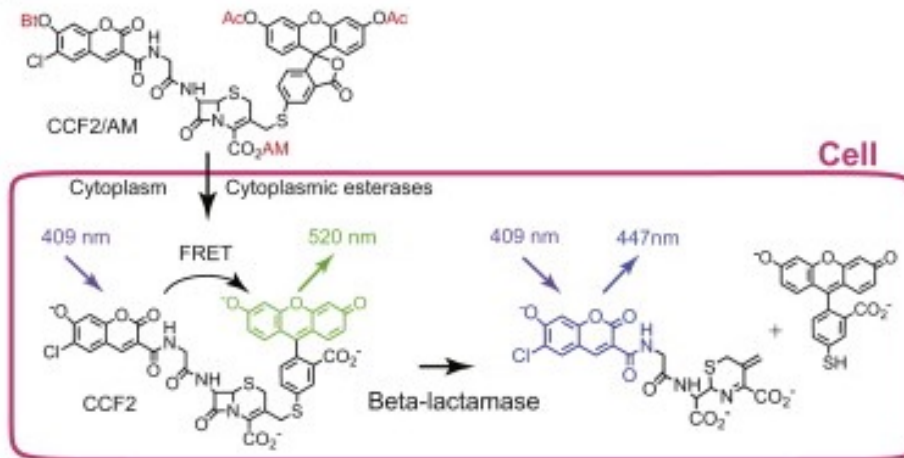
F: 125 μM MgCl<sub>2</sub>

G: 125 μM ZnCl<sub>2</sub>

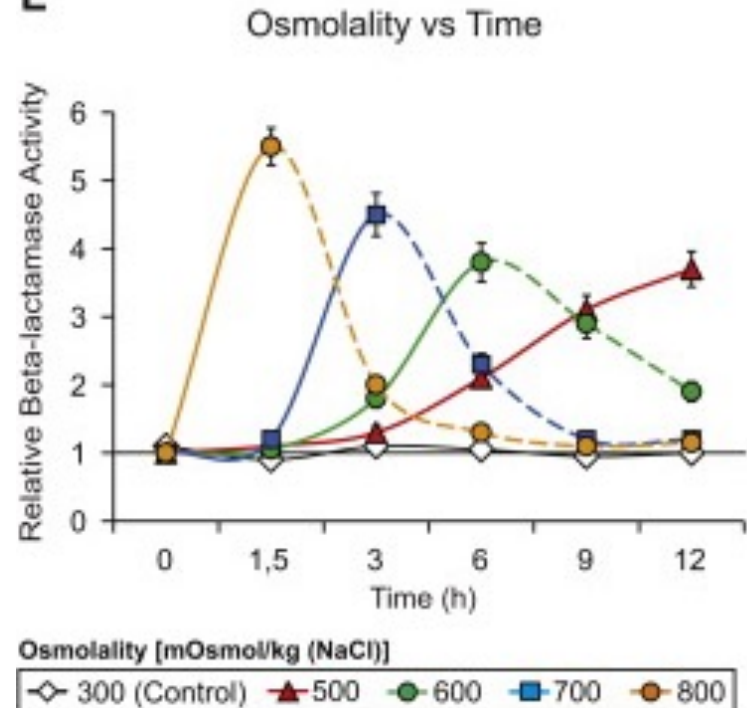
6xOct4-TK-LUC	+	+	+	+		
TK-LUC (Control)					+	+
TK-Renilla	+	+	+	+	+	+
Lenti	+					
Lenti-Oct4		+				
Protein Buffer (Control)			+		+	
Oct4-VP16 (500 nM)				+		+

# Optimal transduction time is directly proportional to NaCl-induced hyperosmolarity

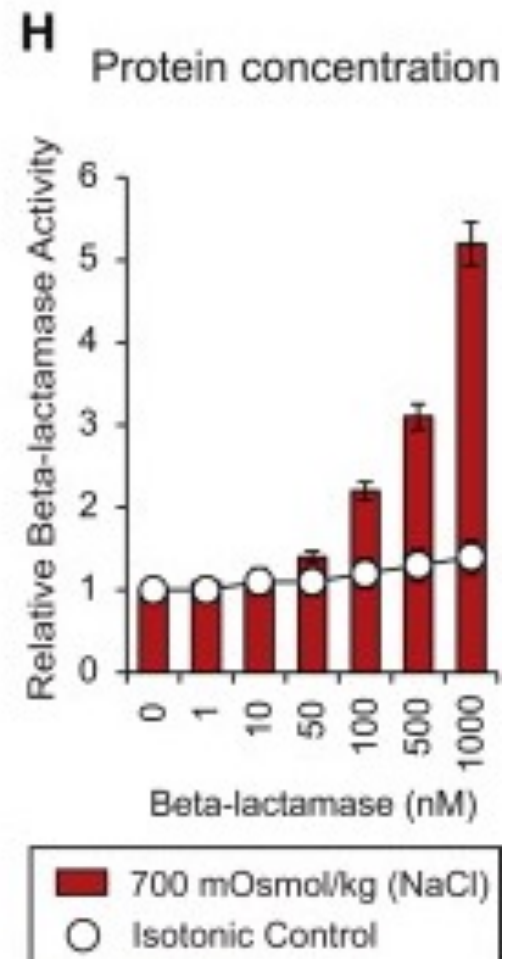
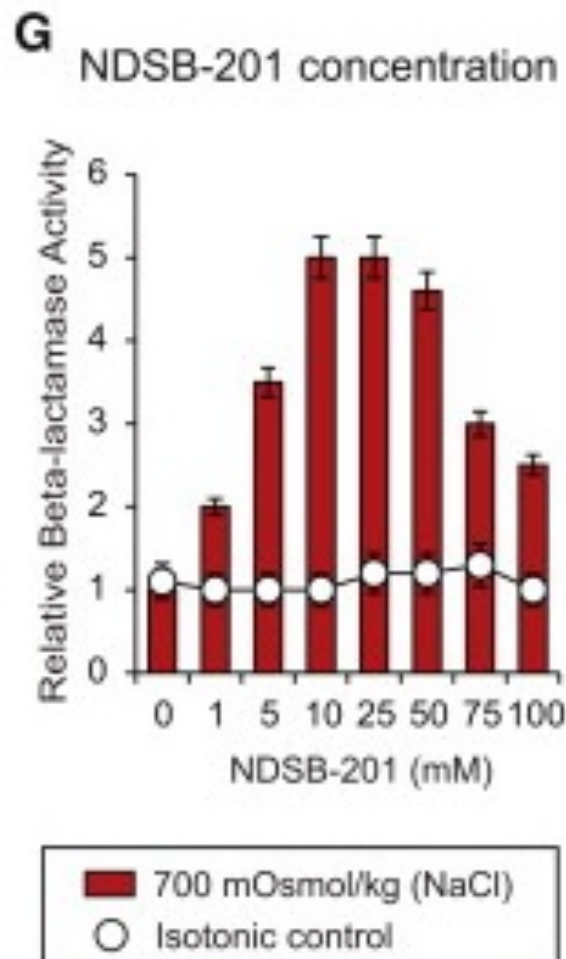
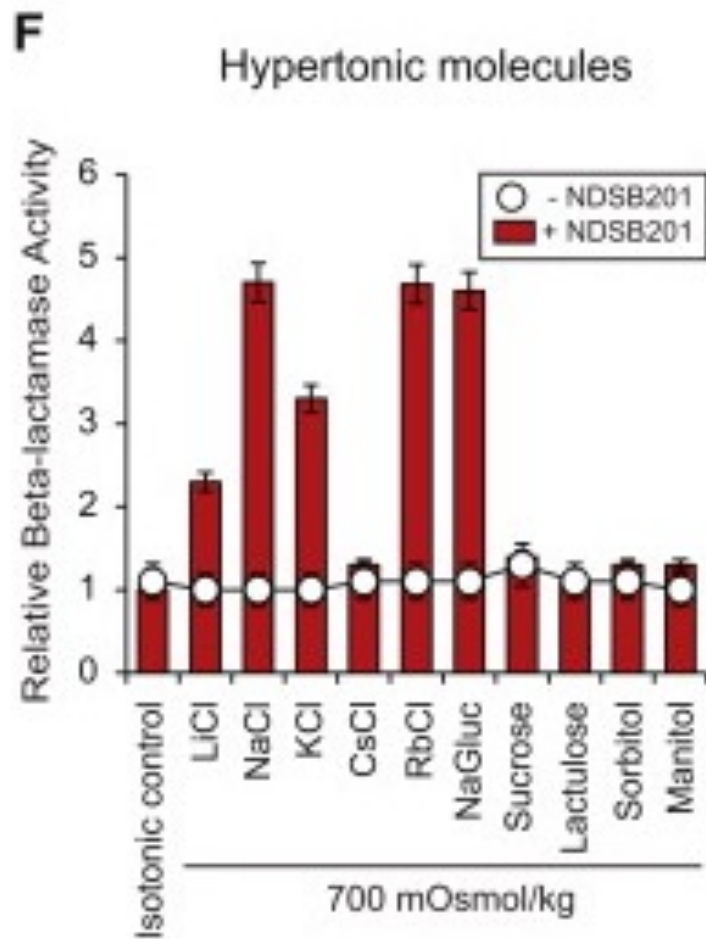
D



E

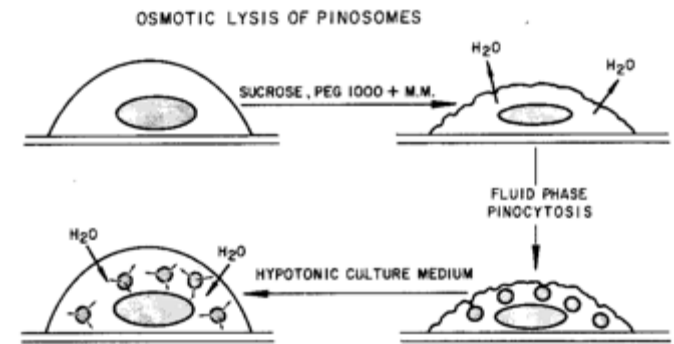


# Cl<sup>-</sup> anions are not essential for induction of hypertonicity

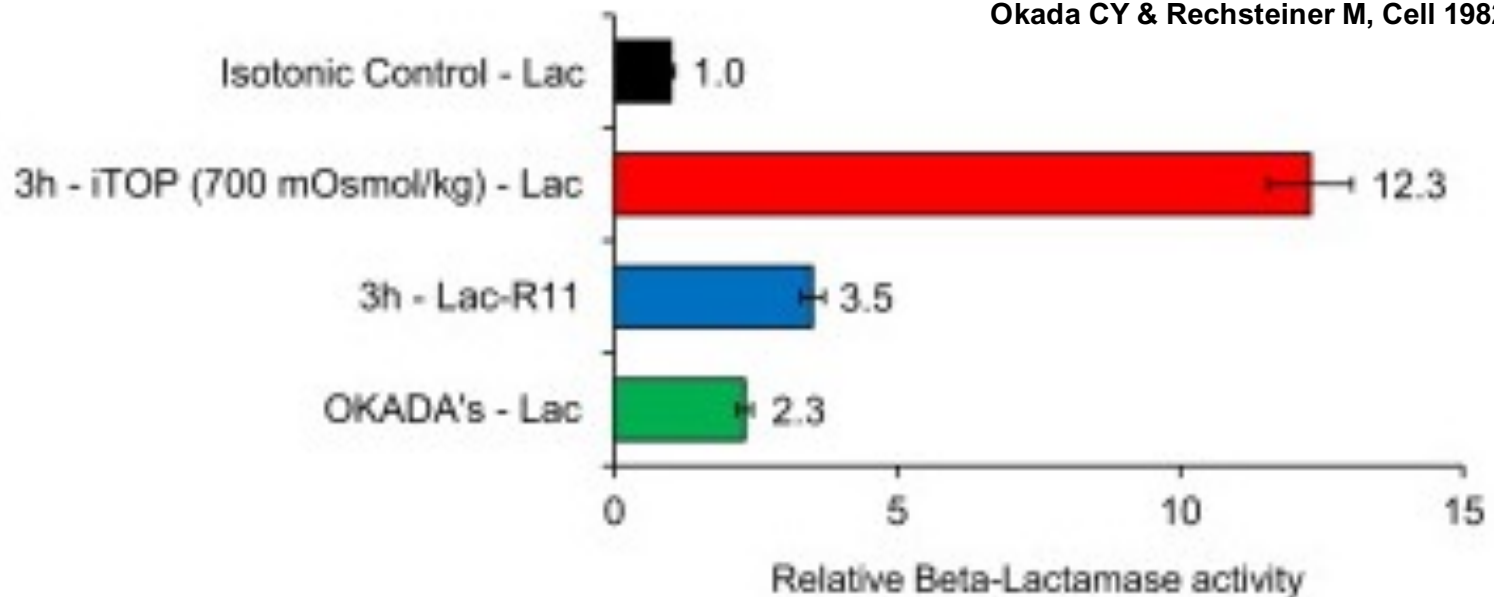


# iTOP vs osmotic lysis of pinocytic vesicles

F



Okada CY & Rechsteiner M, Cell 1982



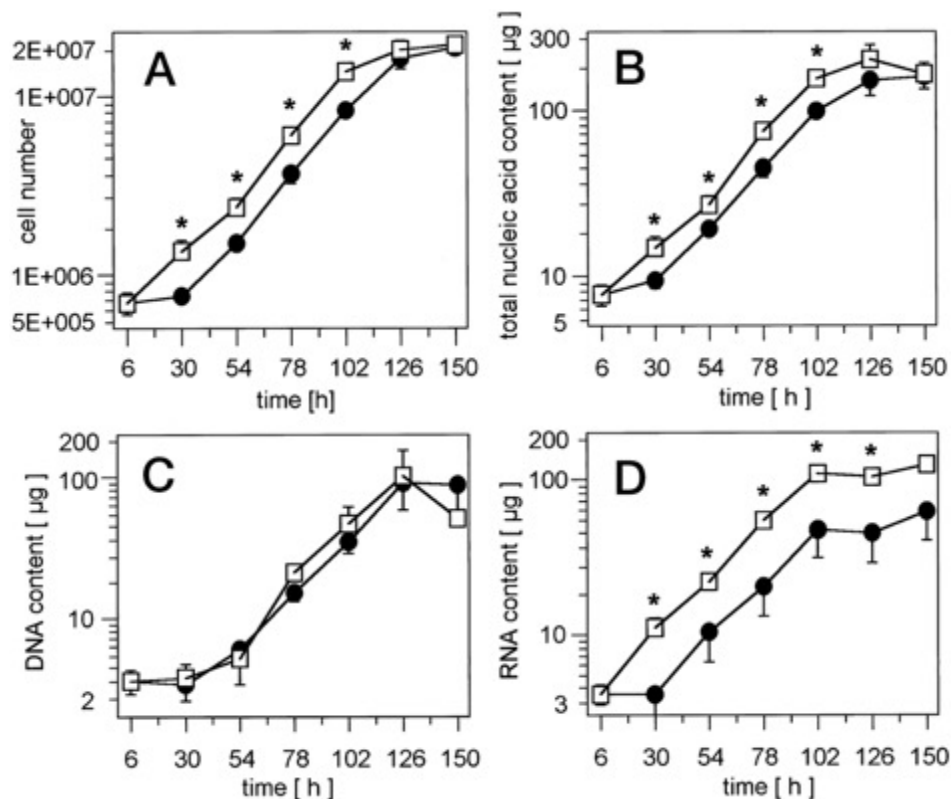
# Hyperosmolality Causes Growth Arrest of Murine Kidney Cells

## INDUCTION OF GADD45 AND GADD153 BY OSMOSENSING VIA STRESS-ACTIVATED PROTEIN KINASE 2\*

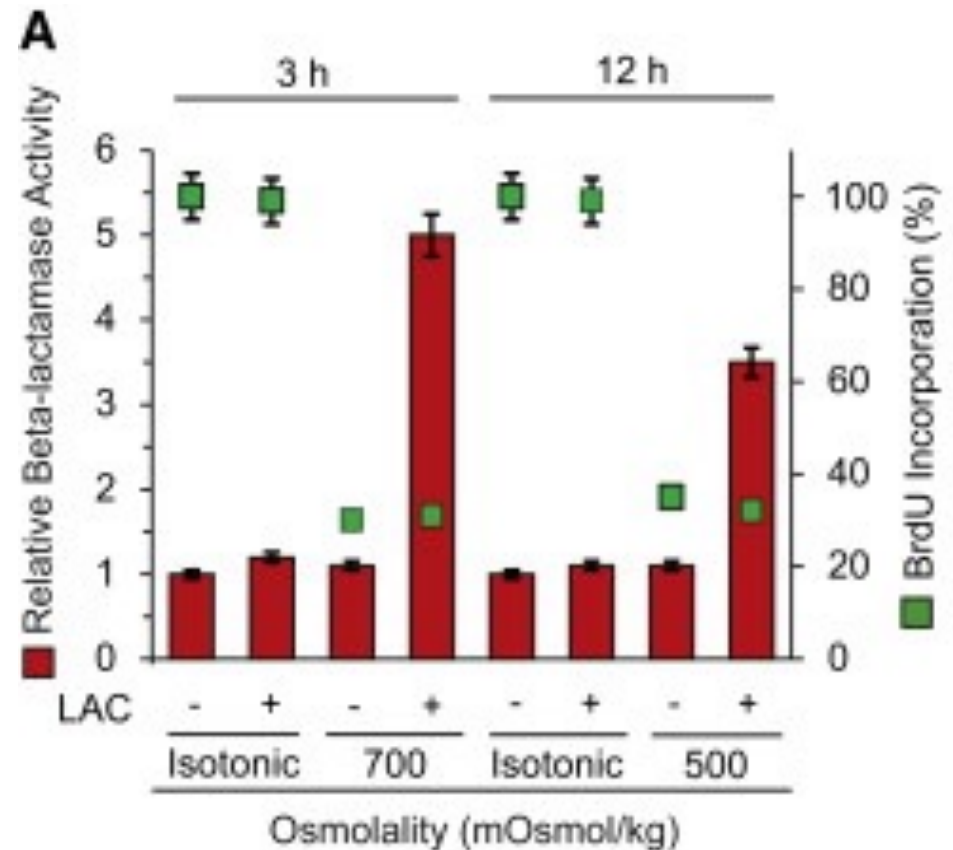
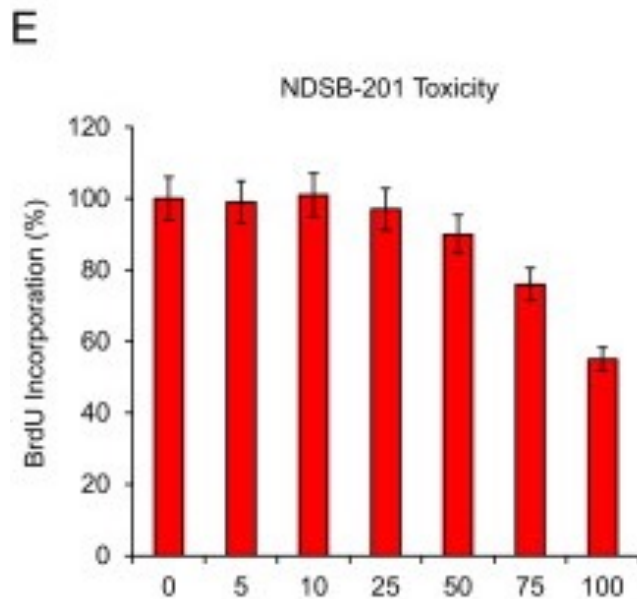
(Received for publication, January 13, 1998, and in revised form, March 16, 1998)

**Dietmar Kültz†, Samira Madhany§, and Maurice B. Burg**

*From the Laboratory of Kidney and Electrolyte Metabolism, NHLBI, National Institutes of Health, Bethesda, Maryland 20892-1603*

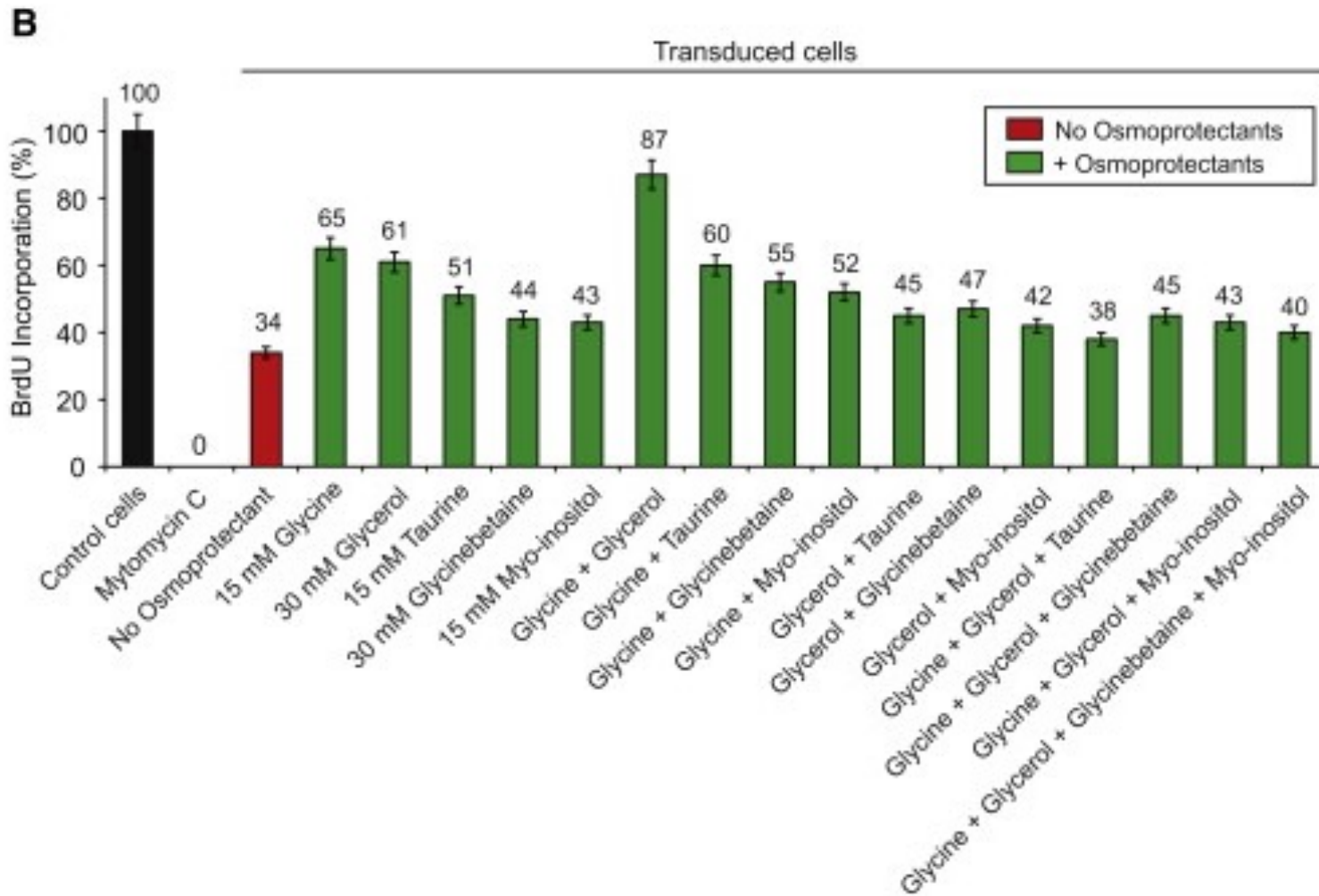


# Hypertonicity induces cell-cycle inhibition



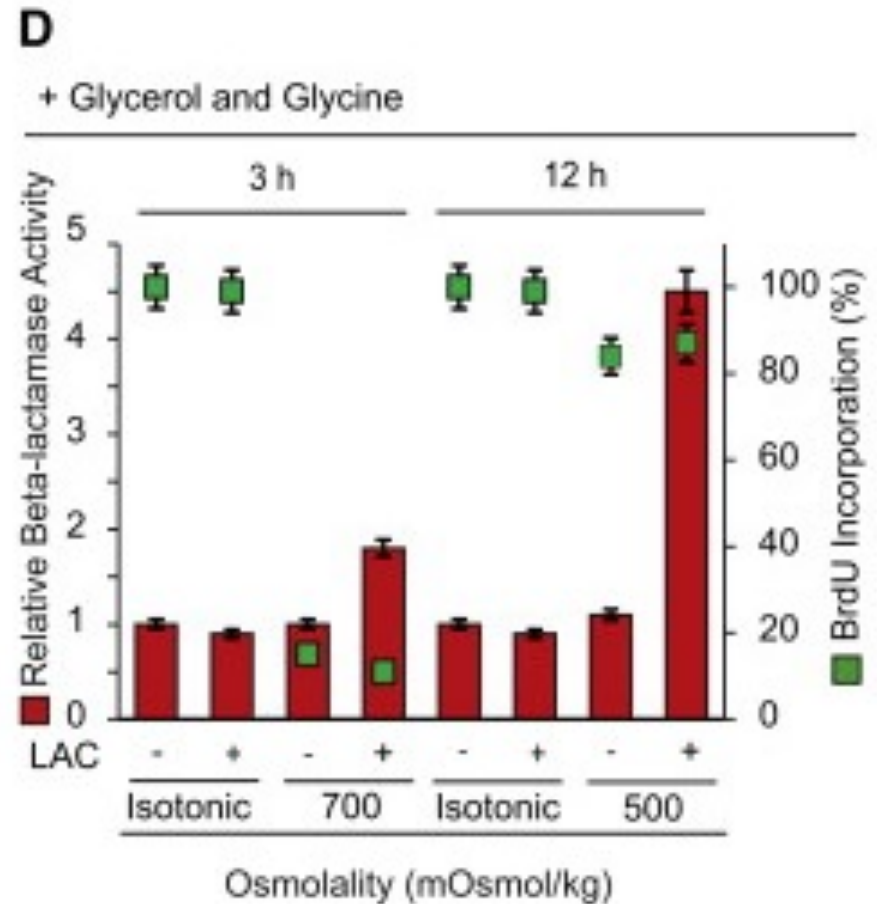
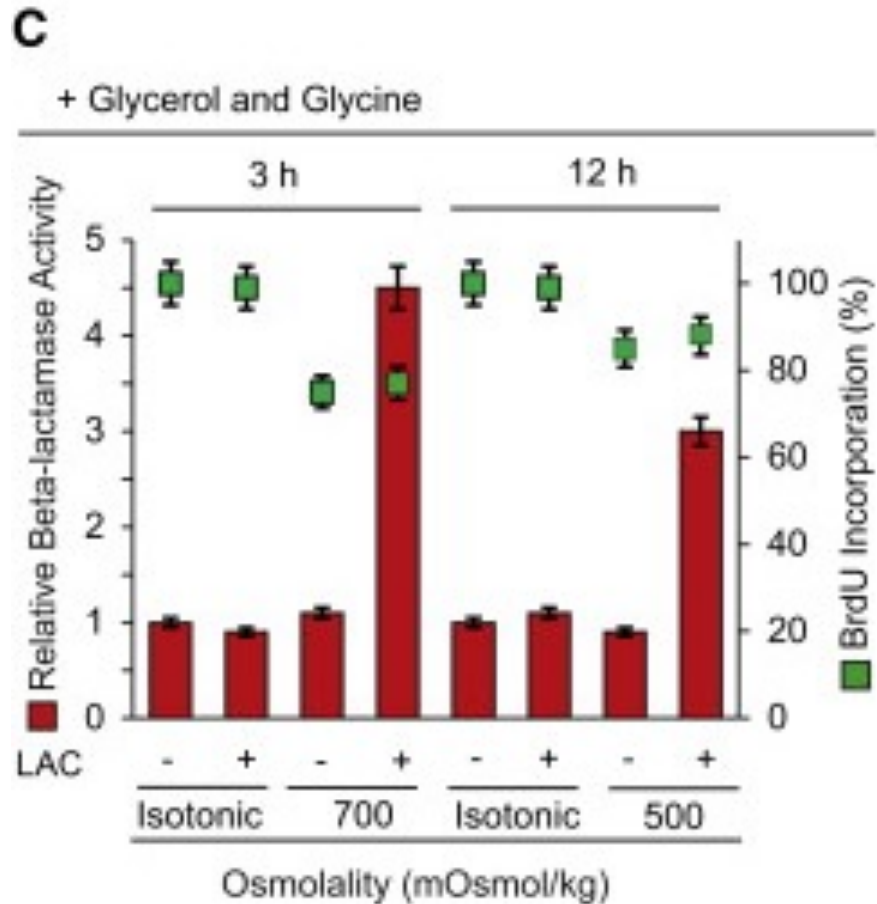


# Glycerol and glycine rescue hypertonicity-induced cell-cycle inhibition

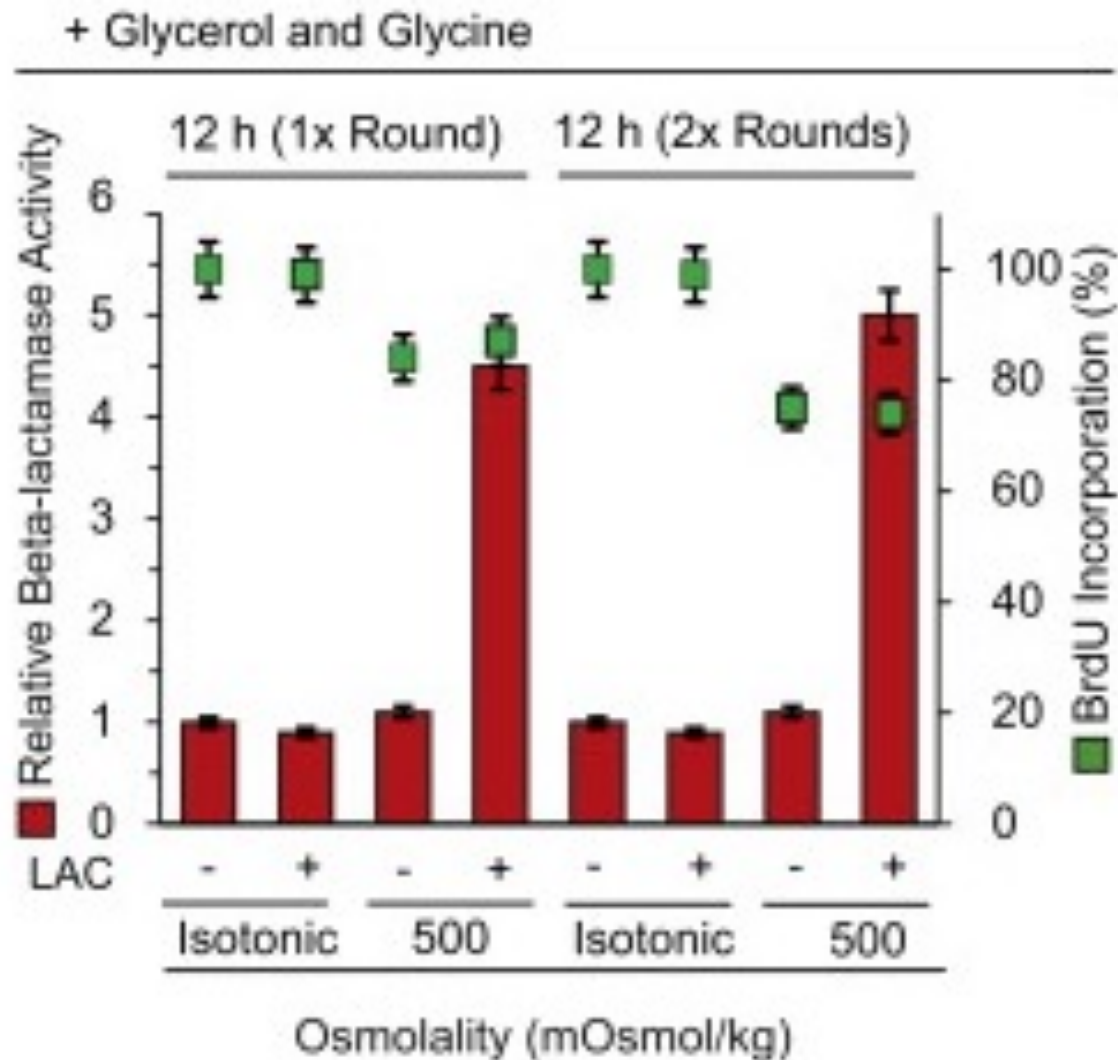




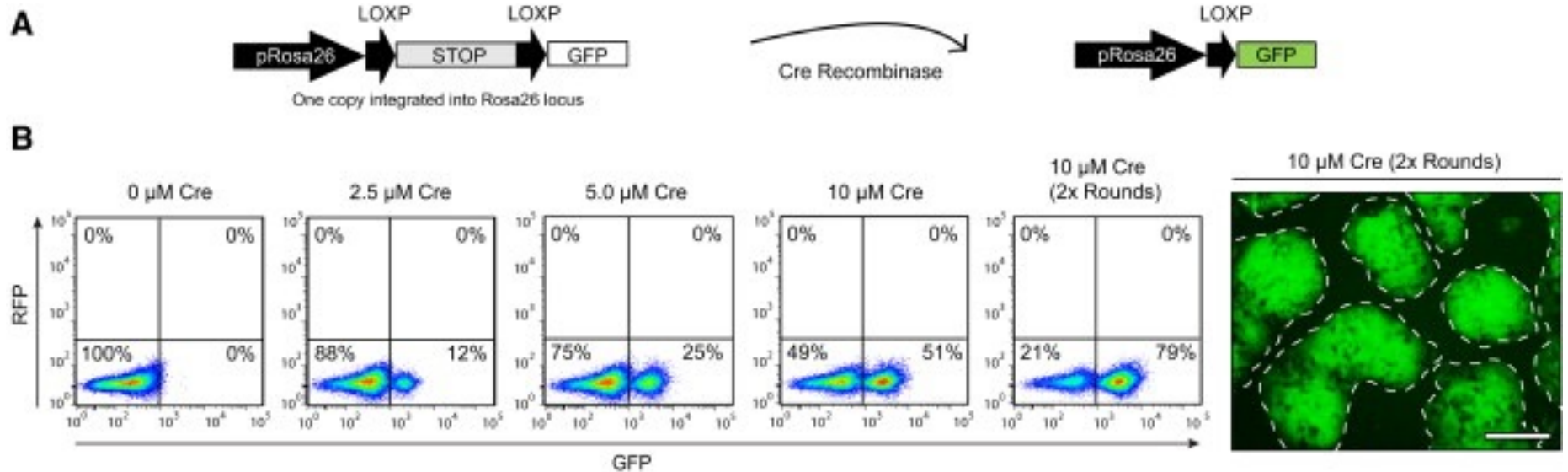
# Addition of glycerol/glycine gives similar transduction efficiencies in MEFs but not in mESCs



**Multiple rounds of hypertonicity lead to high transduction yield and remain cell viability in sensitive cell lines (i.e. mESC)**



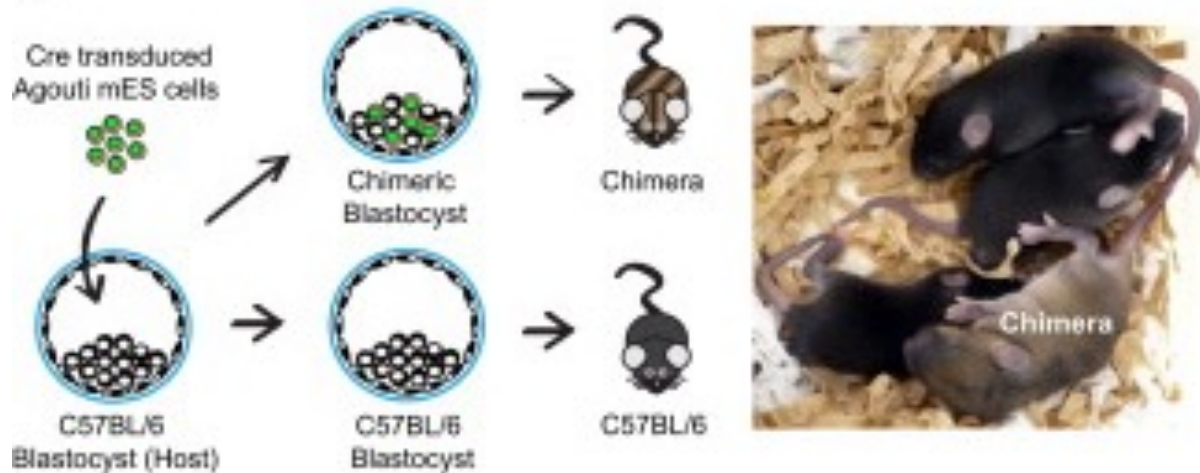
# Efficient protein transduction in multiple primary cell types



„mESCs were transduced with Cre protein at 500 mOsmol/kg for 12 hr“

# Cre protein transduction does not disrupt mESC pluripotency and Cre-transduced mESCs contribute to the germline

C



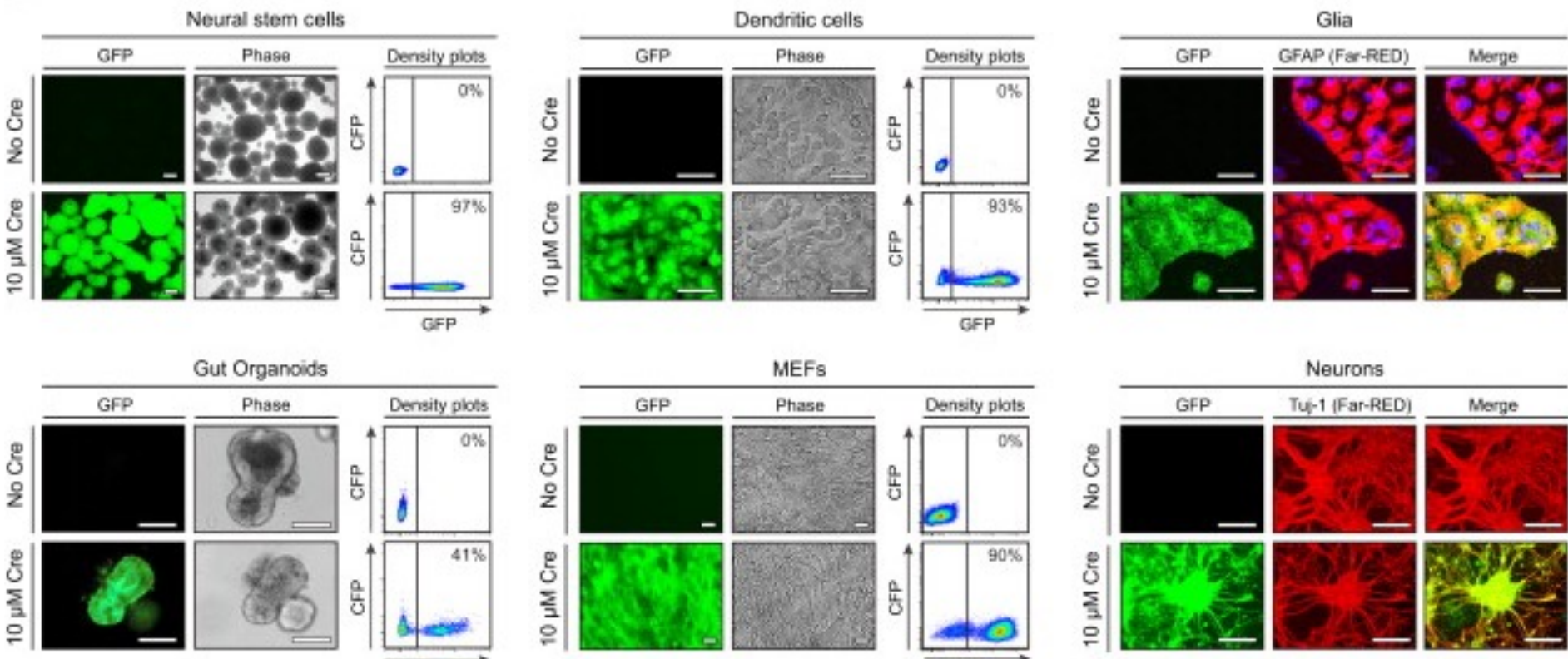
D



# iTOP facilitates the efficient transduction of multiple primary cell lines



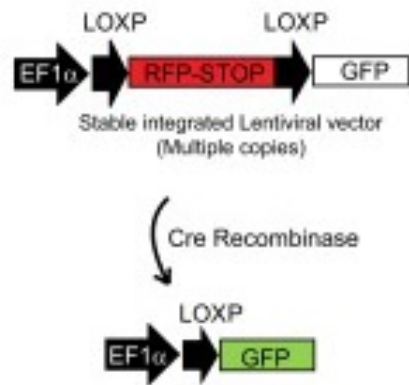
E



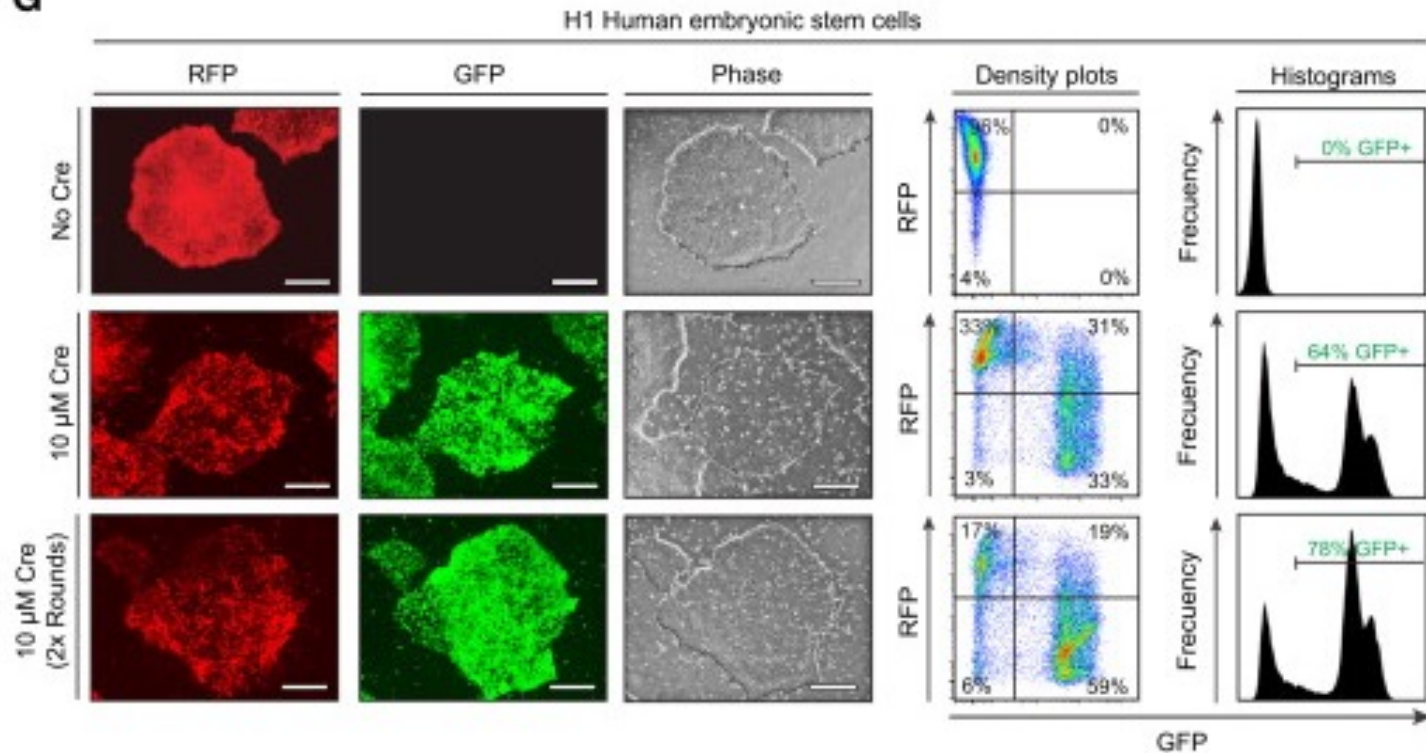


# Efficient transduction with iTOP in human ESCs

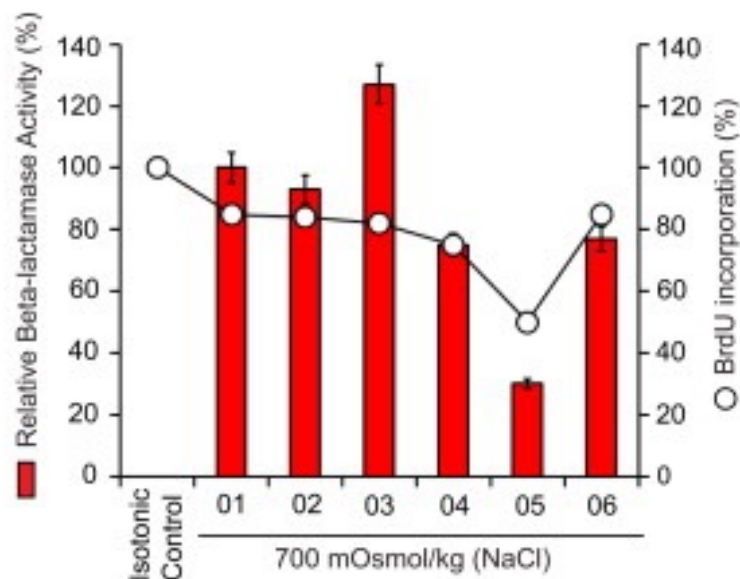
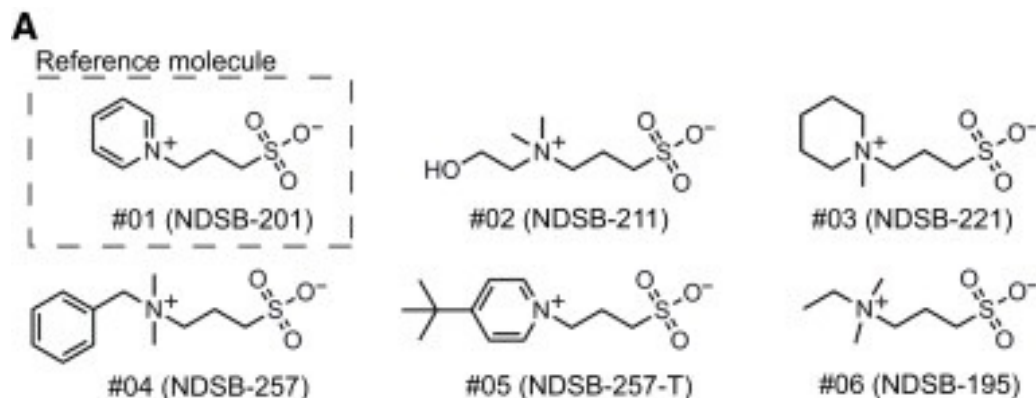
F



G



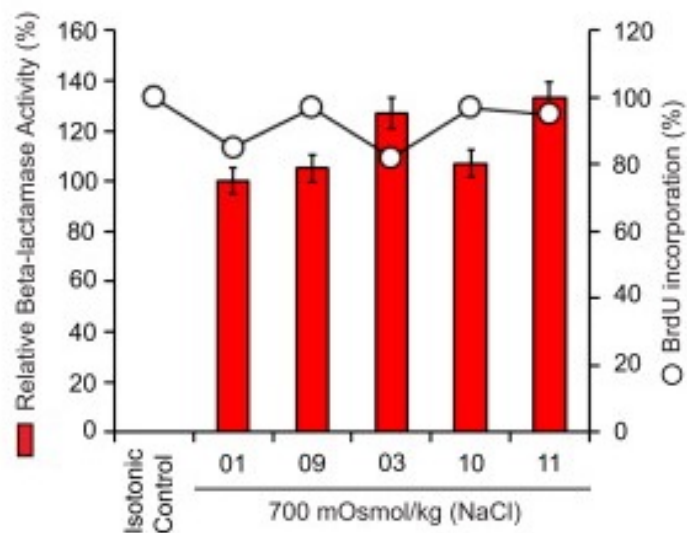
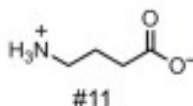
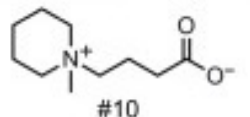
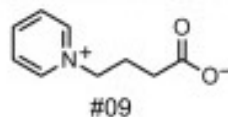
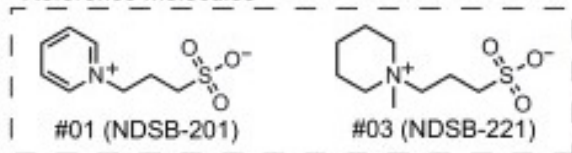
# Structural Features of the Transduction Compound



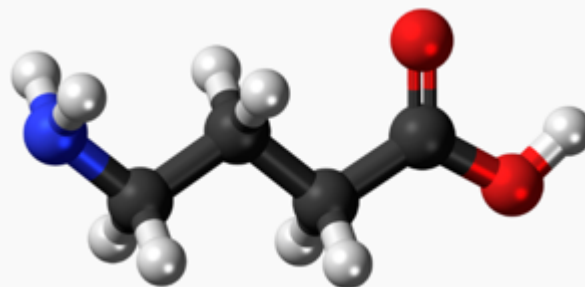
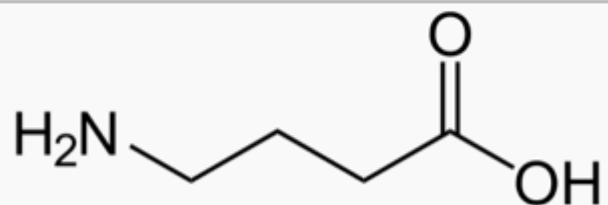
# Replacing sulfonate with carboxyl residues + replacing the amine improves cell viability

B

Reference molecules

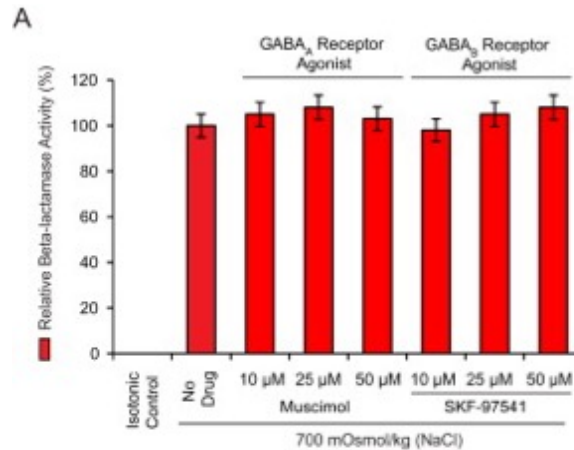


*gamma*-Aminobutyric acid

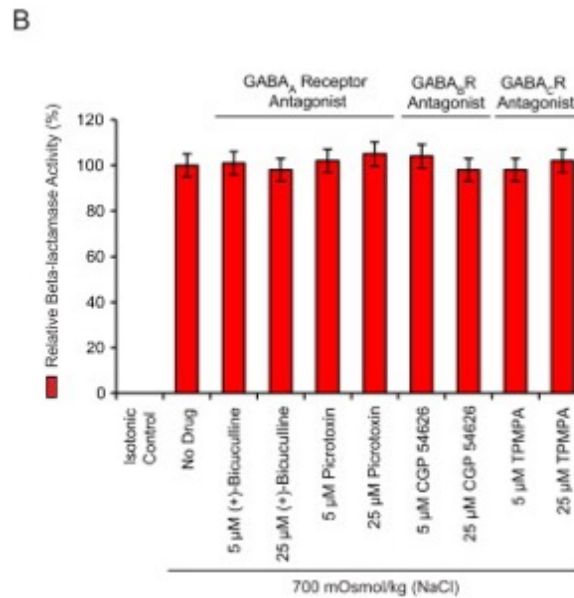




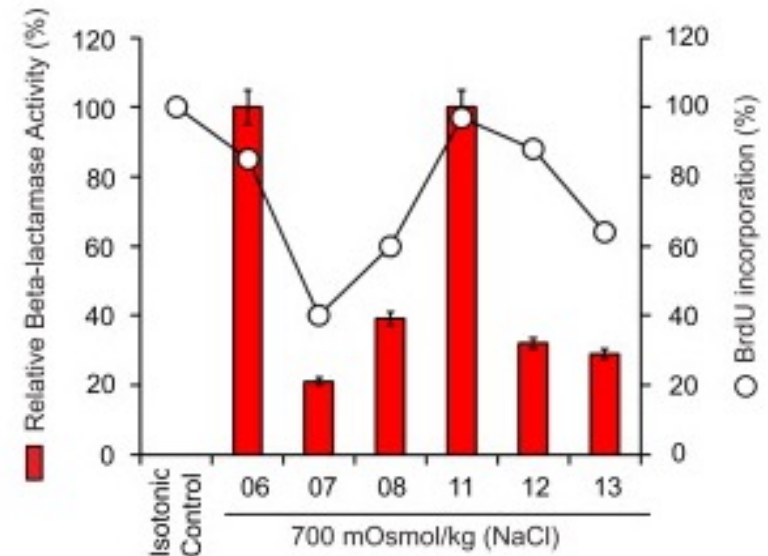
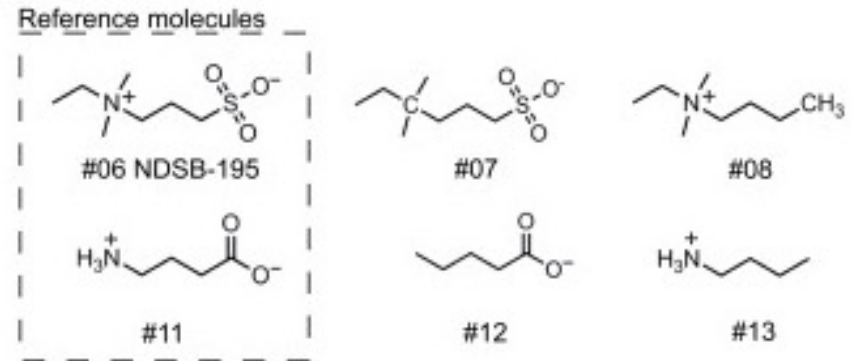
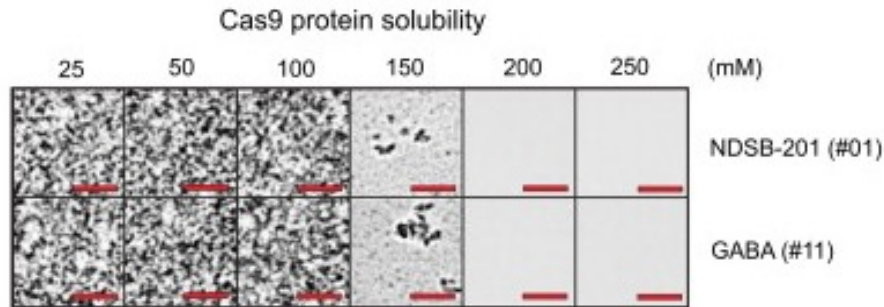
# GABA-R (ant-)agonists do not impact transduction



25 mM GABA needed for iTOP is 5 logs higher than concentrations needed for neurotransmission



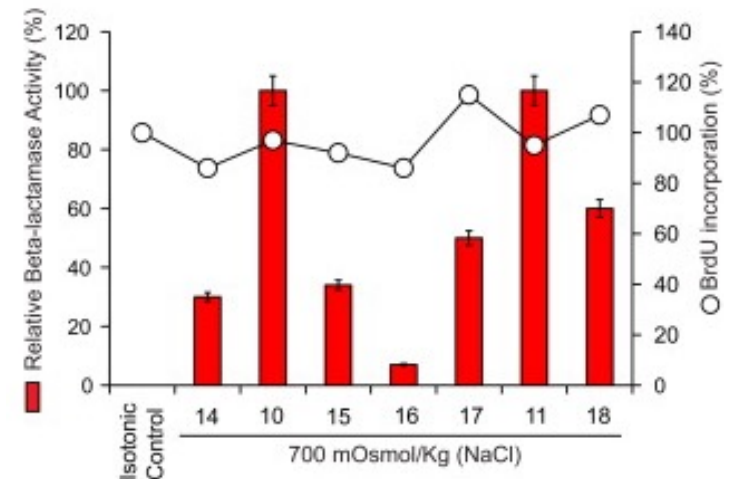
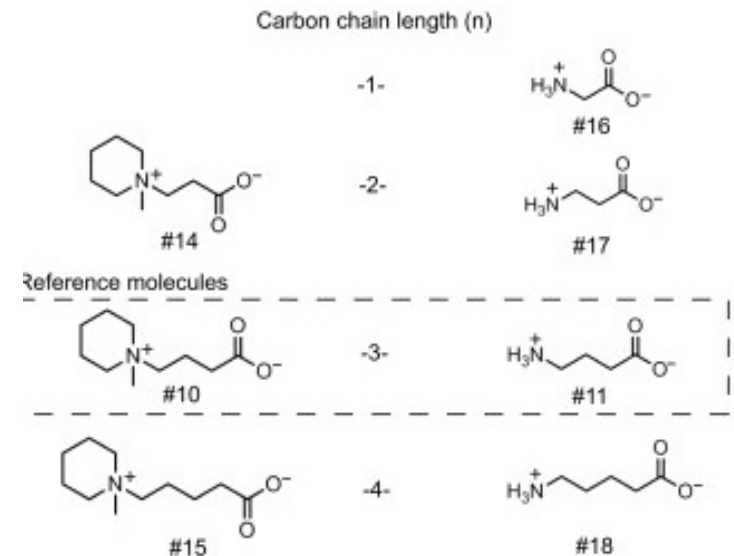
# GABA has similar properties in increasing protein solubility as NDSB-201 and both amine and carboxyl groups are needed for protein transduction



# Determination of the the optimal length of the carbon chain spacer

## The ideal molecule

- zwitterionic @ neutral pH with
  - (-) + charged amino group
  - (-) – charged sulfonyl/carboxyl group
- 3 carbons between  $N_{Rx}^+$  and  $SOO^-/COO^-$  groups



C

Pictures

Main components of Transduction media



- Culture media adjusted with NaCl to 700 mOsmol/kg
- **50 mM compound #14**
- 1  $\mu$ M Beta-lactamase



- Culture media adjusted with NaCl to 700 mOsmol/kg
- **50 mM compound #10**
- 1  $\mu$ M Beta-lactamase



- Culture media adjusted with NaCl to 700 mOsmol/kg
- **50 mM compound #15**
- 1  $\mu$ M Beta-lactamase



- Culture media adjusted with NaCl to 700 mOsmol/kg
- **50 mM compound #16**
- 1  $\mu$ M Beta-lactamase



- Culture media adjusted with NaCl to 700 mOsmol/kg
- **50 mM compound #17**
- 1  $\mu$ M Beta-lactamase



- Culture media adjusted with NaCl to 700 mOsmol/kg
- **50 mM compound #11 (GABA)**
- 1  $\mu$ M Beta-lactamase



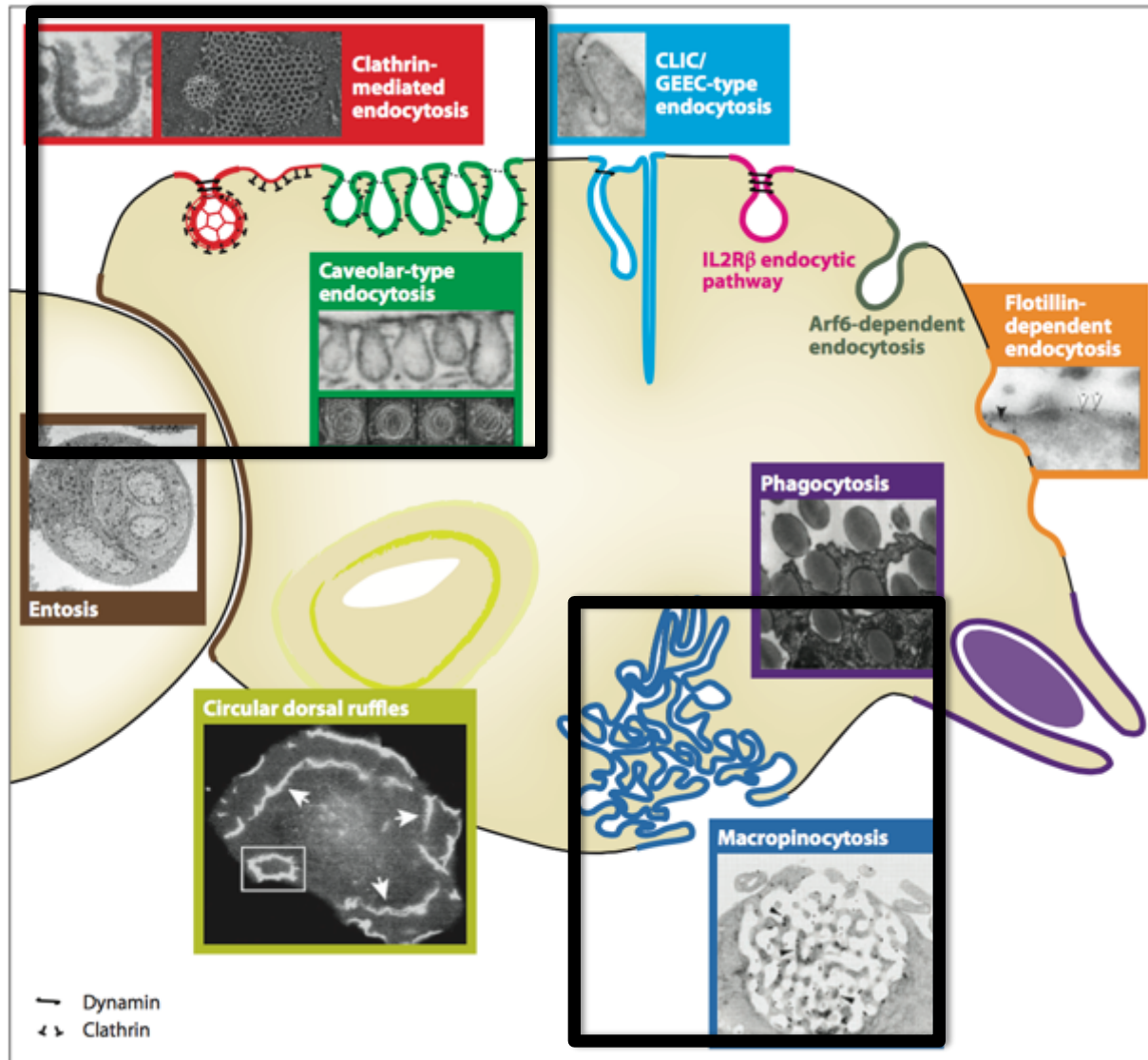
- Culture media adjusted with NaCl to 700 mOsmol/kg
- **50 mM compound #18**
- 1  $\mu$ M Beta-lactamase



- Culture media adjusted with NaCl to 700 mOsmol/kg
- **50 mM compound #11 (GABA)**
- 1  $\mu$ M Beta-lactamase
- 50 % Ethanol

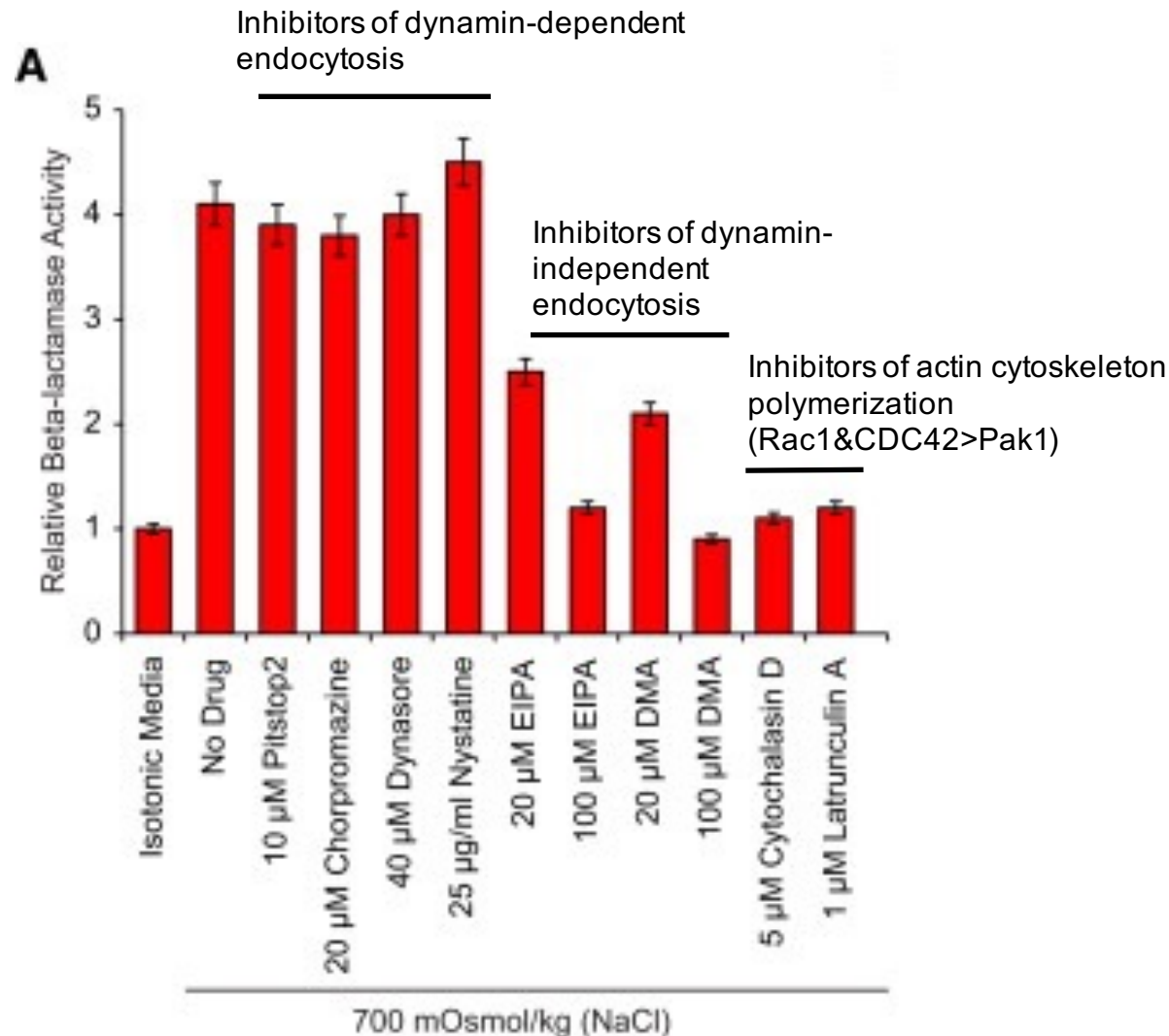
**Transduction  
efficiacies are  
not confounded  
by differences  
in protein  
solubility**

# Dissecting the Mechanism of Protein Transduction



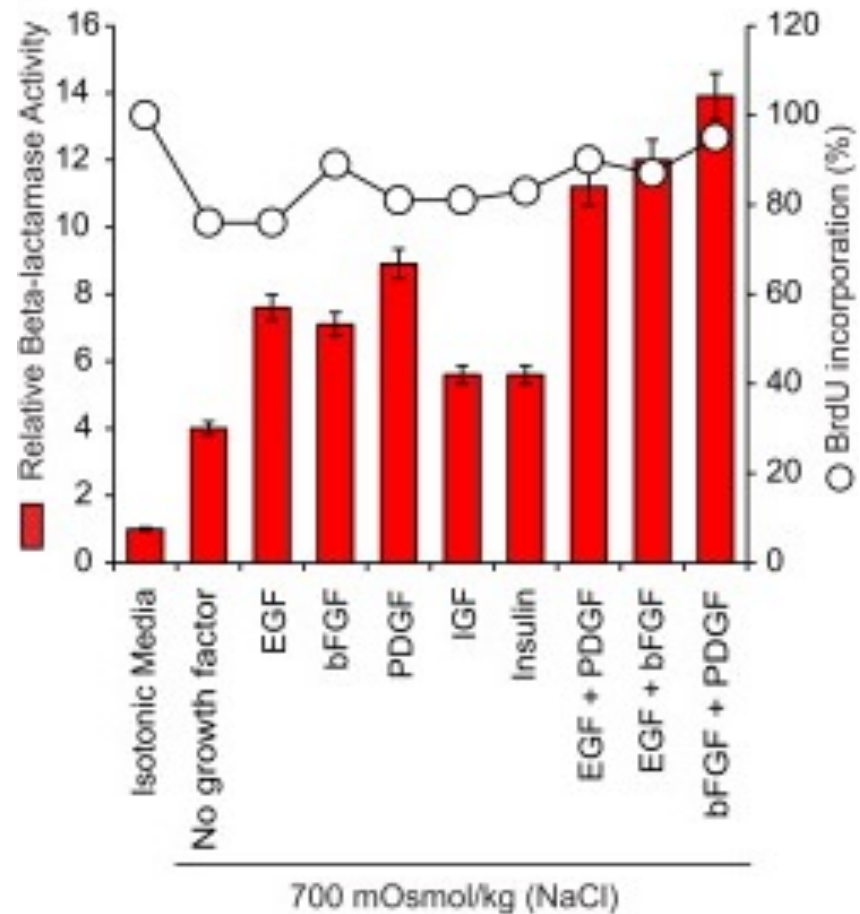
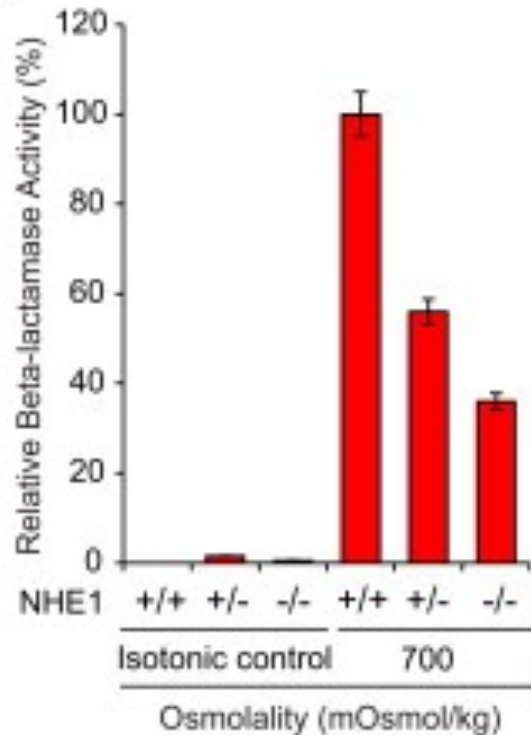
Doherty GJ & McMahon HAT  
Annu Rev Biochemistry 2009

# Protein transduction is mediated by macropinocytosis



**NHE1 Na<sup>+</sup>/H<sup>+</sup> antiporter function is important for protein transduction through macropinocytosis, which is enhanced through TK-acting growth factors**

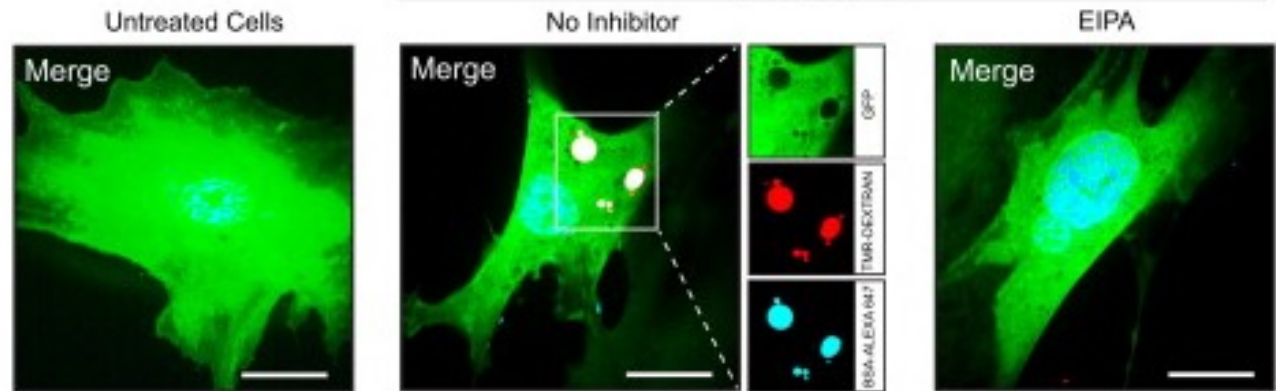
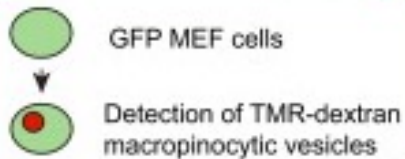
3



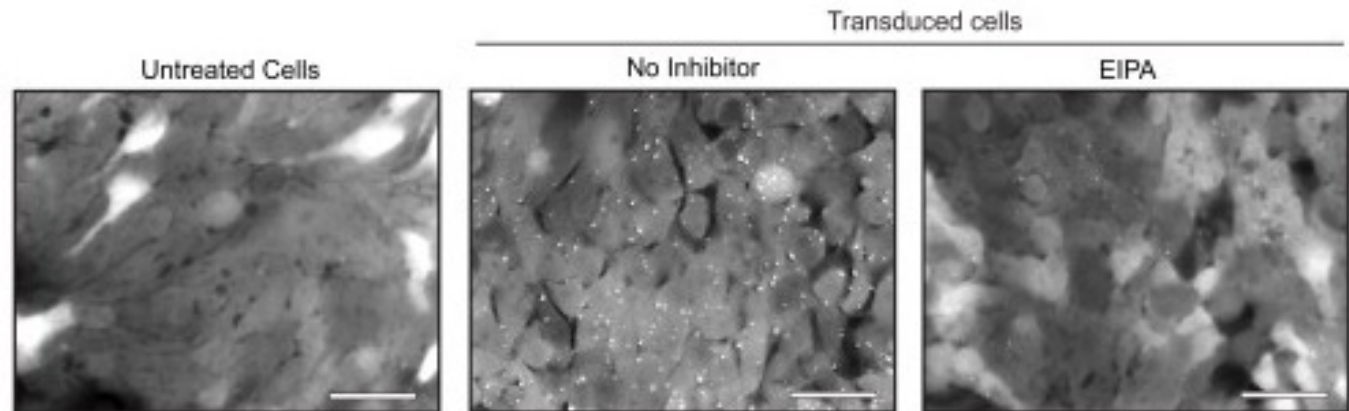
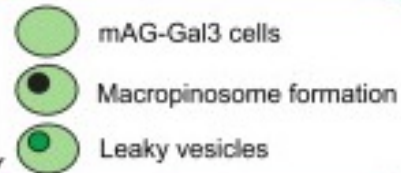


# Quantifying macropinocytosis and vesicle release

## Macropinocytosis

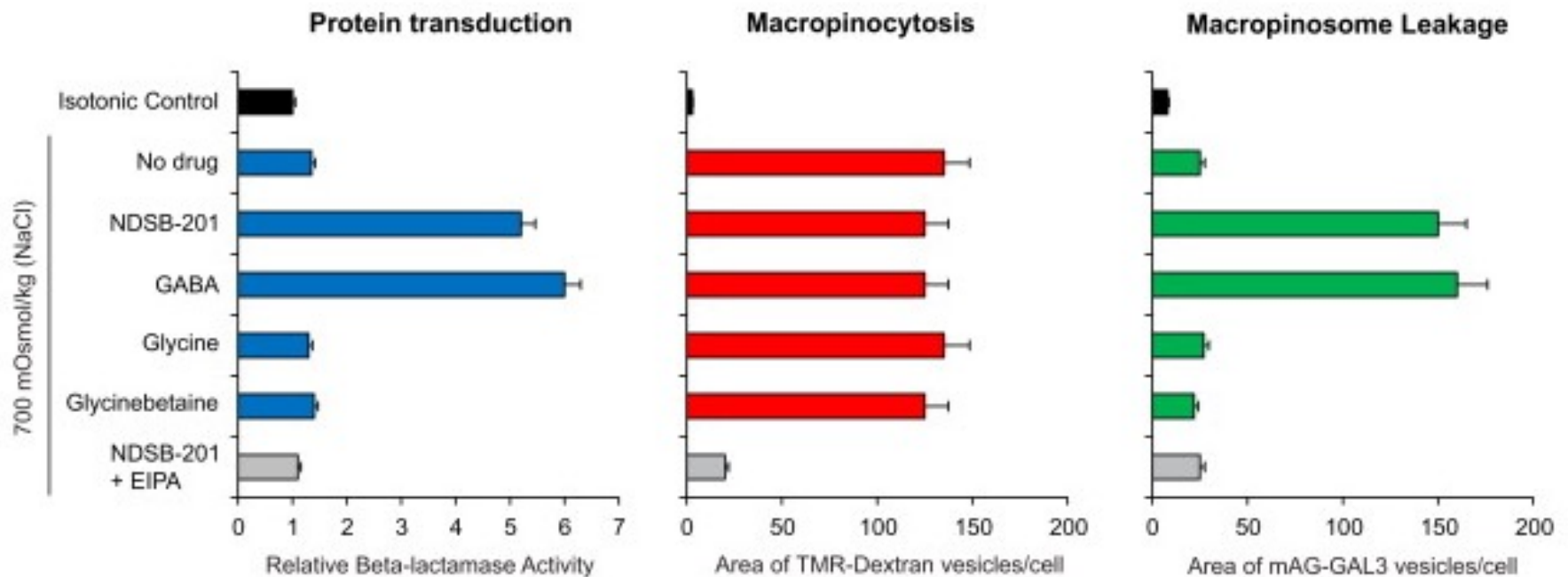


## Macropinocytic vesicle leakage



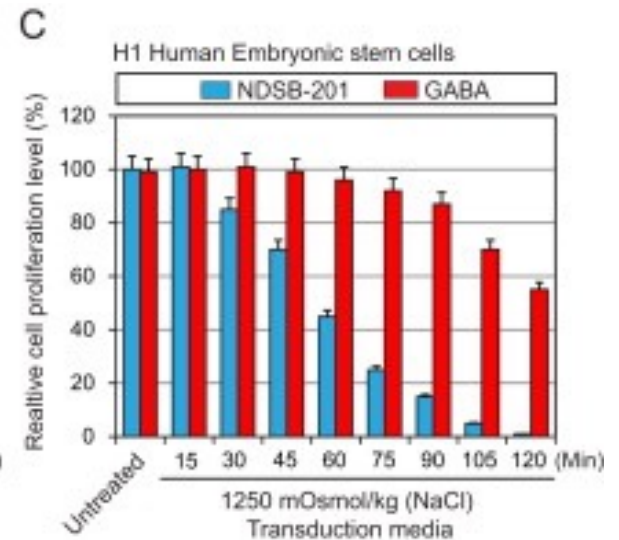
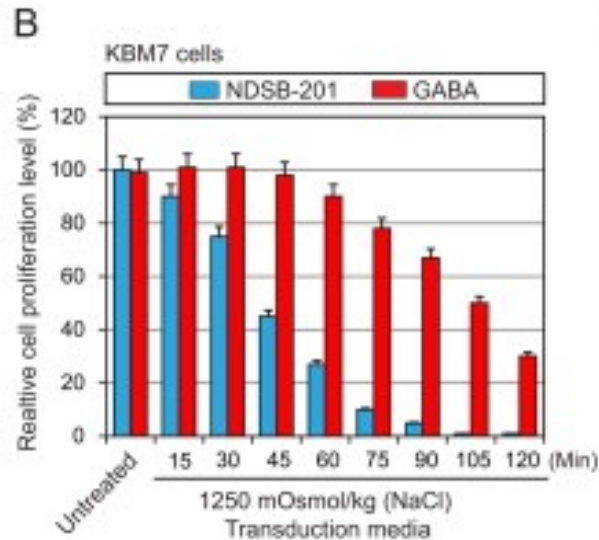
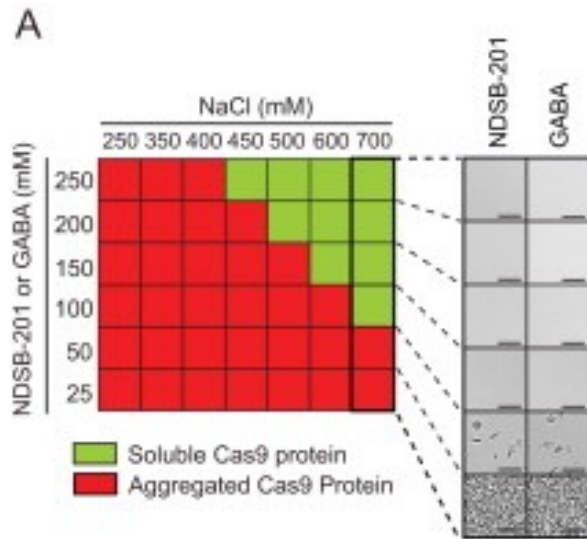
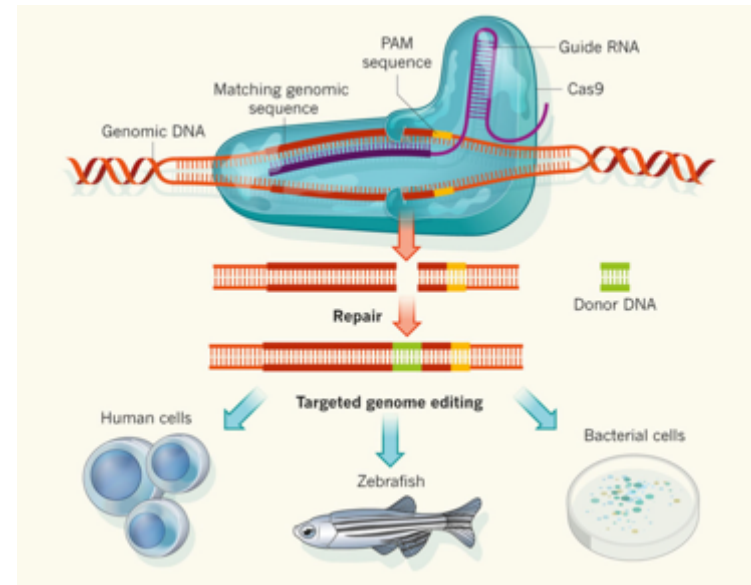


# Quantifying macropinocytosis and vesicle release



# iTOP for Crispr/Cas9 in primary cells

Charpentier E & Doudna JA  
Nature 2013



# High salt concentrations and DNA breaks

Proceedings of the National Academy of Sciences of the United States of America

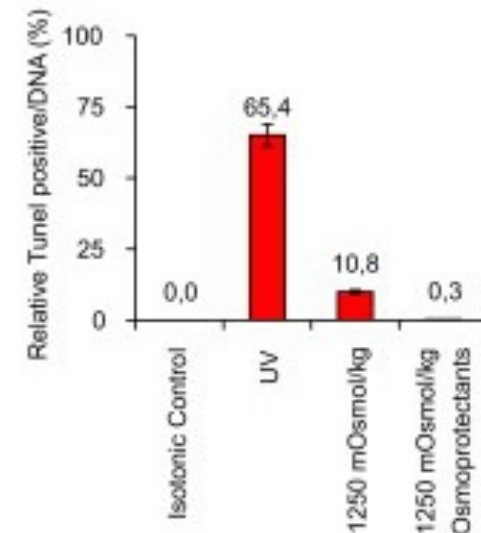
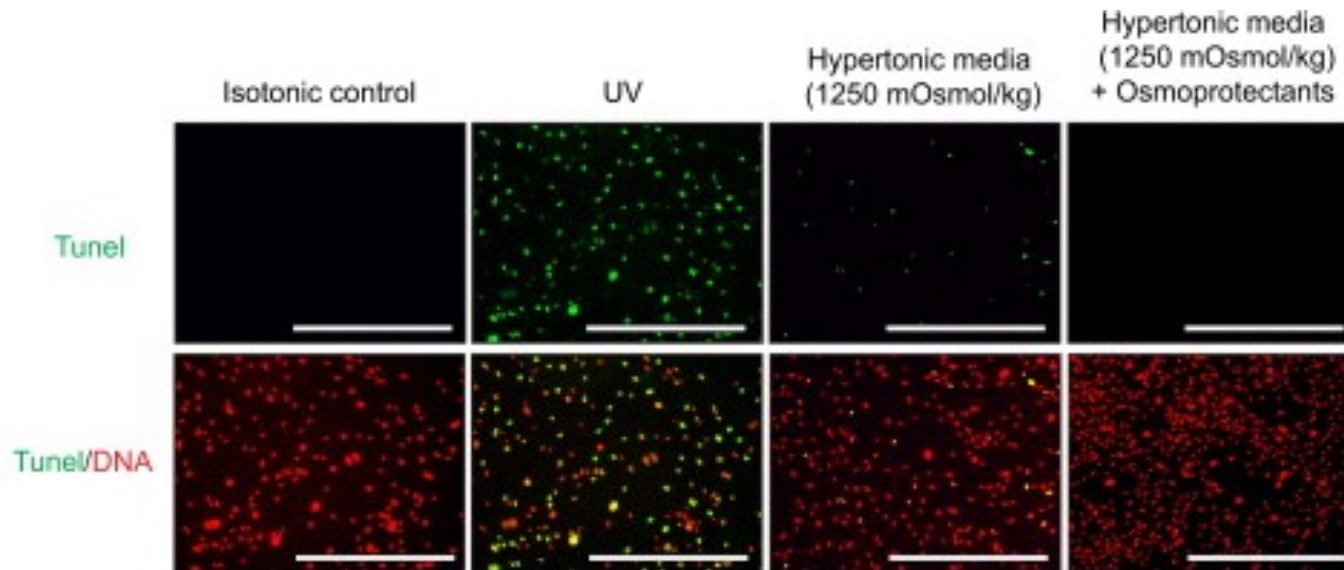
CURRENT ISSUE // ARCHIVE // NEWS & MULTIMEDIA // FOR AUTHORS // ABOUT PNAS // COLLECTED

Home > Current Issue > vol. 108 no. 51 > Christophe E. Redon, 20281–20282, doi: 10.1073/pnas.1117713109

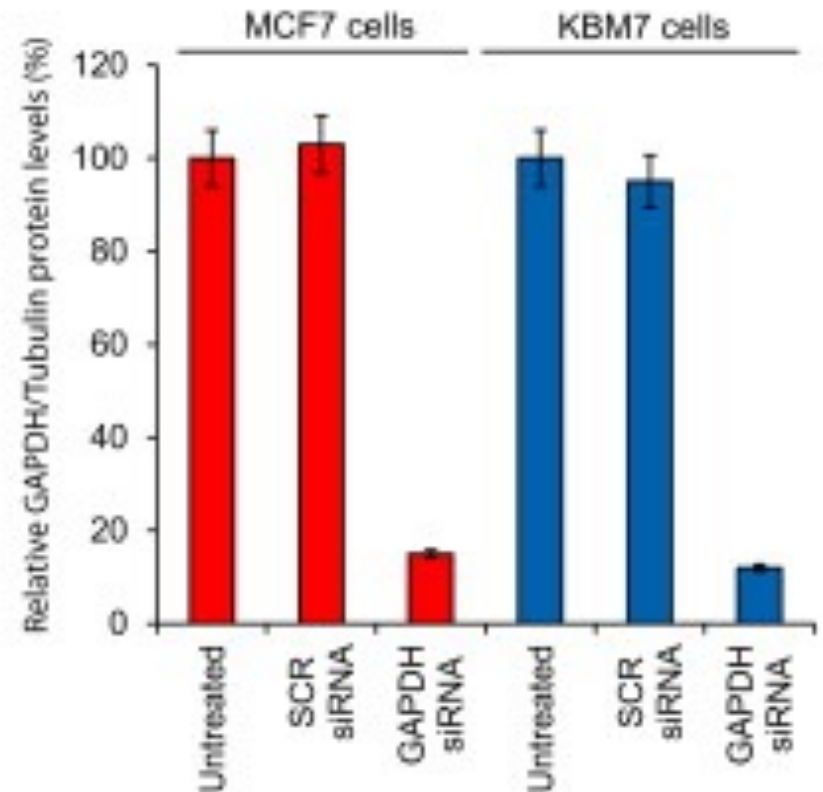
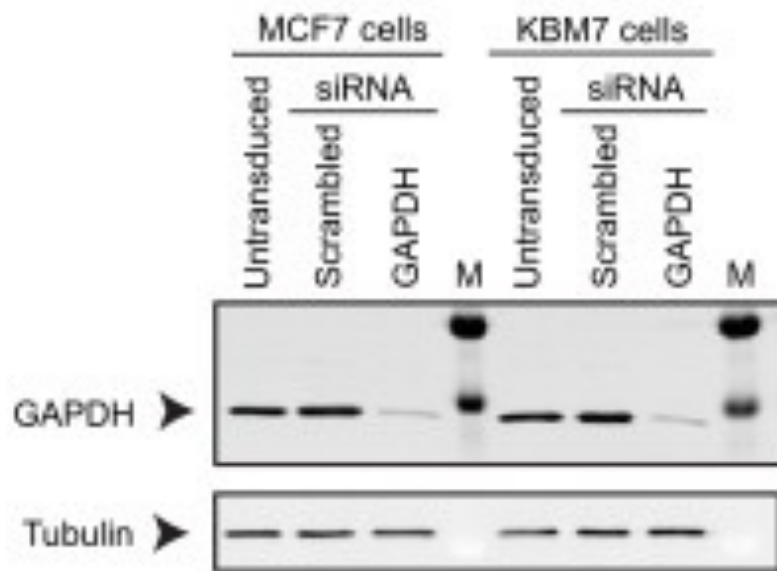


## High salt and DNA double-strand breaks

Christophe E. Redon and William M. Bonner<sup>1</sup>

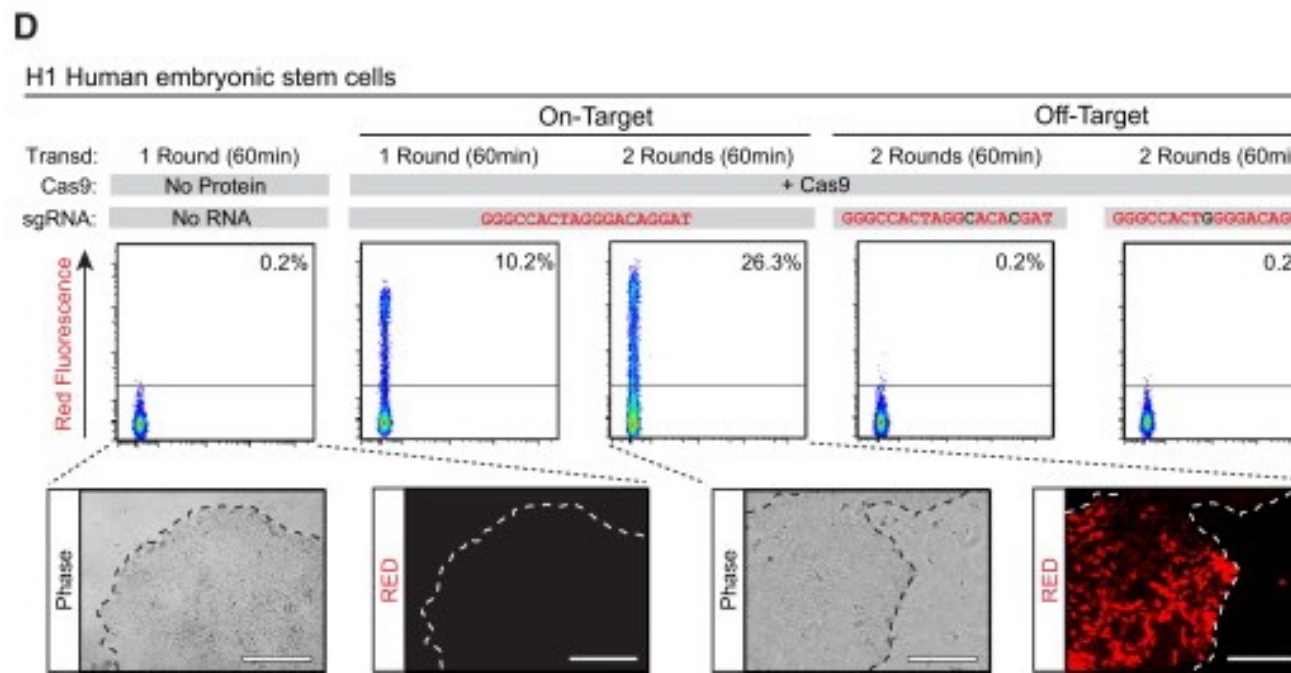
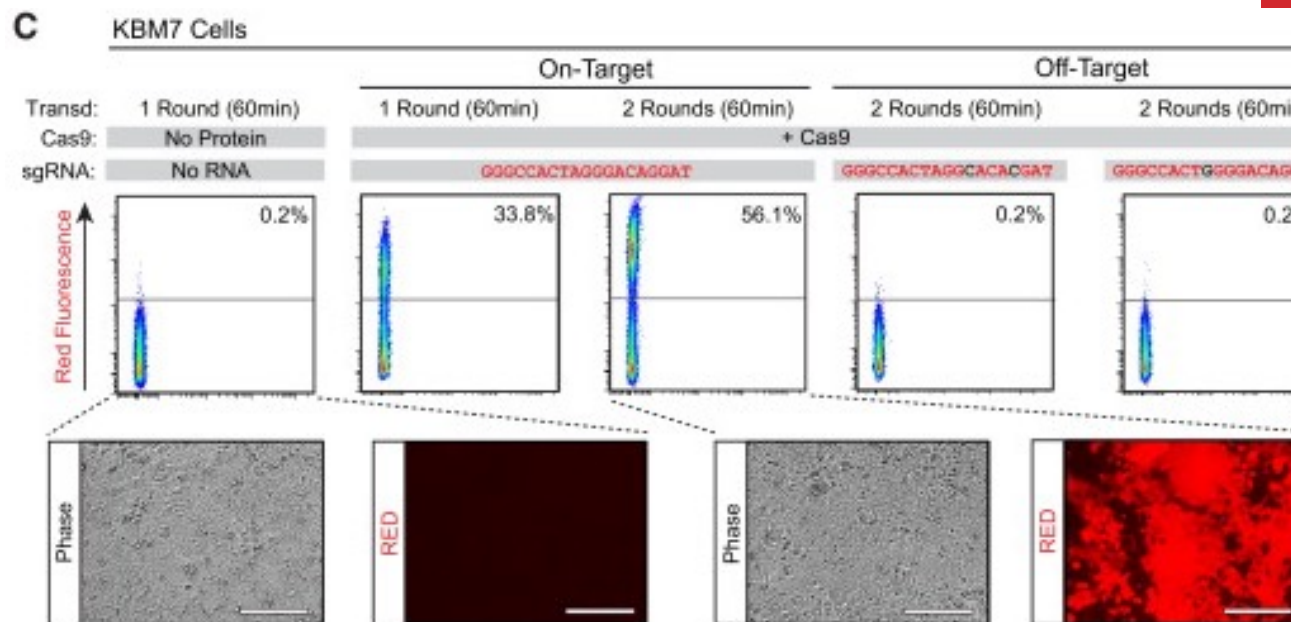


# iTOP does not seem to affect (si)RNA delivery





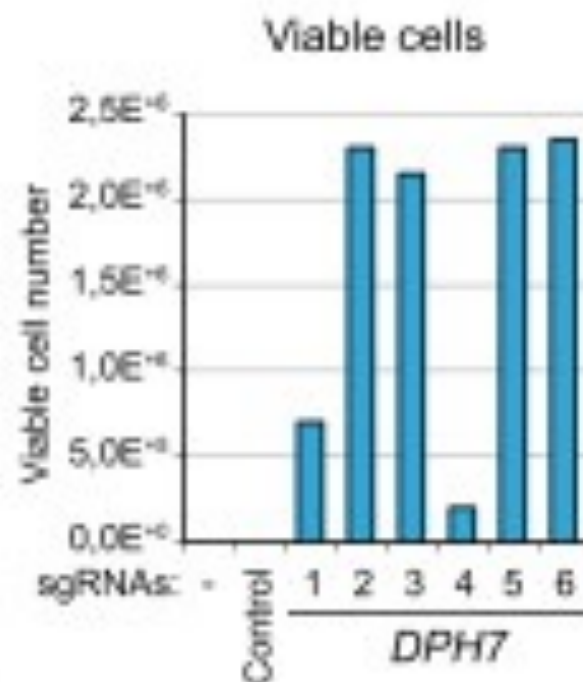
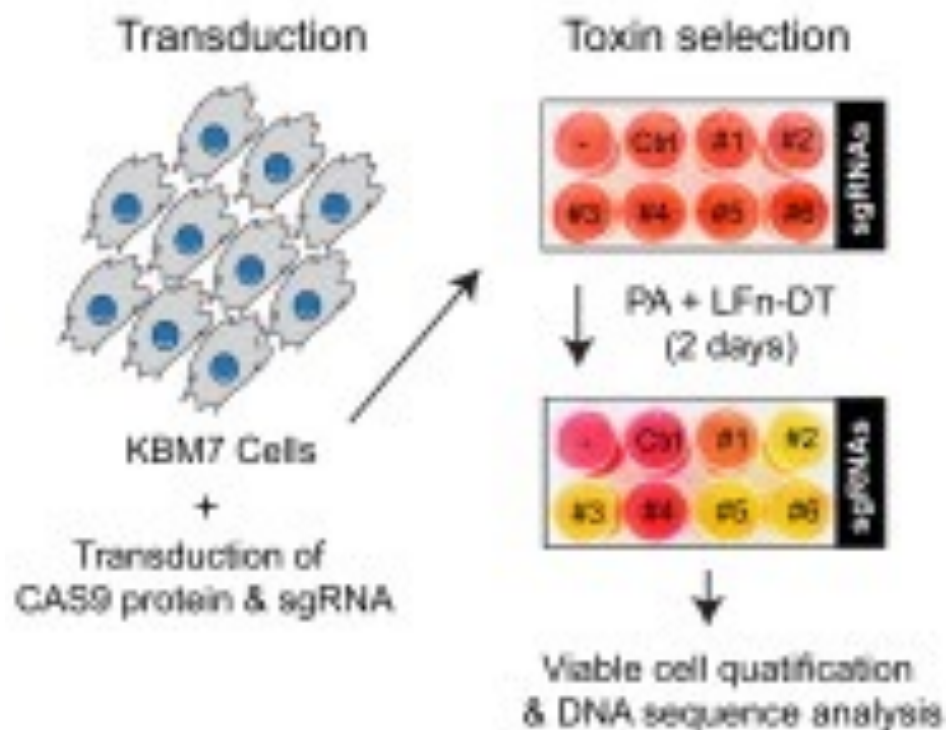
# iTOP/ Crispr-Cas9





# iTOP/Crispr-Cas9

**A**



# iTOP/Crispr-Cas9

## B

*DPH7* sgRNA #2; 11 mutant sequences out of 11 sequences = 100%

```
ATGATGGGCTGTTTGGCCCTGCAAAACGGTGGACACCGAGCTGACCGCGGACTCGGTGGAGTGGTGCDCGCTGCAAGGCT WT
ATGATGGGCTGTTTGGCCCTGCAAAACGGTGGACACCGAGCTGCGCGGACTCGGTGGAGTGGTGCDCGCTGCAAGGCT D1
ATGATGGGCTGTTTGGCCCTGCAAAACGGTGGACACCGAGCTGAC-----TCGGTGGAGTGGTGCDCGCTGCAAGGCT D7 [3x]
ATGATGGGCTGTTTGGCCCTGCAAA-----CGGACTCGGTGGAGTGGTGCDCGCTGCAAGGCT D21
ATGATGGGCTGTTTGGCCCTGCAAAACGGTGG-----GTGGTGCDCGCTGCAAGGCT D27

ATGATGGGCTGTTTGGCCCTGCAAAACGGTGGACACCGAGCTGAACCGCGGACTCGGTGGAGTGGTGCDCGCTGCAAGGC +1 [4x]
ATGATGGGCTGTTTGGCCCTGCAAAACGGTGGACACCGAGCTGATCCGCGGACTCGGTGGAGTGGTGCDCGCTGCAAGGC +1
```

*DPH7* sgRNA #6; 12 mutant sequences out of 12 sequences = 100%

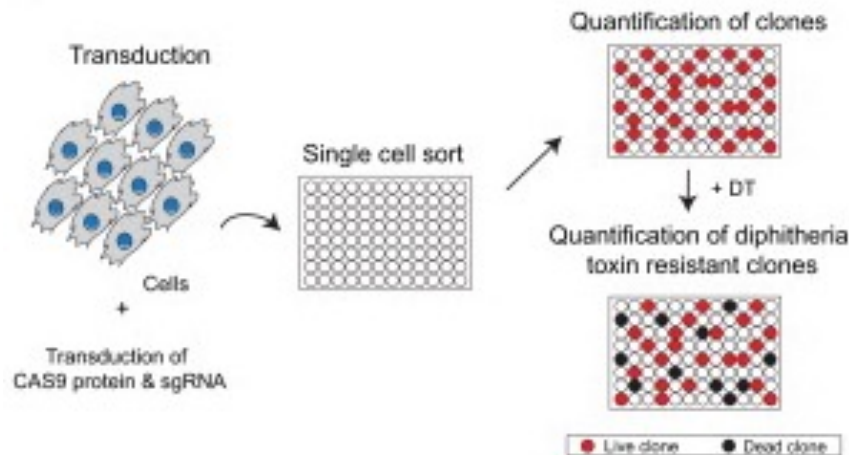
```
AGGGCAGCTCCADCTCCTGATGGTGAATGAGACGAGGCCCCAGGCTGCAGAAASTGGCTTCATGGCAGGCACATCAATTC WT
AGGGCAGCTCCADCTCCTGATGGTGAATGAGACGAGGCCCCAGGCTGCAGAAAG-----TGGCAGGCACATCAATTC D8 [3x]
AGGGCAGCTCCADCTCCTGATGGTGAATGAGACGAGGCCCCAGGCTGCAGAAAG-----GCACATCAATTC D14/+1
AGGGCAGCTCCADCTCCTGATGGTGAATGAGACGAGGCCCCAGGCTG-----CATCAATTC D24 [2x]
AGGGCAGCTCCADCTCCTGATGGTGAATGAGACGAGGCCCCAGGCT-----CATCAATTC D25
AGGGCAGCTACADGTCCTG-----ATGGCAGGCACATCAATTC D41
AG-----GGCAGGCACATCAATTC D64

AGGGCAGCTCCADCTCCTGATGGTGAATGAGACGAGGCCCCAGGCTGCAGAAASTGGCTTCATGGCAGGCACATCAATTC +1
AGGGCAGCTCCADCTCCTGATGGTGAATGAGACGAGGCCCCAGGCTGCAGAAASTGGCTTCATGGCAGGCACATCAATTC +1
TGTCTGGGAACCAAGA-----//-----GGCAGGCACATCAATTC D155
```

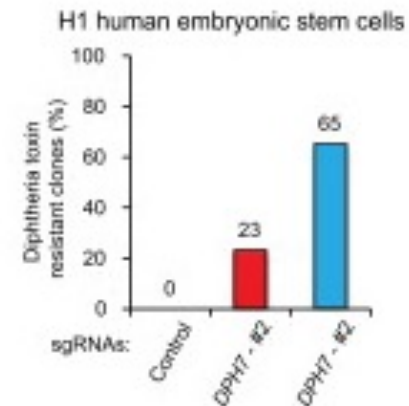
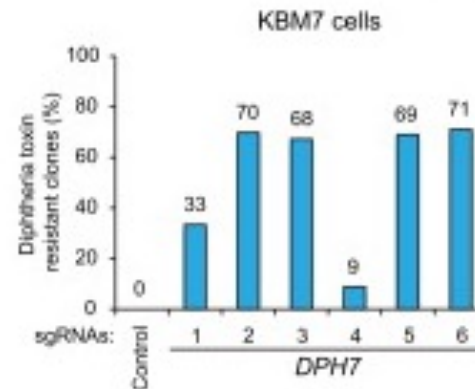


# Biallelic deletion efficiency with iTOP/Crispr-Cas9

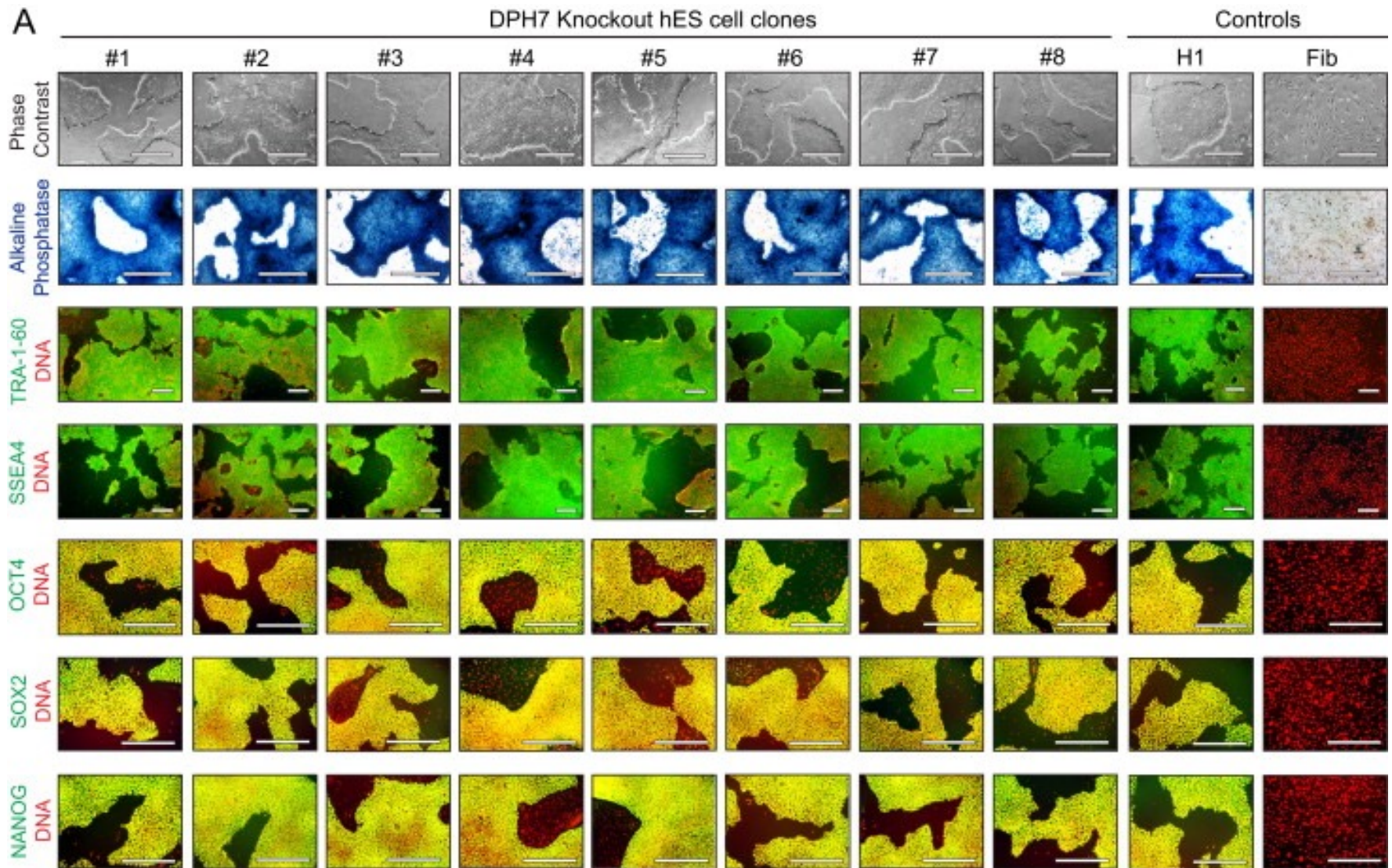
D



Biallelic *DPH7* gene knockout efficiency



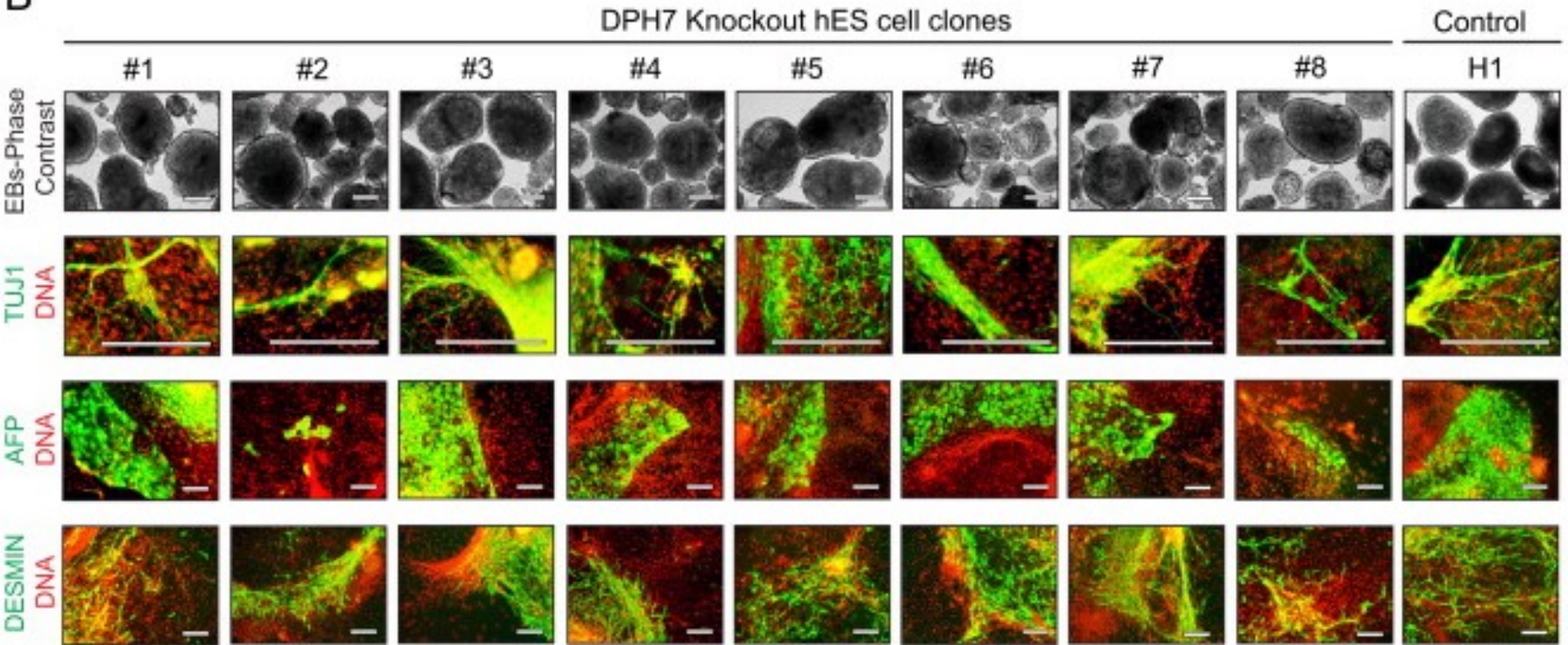
# Pluripotency markers are retained in iTOP/Crispr-Cas9 hESCs





# iTOP/Crispr-Cas9 treated hESCs can differentiate into cells of all 3 germ layers

B



# Summary/critical remarks

## - Pro:

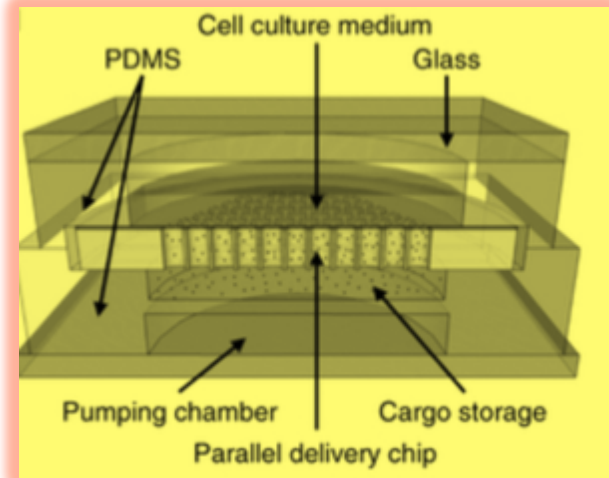
- Very convenient (no biosafety issues <> Lentivirus, AAV etc), fast (building lentivirus, AAV for a couple of weeks) and cheap technique (250g of NDSB 201 around 100 sFr),  
*5x transduction buffer*
  - *500 mM NaCl, 25 mM NaH<sub>2</sub>PO<sub>4</sub>, 250 mM NDSB-201, 150 mM glycerol, 75 mM glycine, 1.25 mM MgCl<sub>2</sub>, 1 mM 2-mercaptoethanol at pH 8.0*
- Very efficient with almost no cell toxicity
- Works well with primary cell lines (<> Crispr/Cas9 not so much)

## - Contra:

- Only transient transfection
- If applied more than 2 rounds > cell viability?
- Authors claim possibly suitable for treatment of genetic disease, no such data evident from present report

**WU Y ET AL., NATURE METHODS 2015**

# **MASSIVELY PARALLEL DELIVERY OF LARGE CARGO INTO MAMMALIAN CELLS WITH LIGHT PULSES**



# Physical approaches to bypass endocytosis

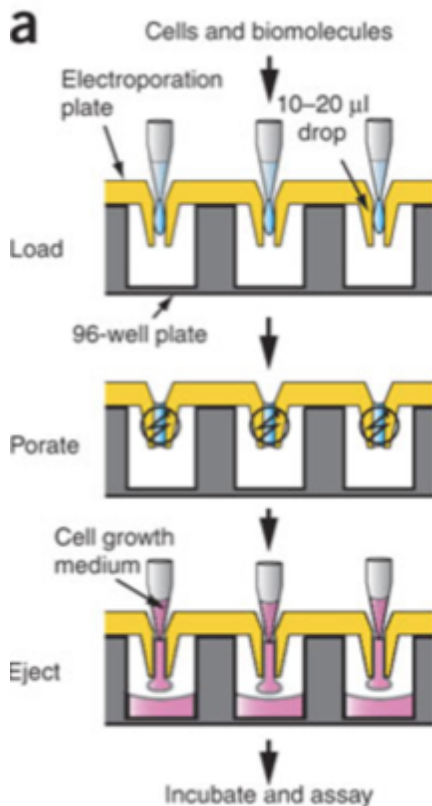
- 1) Create transient pores
- 2) Cargo delivery before pores reseal



## Ultrasound

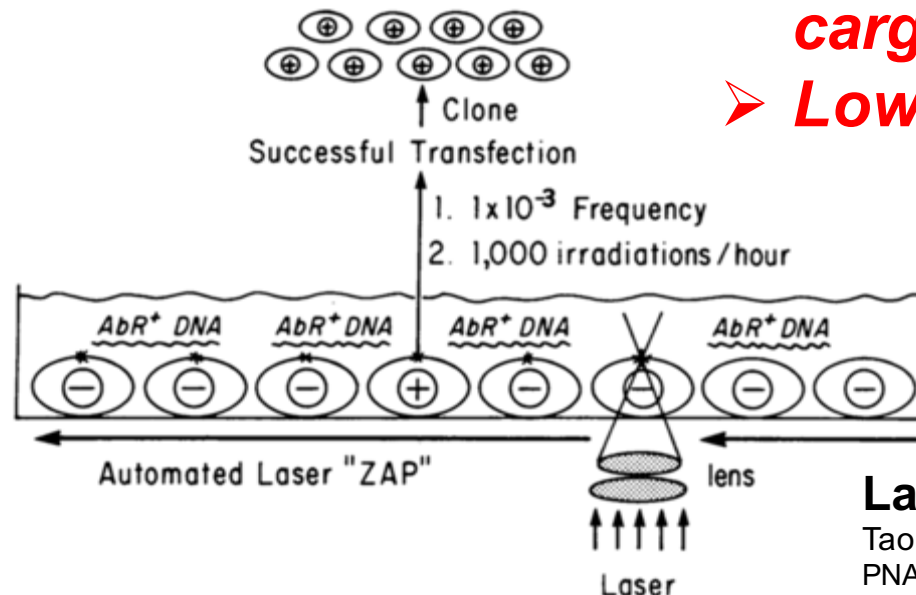
Mitragotri S  
Nat Rev Drug Discovery 2005

- Traumatic cell lysis with larger cargos
- Low throughput



## Drop electroporation

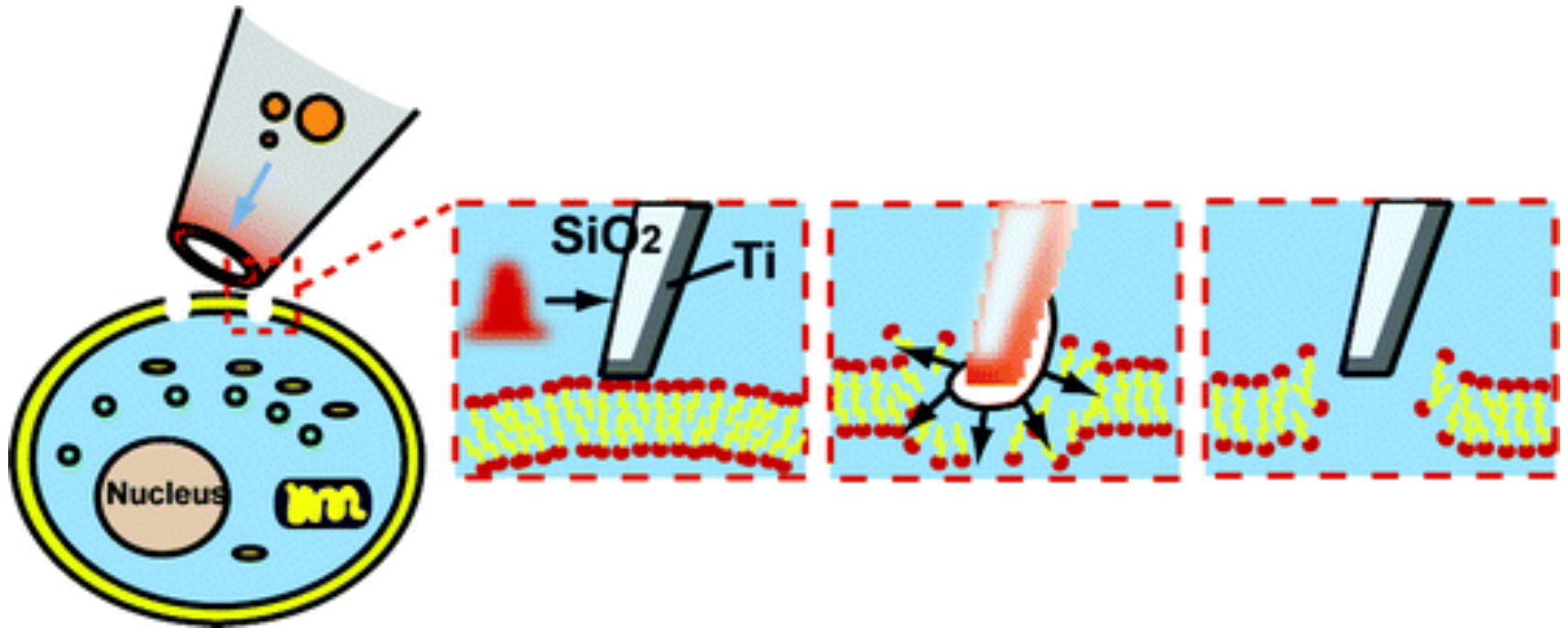
Guignet EG & Meyer T  
Nat Methods 2008



## Laser micropuncture

Tao W et al.  
PNAS 1987

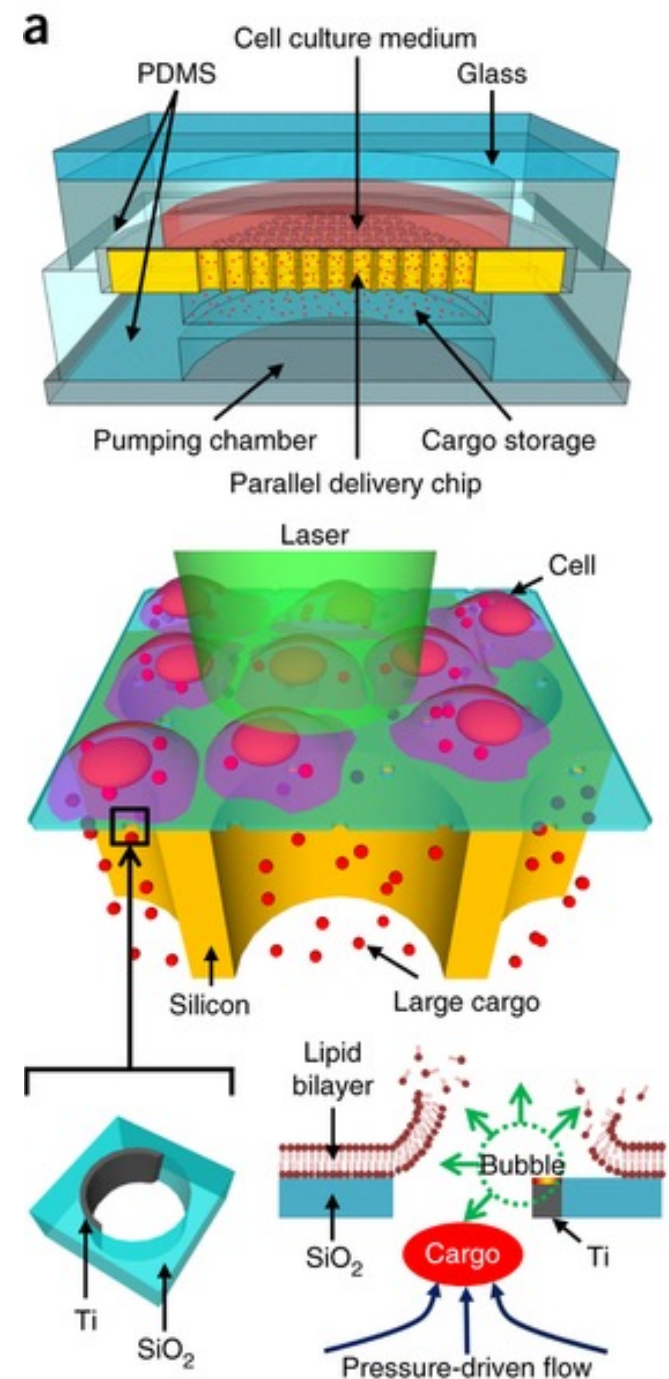
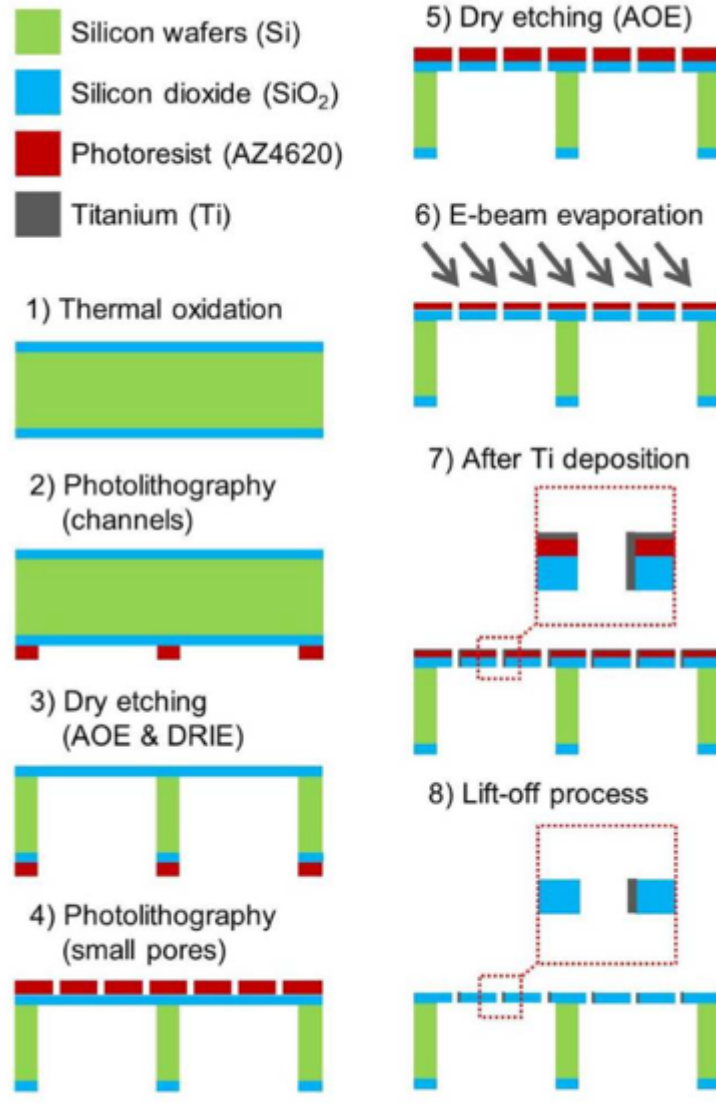
# Large cargo delivery – a photothermal nanoblade



Wu T et al., Analytical Chemistry 2011



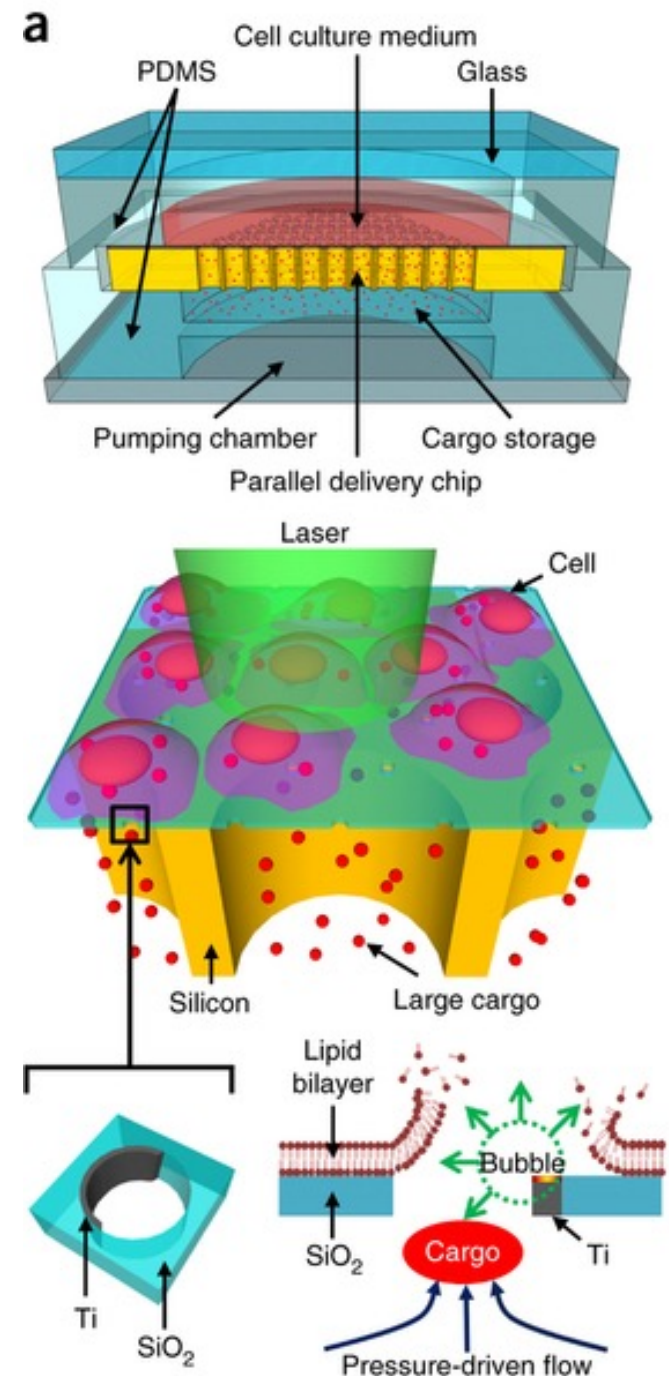
# BLAST – biophotonic laser-assisted surgery tool



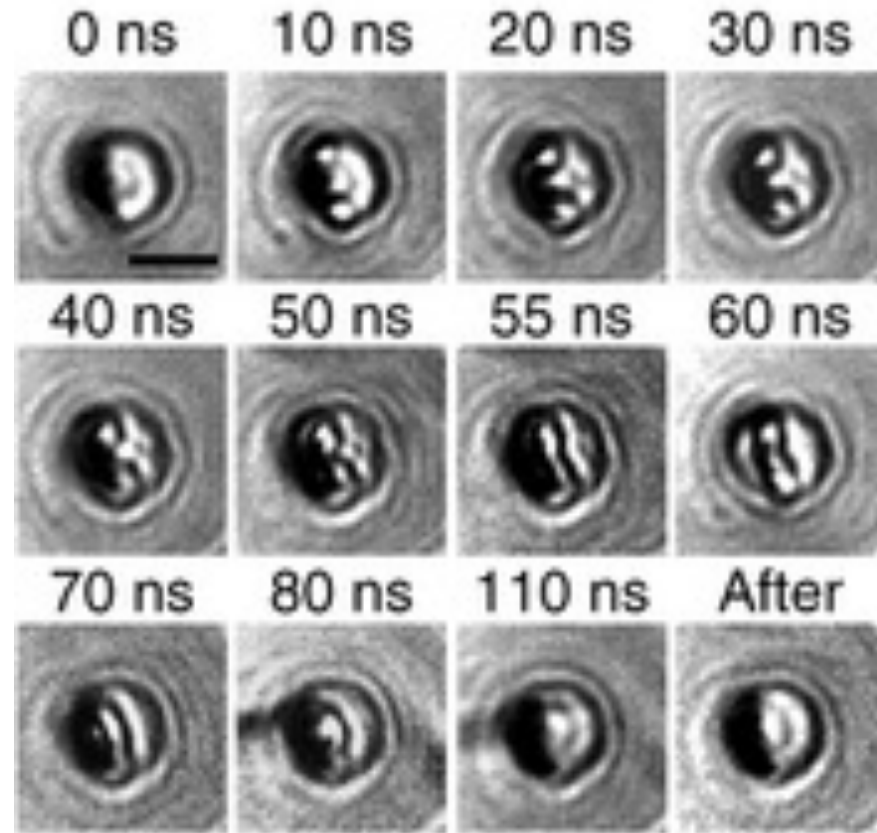
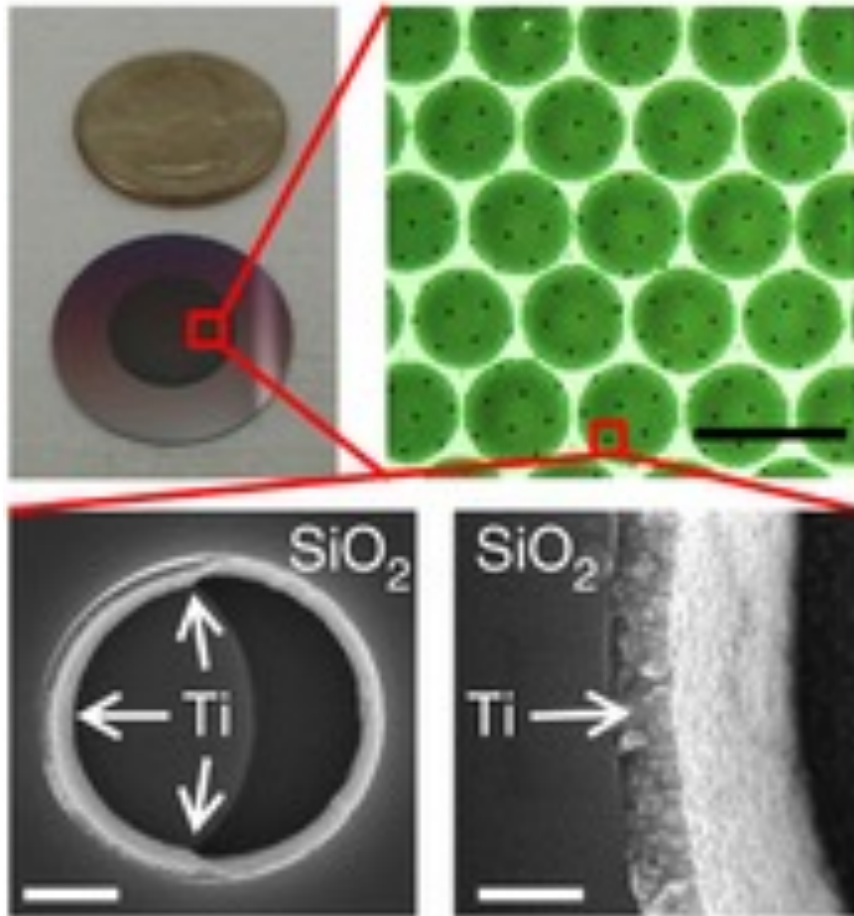
# BLAST – biophotonic laser-assisted surgery tool

- (i) Cells are cultured or made to adhere on a silicon chip
- (ii) The chip is assembled with a microliter chamber loaded with the cargo to be delivered
- (iii) A nanosecond-pulse laser scans rapidly across the entire chip to generate membrane pores in cells and immediately thereafter the elastic chamber is pressurized to deliver cargo through these transient pores (10s)

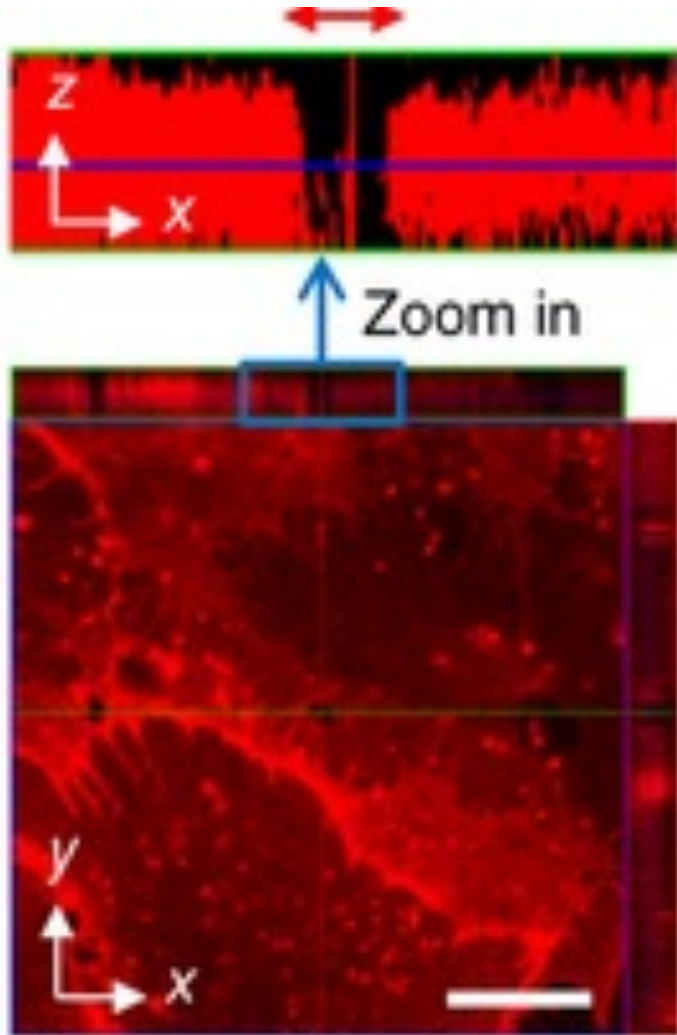
- **Each batch can deliver cargo into 100'000 cells within the chip area (1cm<sup>2</sup>)**
- **Each batch delivery takes 1 min**



# Mechanism of opening transient cell-membrane pores

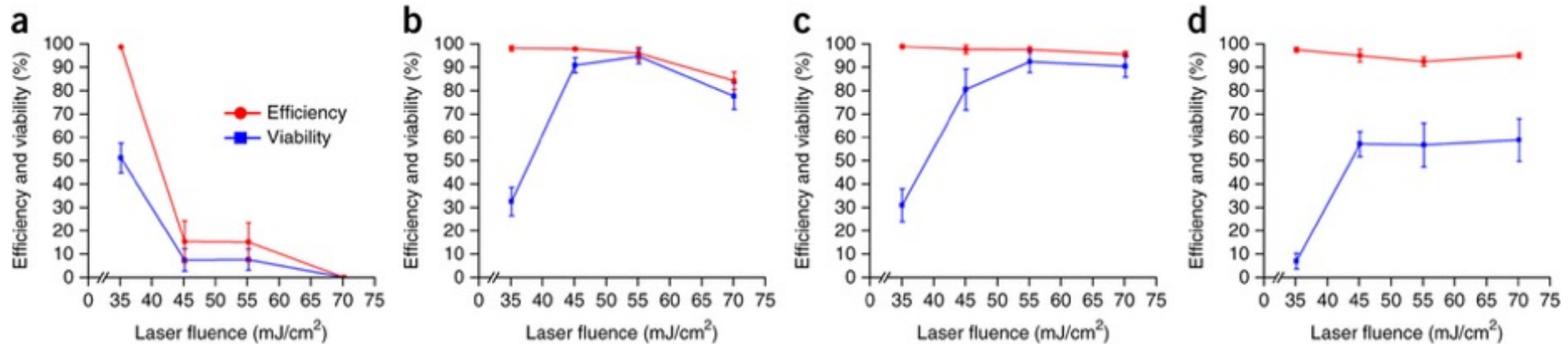


# Rapid bubble bursts are highly localized





# Cargo delivery efficiency and cell viability is dependent on SiO<sub>2</sub> hole density and laser energy



1.15

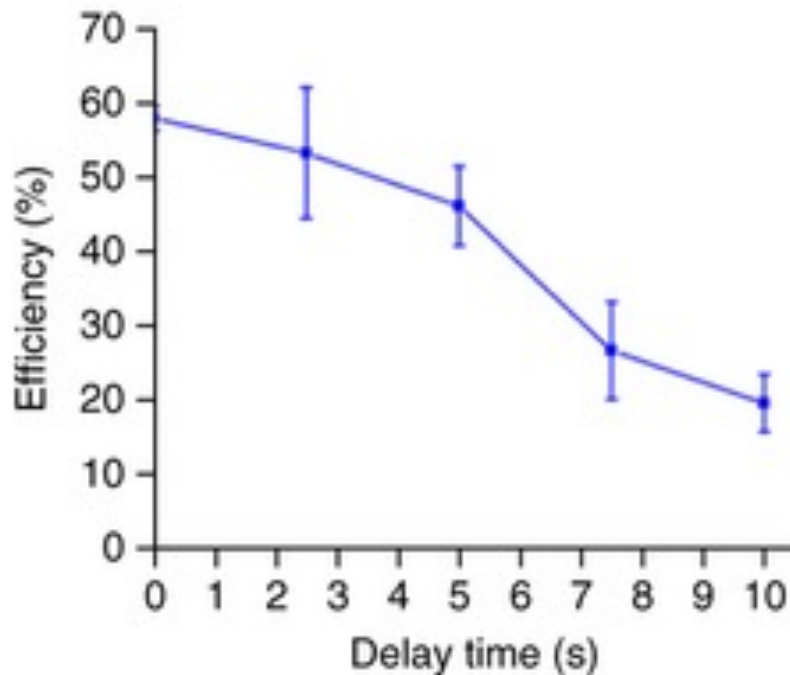
0.28

0.18

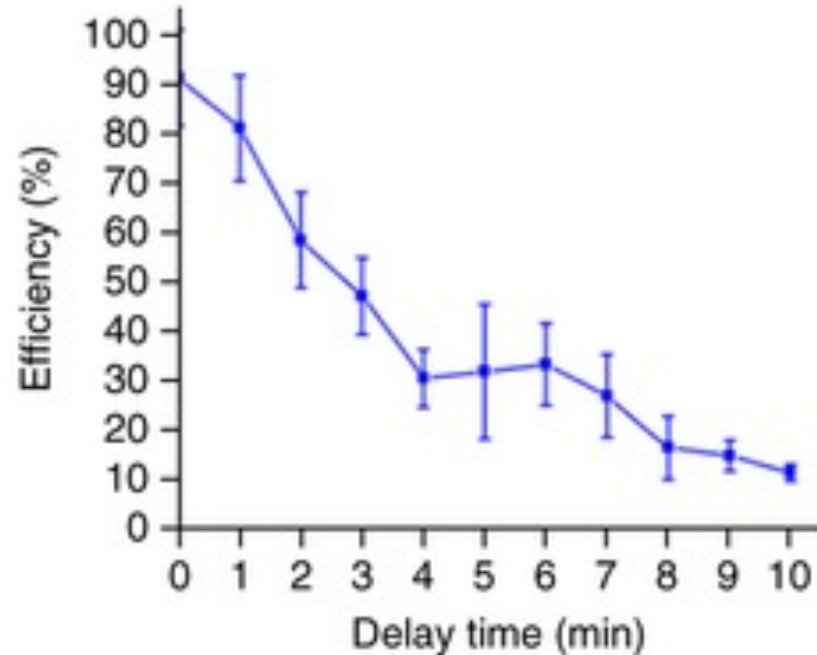
0.12

Holes/10 μm<sup>2</sup>

# Larger cargo tolerates less delivery delay due to rapid pore resealing



**GFP-tagged *F. novicida***  
(ca 1  $\mu\text{m}$  in diameter)

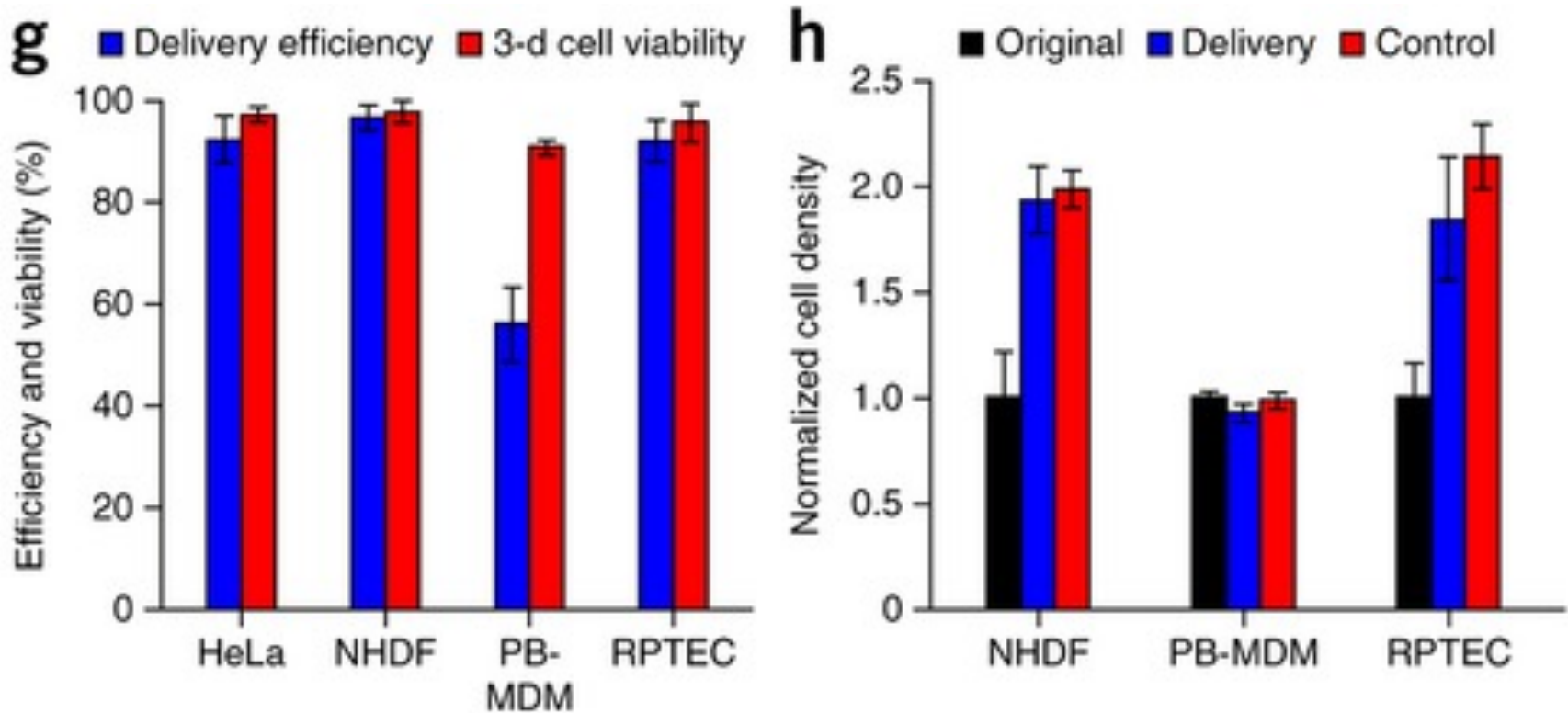


**Calcein**  
(622 Da)

> Complete membrane resealing took more than 10 minutes



# Cargo delivery does not affect neither 3-day cell viability nor cell proliferation



human peripheral blood monocyte–derived macrophages (**PB-MDMs**)  
primary normal human dermal fibroblasts (**NHDFs**)  
human primary renal proximal tubule epithelial cells (**RPTECs**)  
one cancer cell line (**HeLa**)

**Efficient delivery of**

a) 2  $\mu\text{m}$  fluorescent polystyrene beads

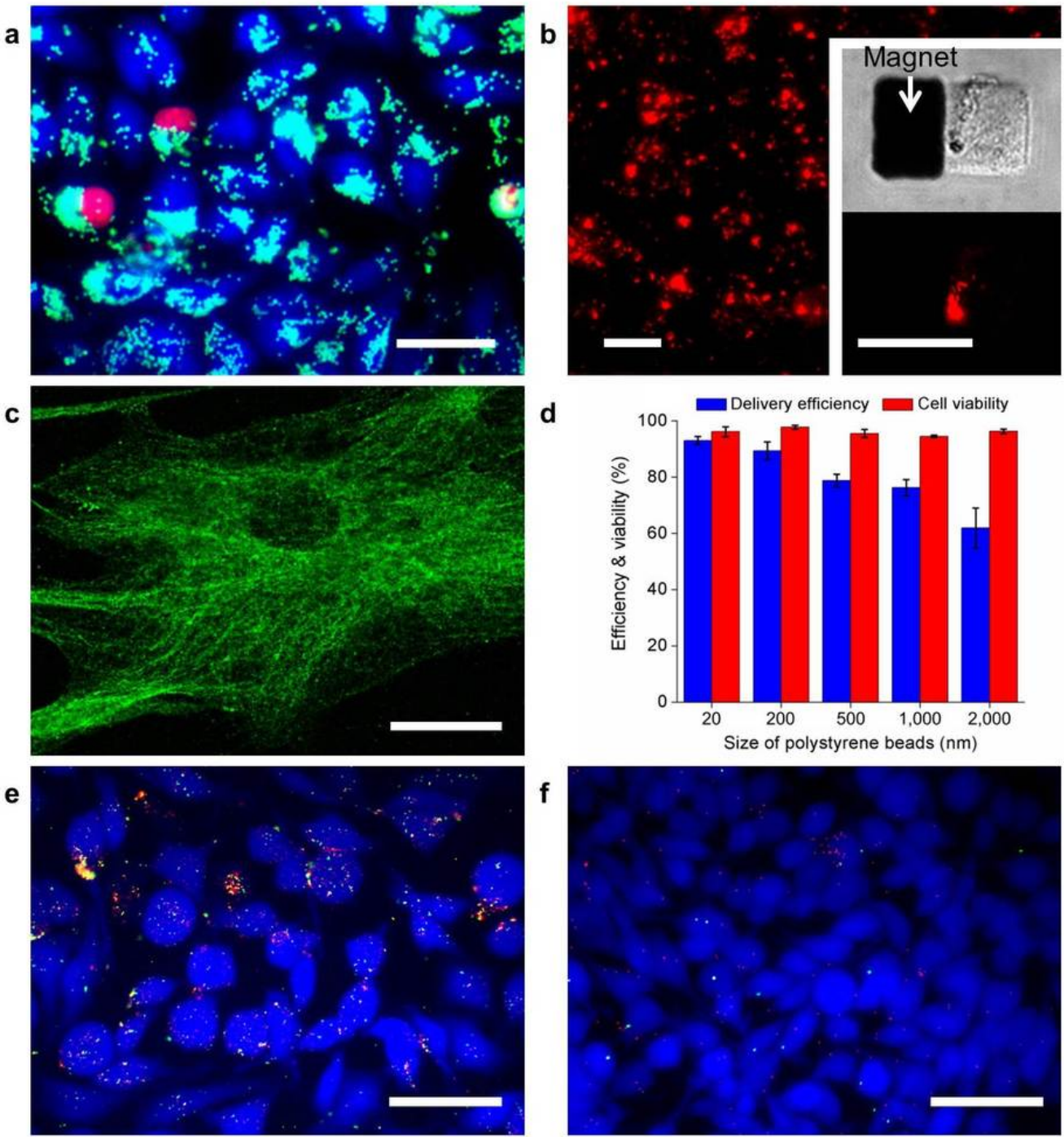
b) 200 nm magnetic beads

c) anti-tubulin AB

d) Delivery efficiency of 5 differentially sized polystyrene beads

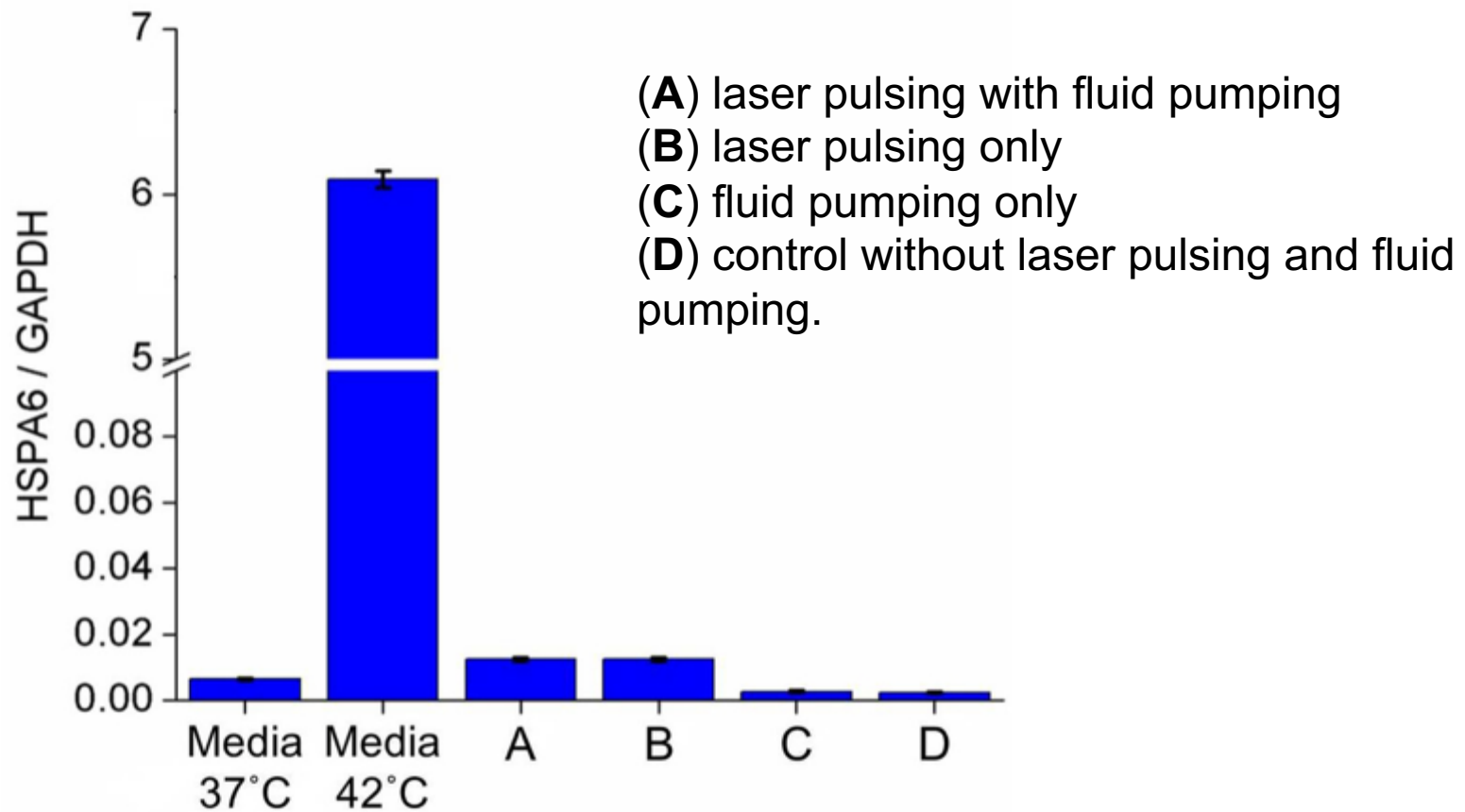
e) co-delivery of 100&600 nm beads

f) Like e) w/o laser pulsing

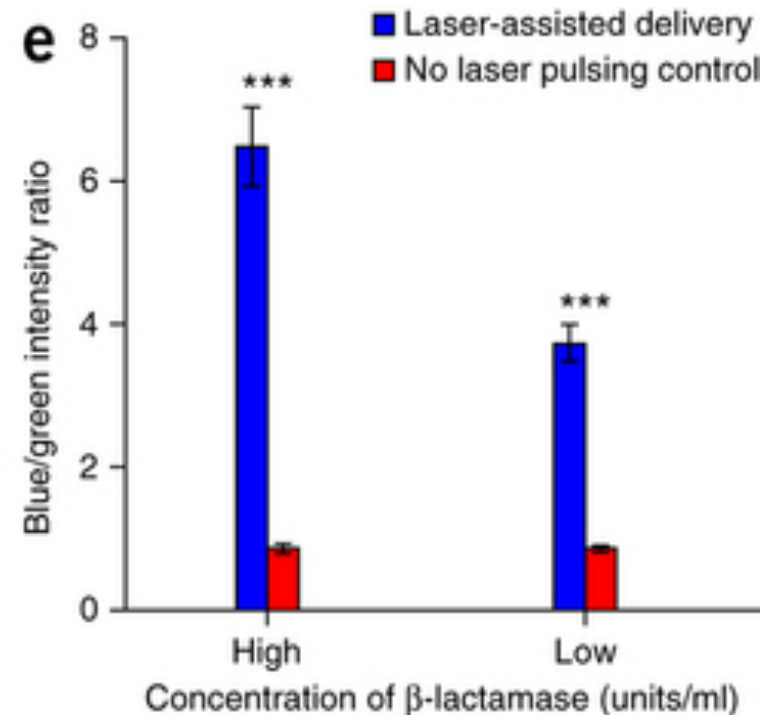
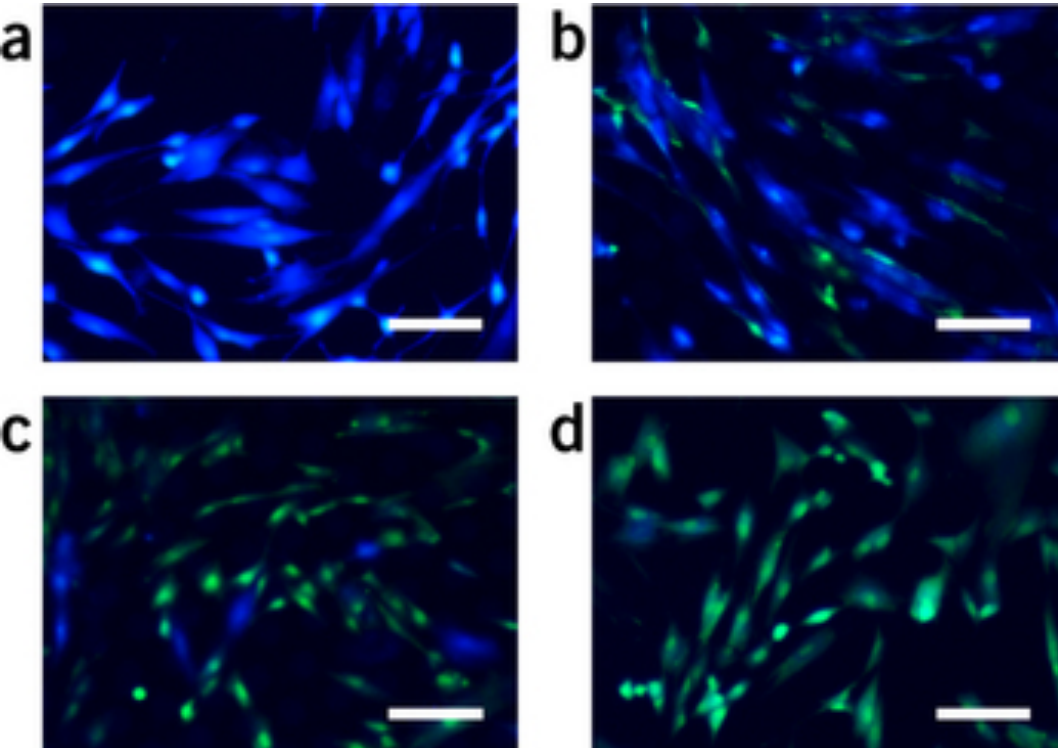
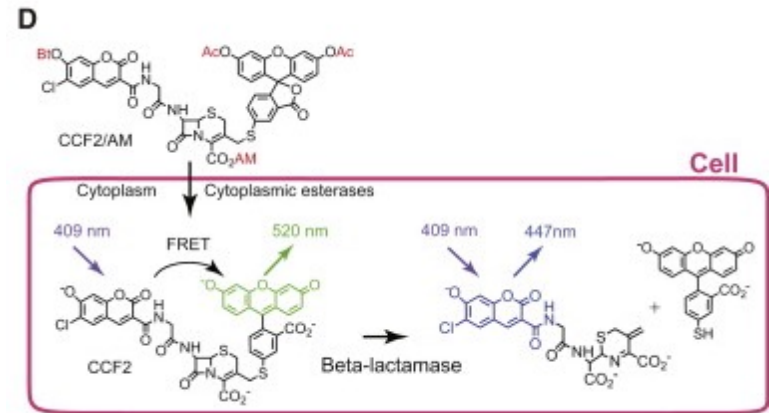


# The general cell stress sensor heat-shock gene HSPA6 is not upregulated by BLAST

Supplementary Figure 5: Evaluation of stress levels of HeLa cells after BLAST delivery.

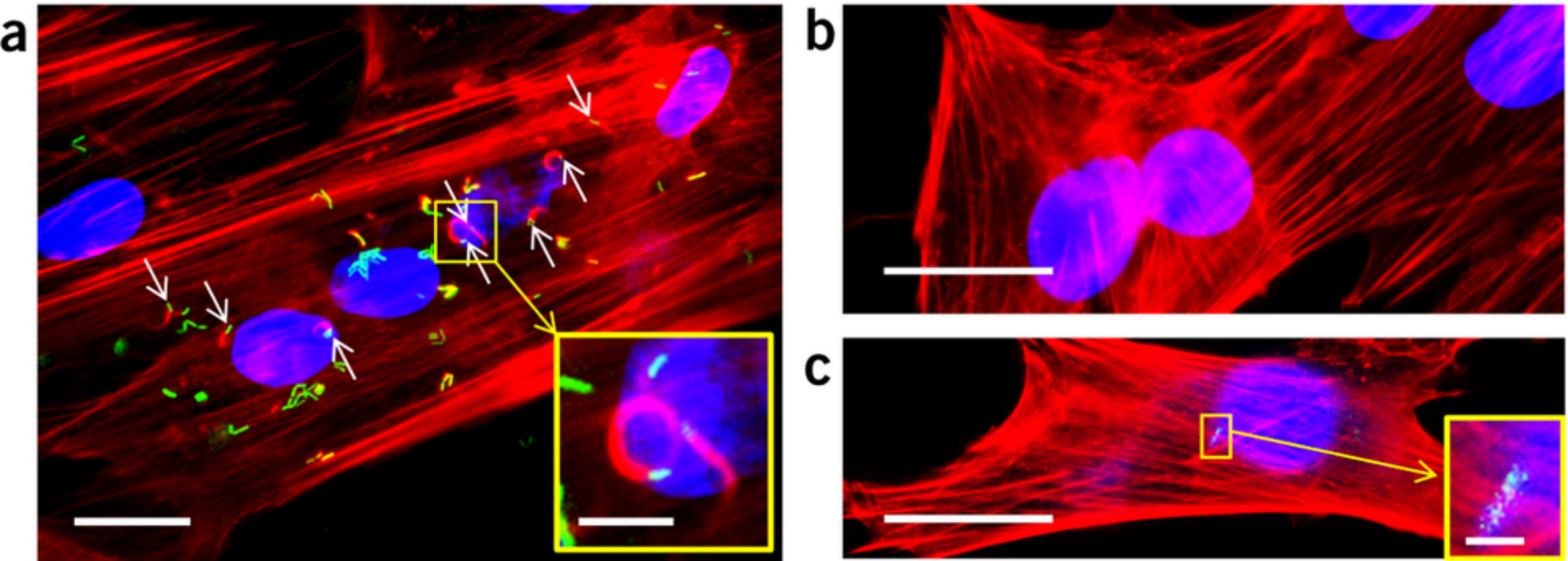


# BLAST facilitates efficient cytosolic delivery

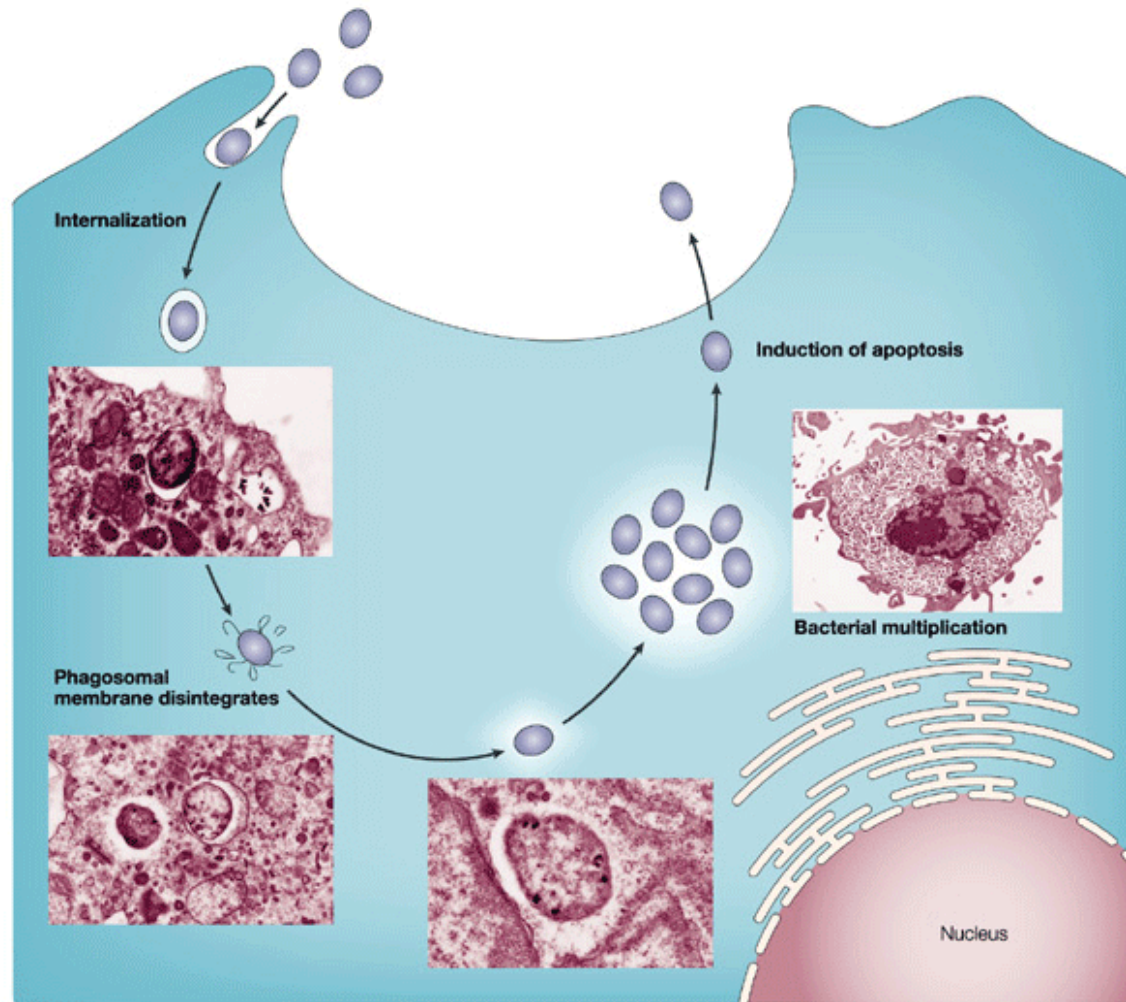




# BLAST-induced uptake of *Listeria monocytogenes* induces actin comet tails



# BLAST allows studying genes important for the replication cycle of *Francisella*





***iglC* is important for cytosolic replication after phagosomal escape in *F. novicida***

