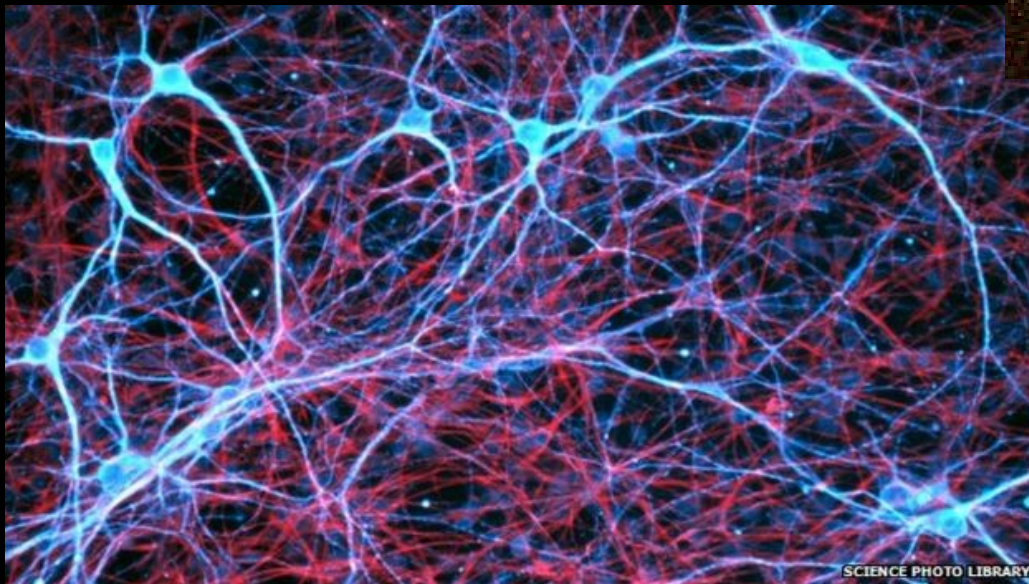
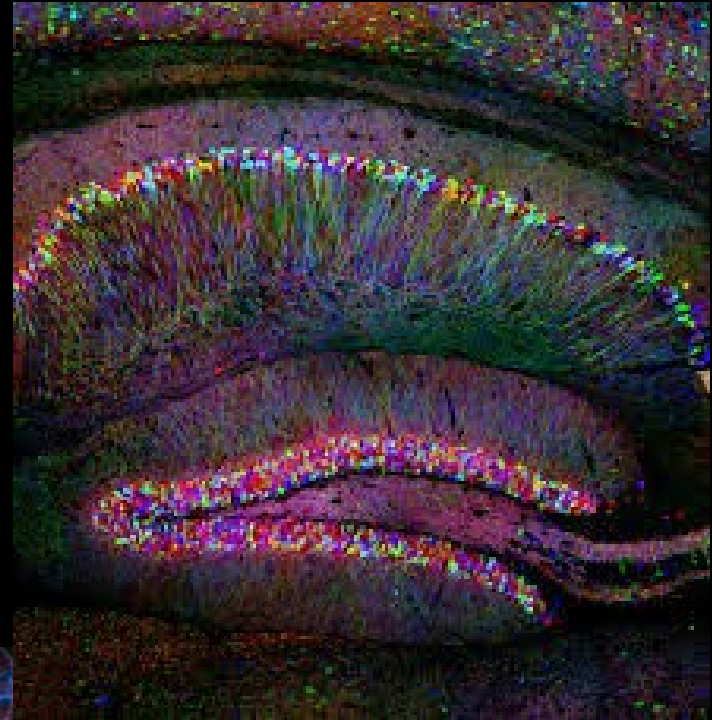


# Imaging Voltage in Neurons

Overview and recent advances

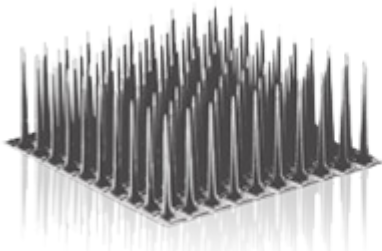
Technical Journal Club  
January 26, 2016  
Angie Wulf

# motivation

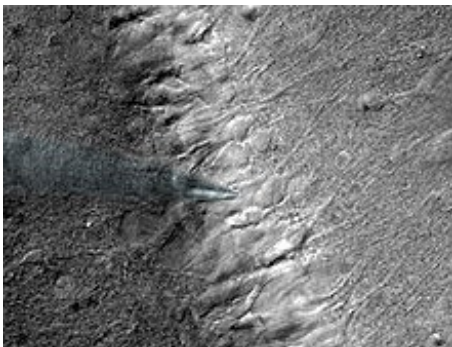


# Methods to measure neuronal activity

- electrodes
  - extracellular: invasive
- calcium imaging

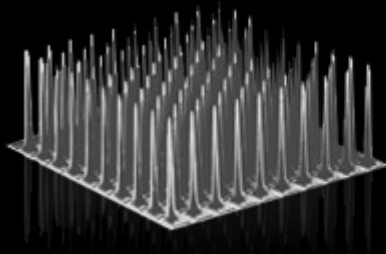


- intracellular: limited to cell soma and large dendrites, few cells

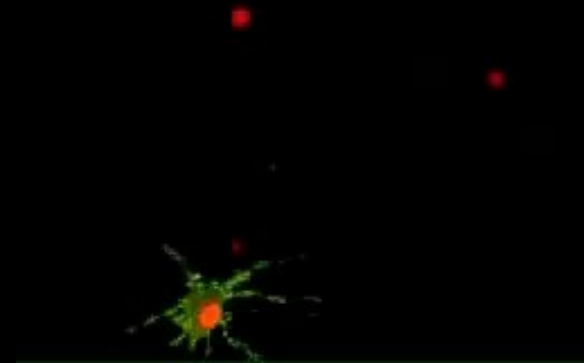


# Methods to measure neuronal activity

- electrodes
  - extracellular: invasive
- calcium imaging
  - Biased to suprathreshold events
  - slow

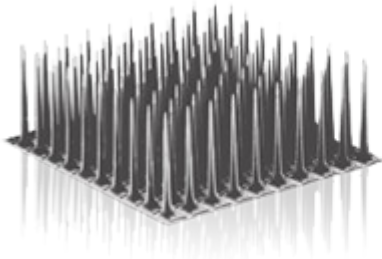


- intracellular: limited to cell soma and large dendrites, few cells

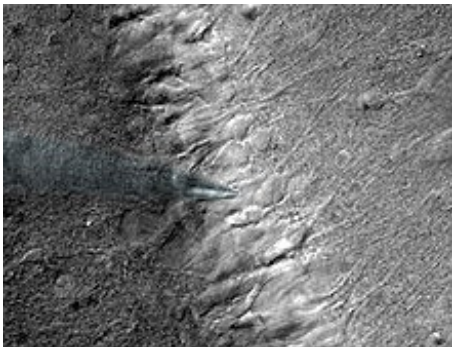


# Methods to measure neuronal activity

- electrodes
  - extracellular: invasive



- intracellular: limited to cell soma and large dendrites, few cells



- calcium imaging
  - Biased to suprathreshold events
  - slow

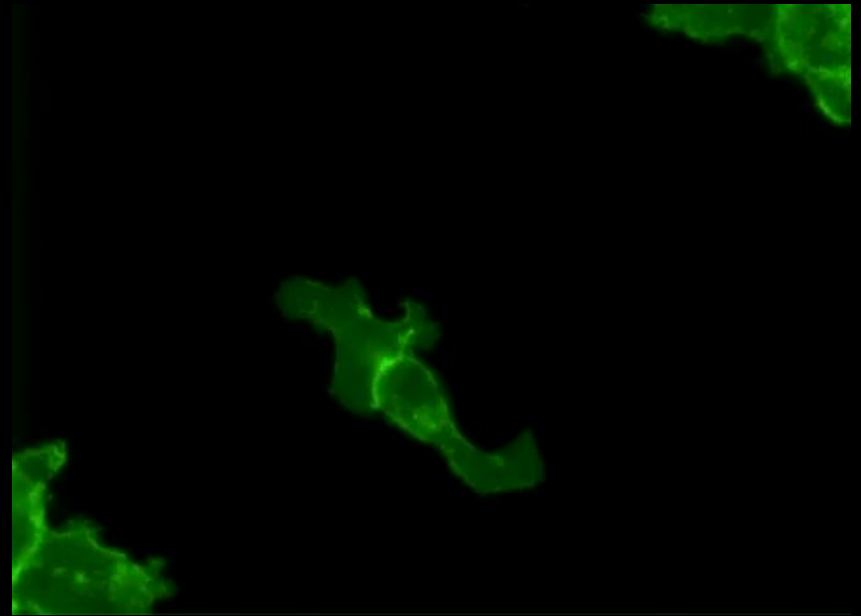
- voltage imaging
  - all events
  - all intracellular compartments
  - many cells

# Definition: voltage indicator

- indicator that changes its fluorescence upon changes of membrane voltage

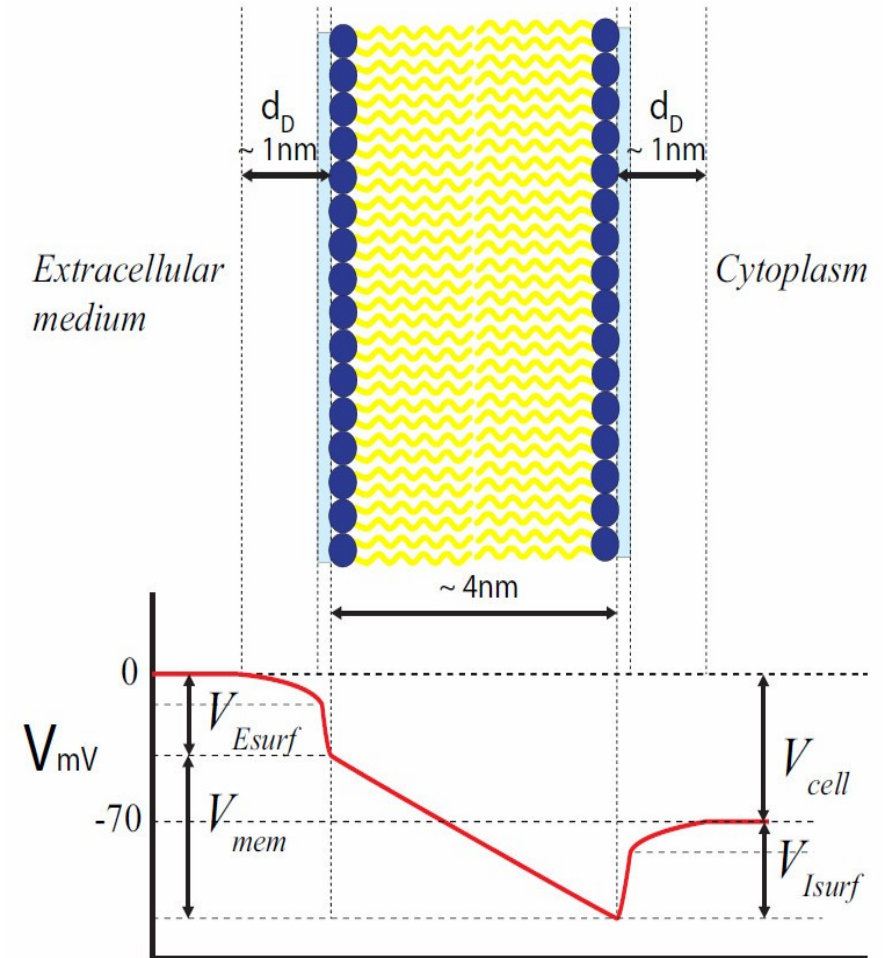
# Definition: voltage indicator

- indicator that changes its fluorescence upon changes of membrane voltage



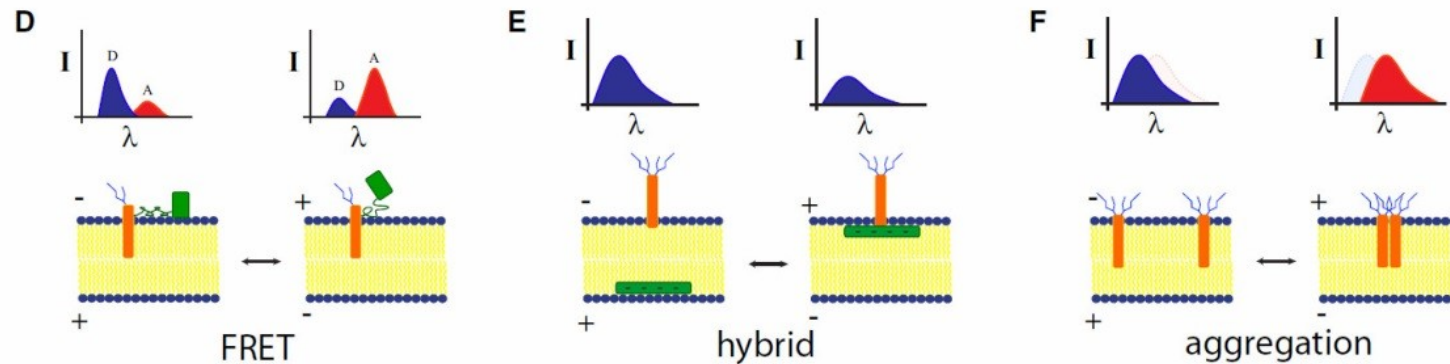
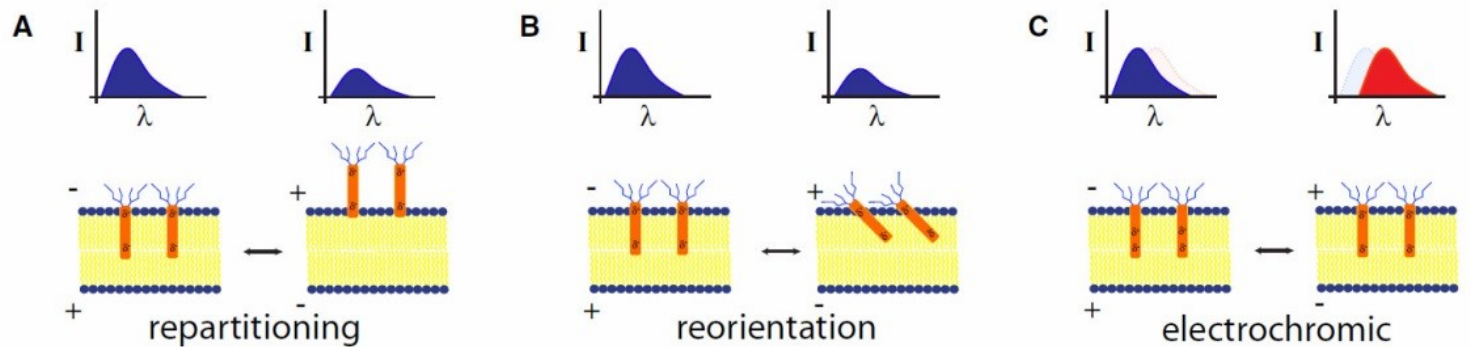
# Requirements

- Challenges
  - Membrane properties
  - (photo-) toxicity
  - Biological activity
- An ideal voltage indicator is
  - Sensitive & bright with good S/N ratio
  - Fast
  - Non-toxic
  - Biologically „inert“



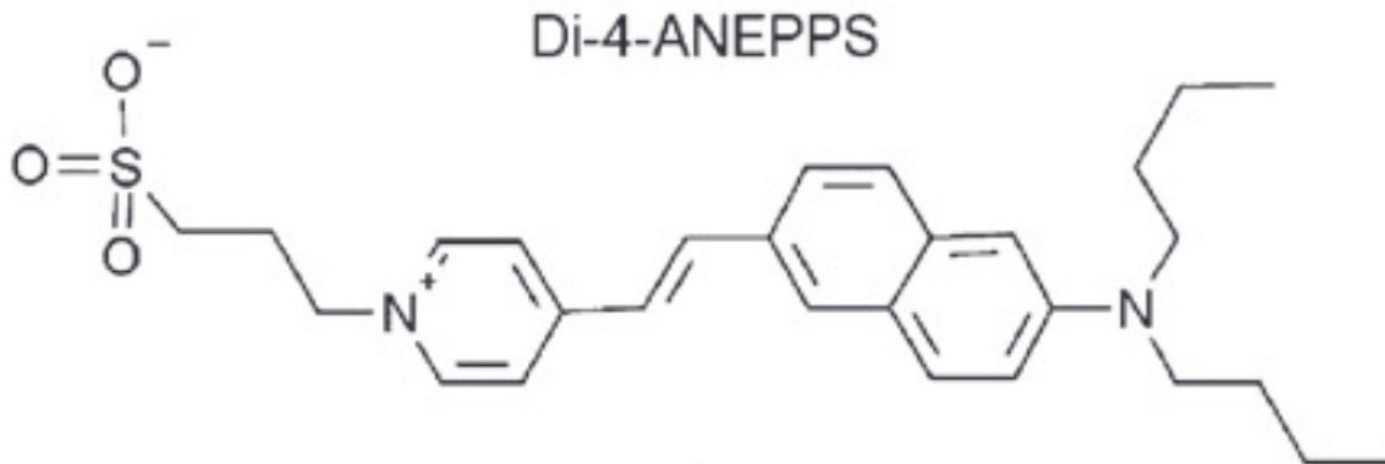
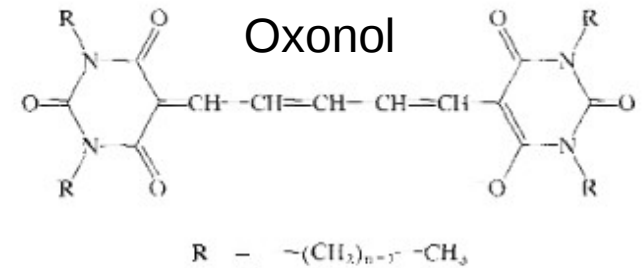


# Mechanisms of voltage sensing

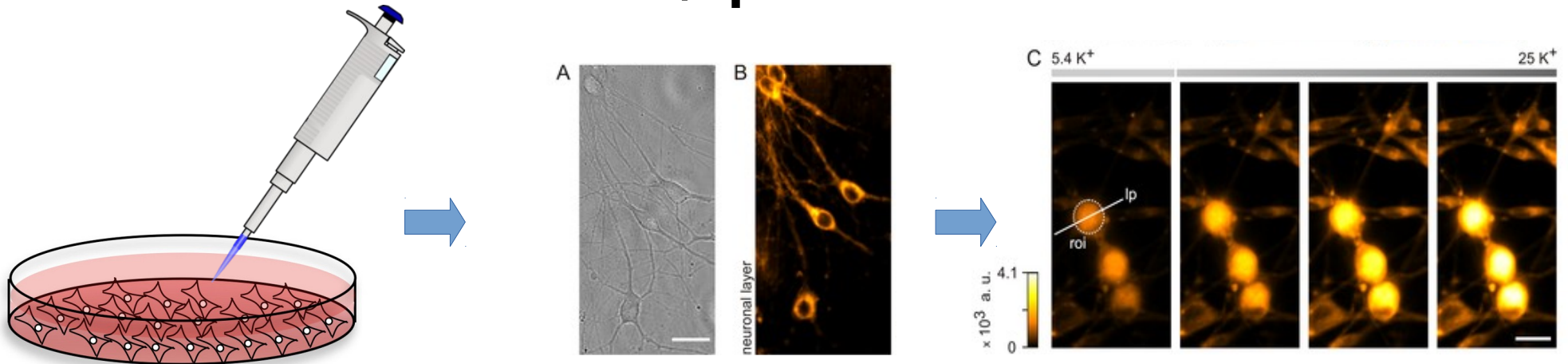


# Voltage Sensitive Dyes (VSDs)

- Organic molecules
- Enrich in the membrane
- Change absorption or emission spectra upon changes in the electric field around them



# VSDs: use, pro's and con's



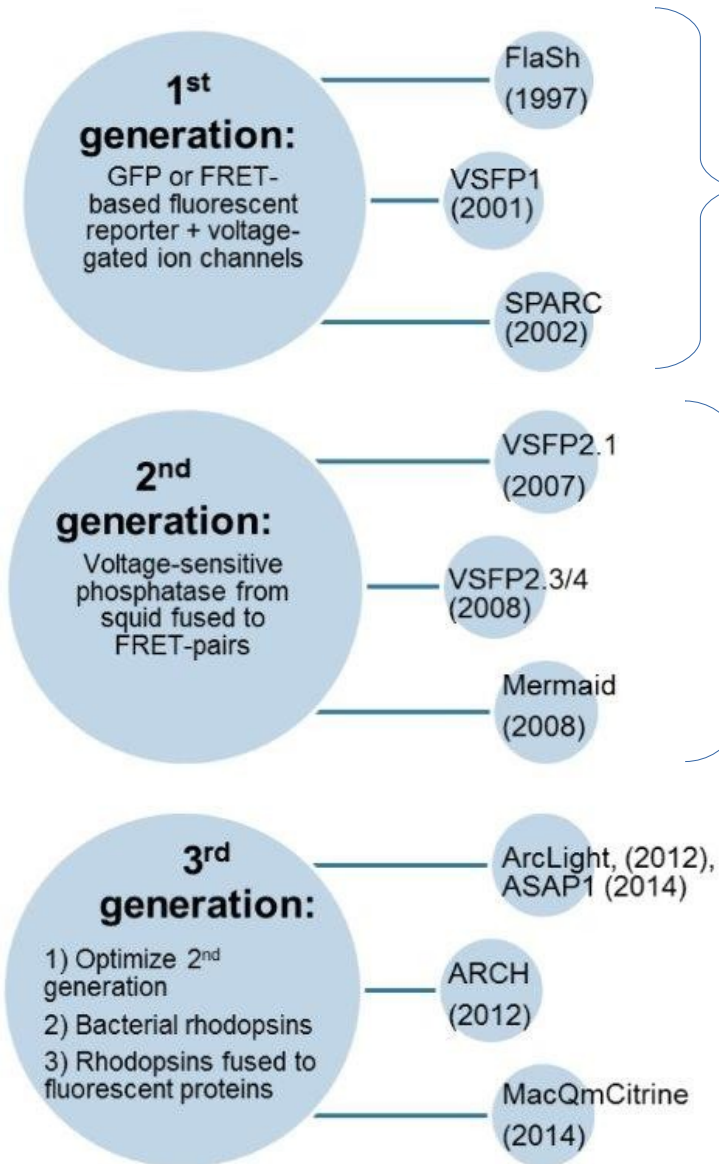
- simple to use
- fast kinetics



- some cells hard to label
- only transient
- no targeting, unspecific labeling

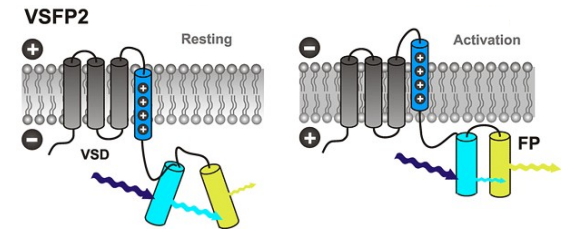
=> excellent for single use where cell labelling is not so important

# Genetically Encoded Voltage Indicators



No expression on the plasma membrane in mammalian cells

Work well in mammalian cells but  
- small signals  
- slow



# Paper 1

[Front Cell Neurosci.](#) 2015 Apr 24;9:147. doi: 10.3389/fncel.2015.00147. eCollection 2015.

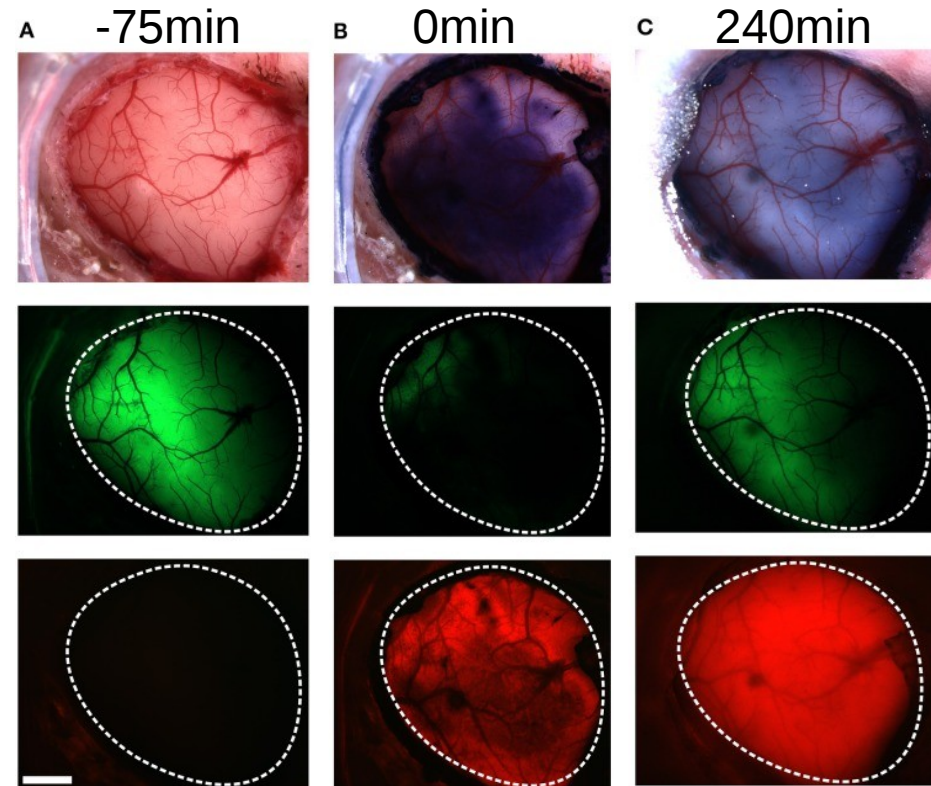
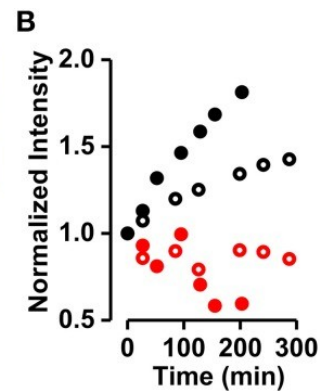
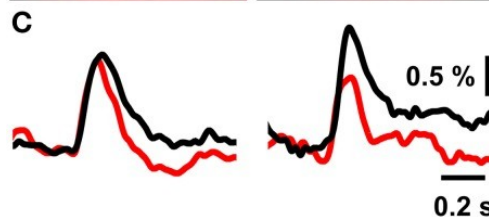
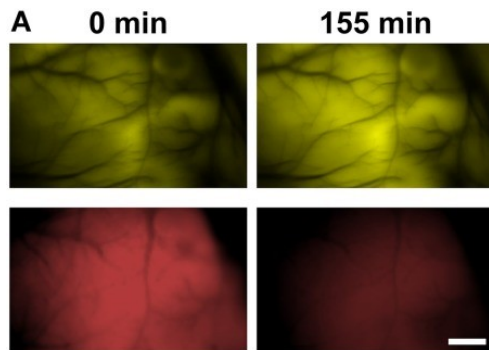
**Comparative performance of a genetically-encoded voltage indicator and a blue voltage sensitive dye for large scale cortical voltage imaging.**

[Mutoh H](#)<sup>1</sup>, [Mishina Y](#)<sup>2</sup>, [Gallero-Salas Y](#)<sup>3</sup>, [Knöpfel T](#)<sup>4</sup>.



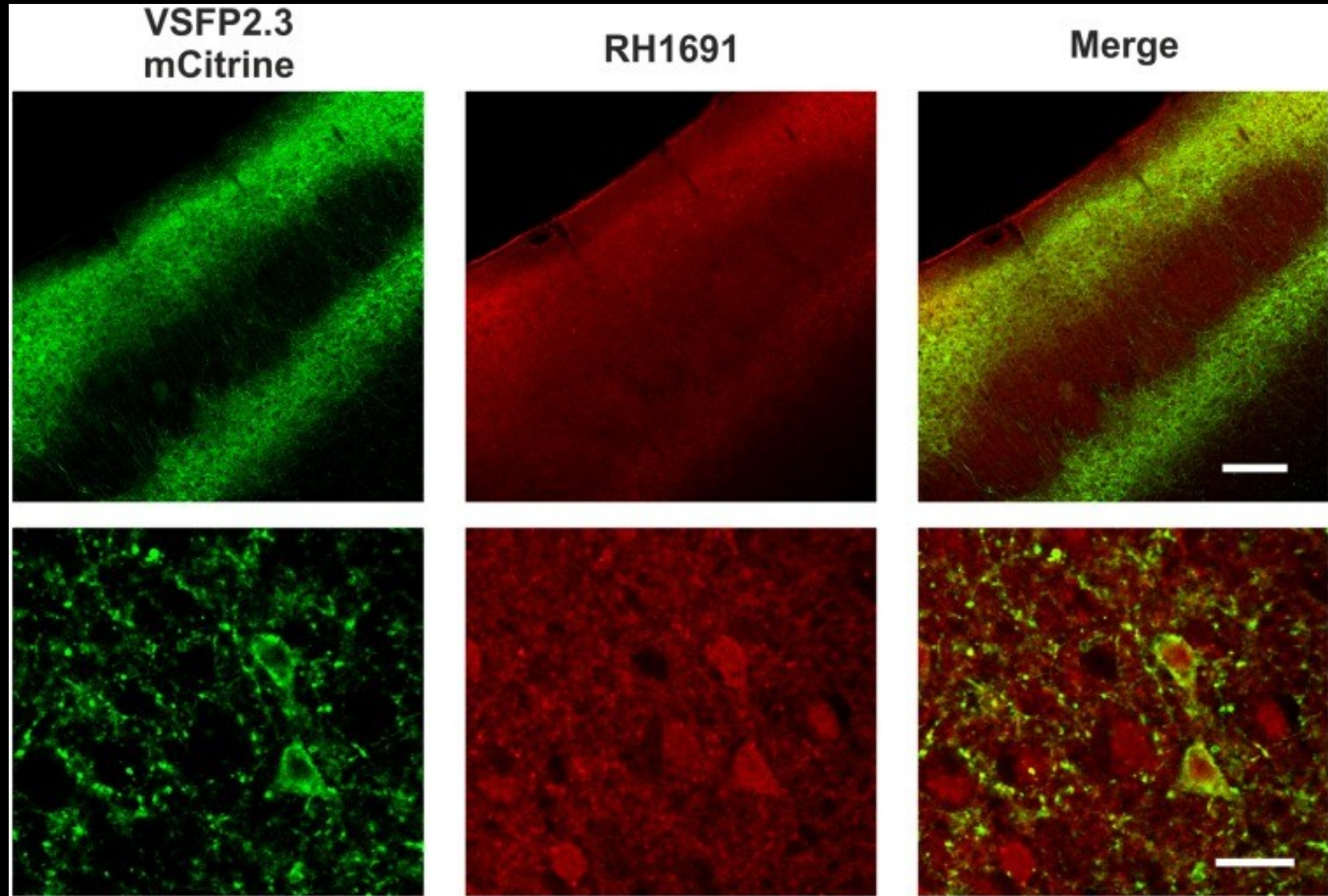
# staining and imaging procedure

- In utero electroporation of VSFP2
- VSFP2 imaging through intact dura
- dye staining for 1h, 15min wash



Scale bar: 1mm

# Post-fixation imaging

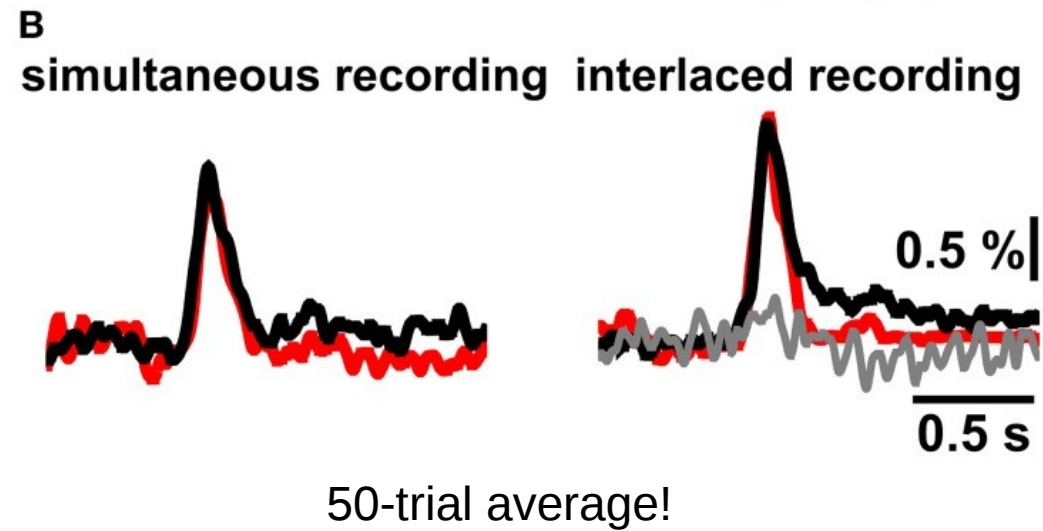
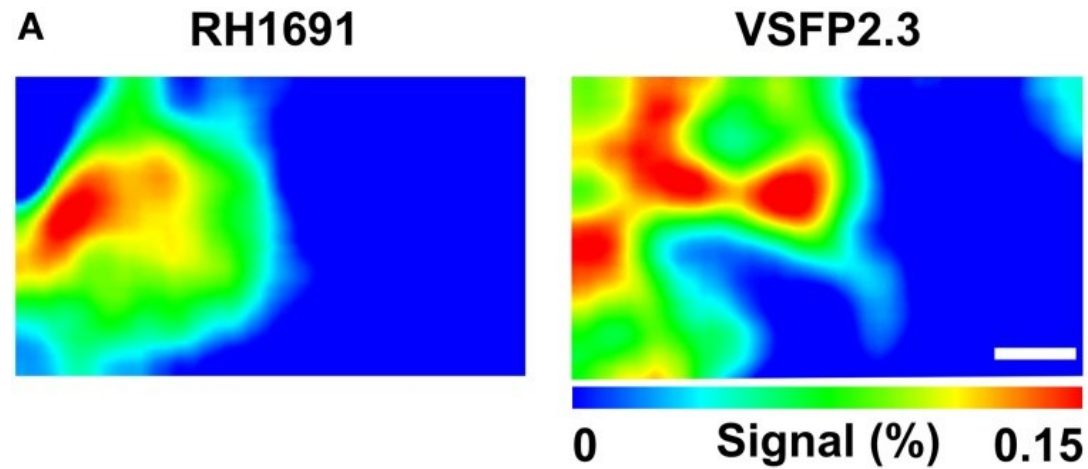


=> VSD labels all cells while GEVI is specific

Scale bar  
Top: 200um  
Bottom: 20um

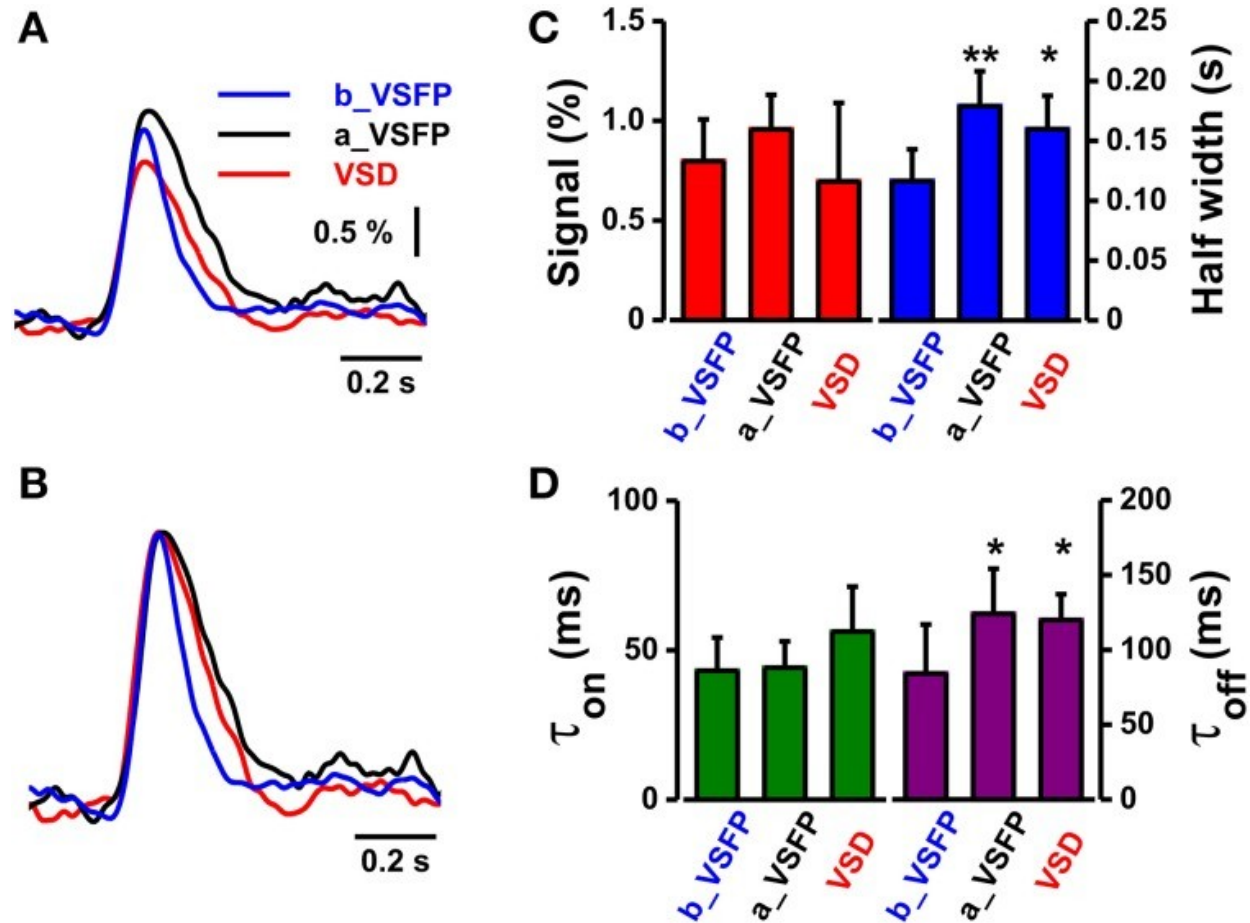


# direct comparison I



=> no difference

# direct comparison II



=> VSFP is better before opening the dura, after, both are same

# Paper 1: discussion and conclusion

- VSFP2 and RH1691 show similar responses to whisker deflection
- VSFP2 is faster if dura/skull is left intact
- GEVIs have many advantages
  - more stable signal
  - targeting
  - non-invasive imaging conditions
  - chronic imaging

# Paper 2

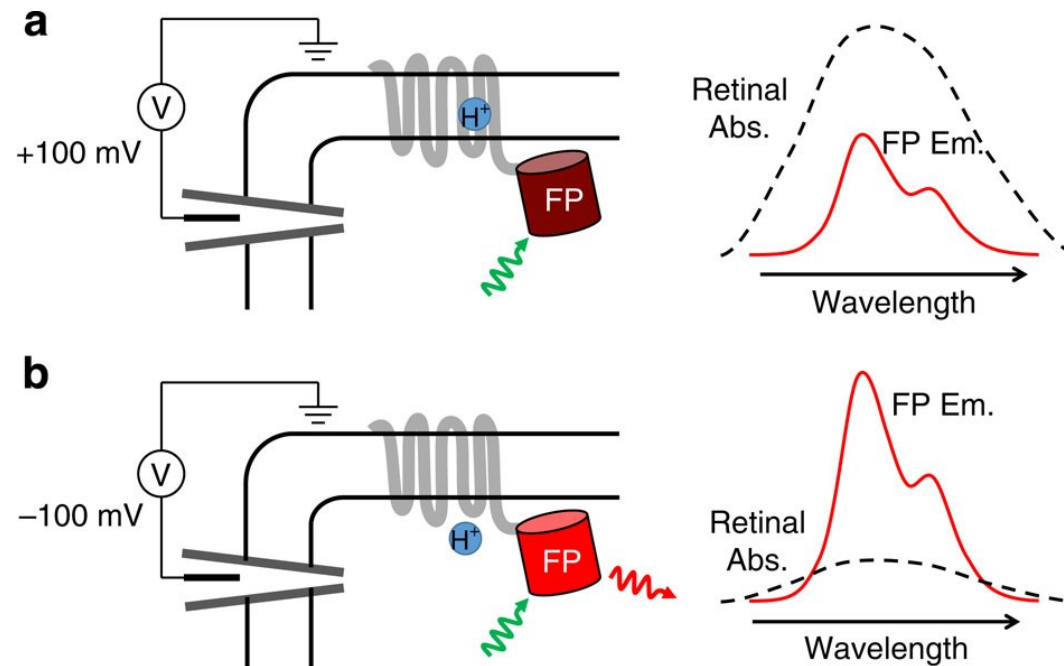
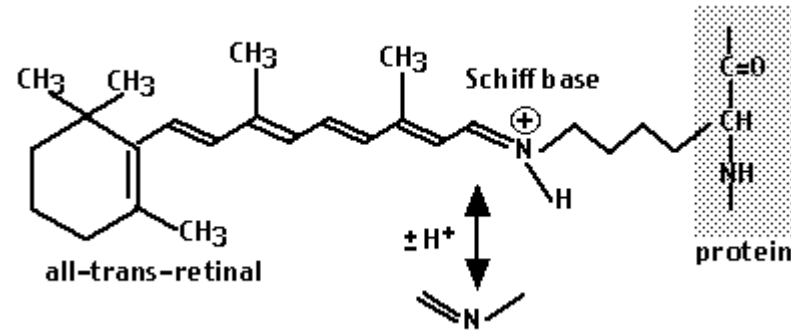
Science. 2015 Dec 11;350(6266):1361-6. doi: 10.1126/science.aab0810. Epub 2015 Nov 19.

**High-speed recording of neural spikes in awake mice and flies with a fluorescent voltage sensor.**

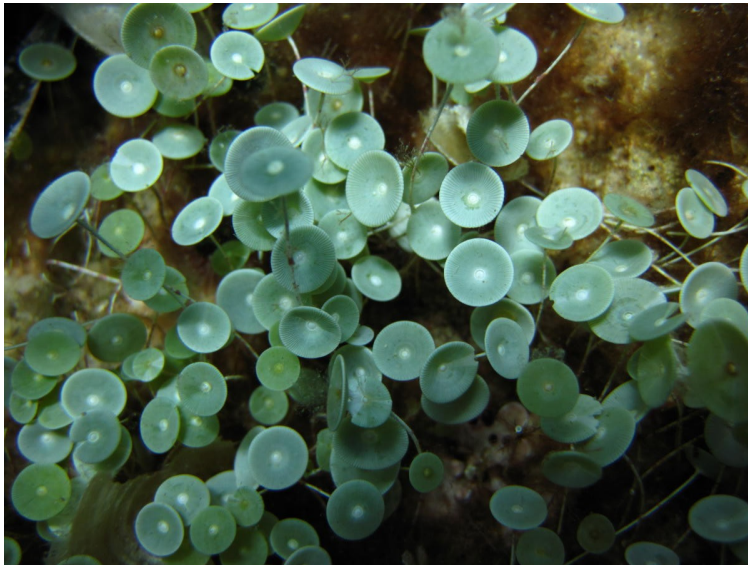
Gong Y<sup>1</sup>, Huang C<sup>2</sup>, Li JZ<sup>3</sup>, Grewe BF<sup>3</sup>, Zhang Y<sup>4</sup>, Eismann S<sup>3</sup>, Schnitzer MJ<sup>5</sup>.

# indicator mechanism: eFRET

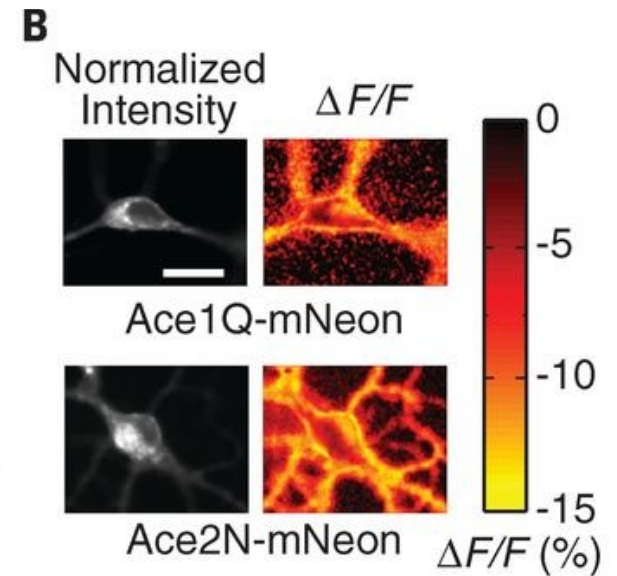
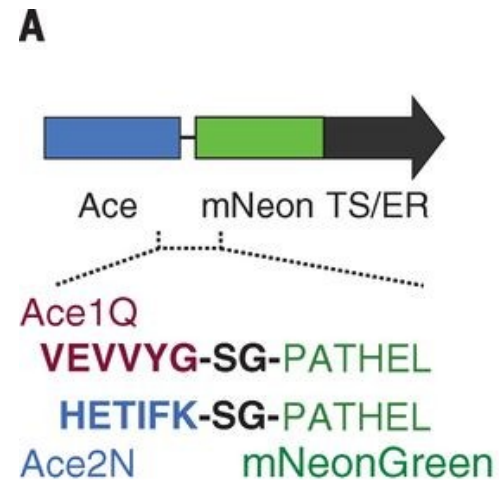
rhodopsin



# indicator generation & characterization I



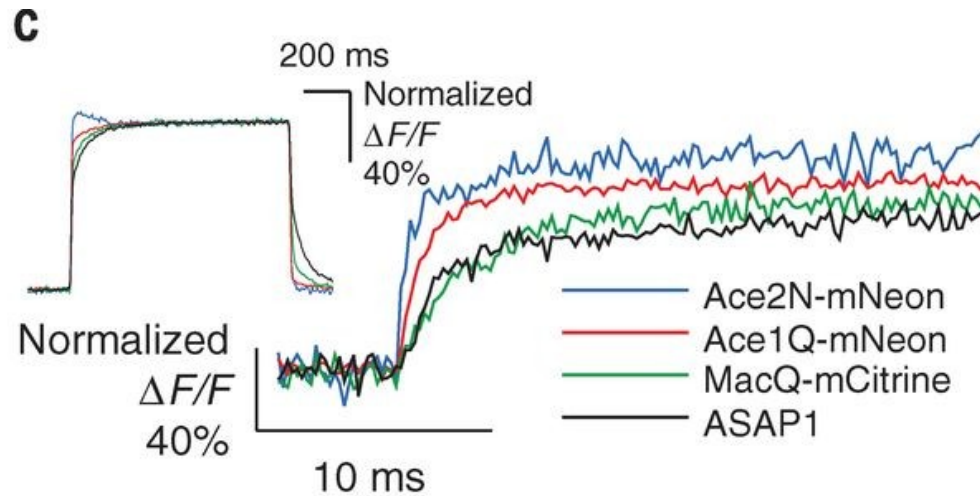
Acetabularia acetabulum



=> high FRET-efficiency  
=> little intracellular aggregation

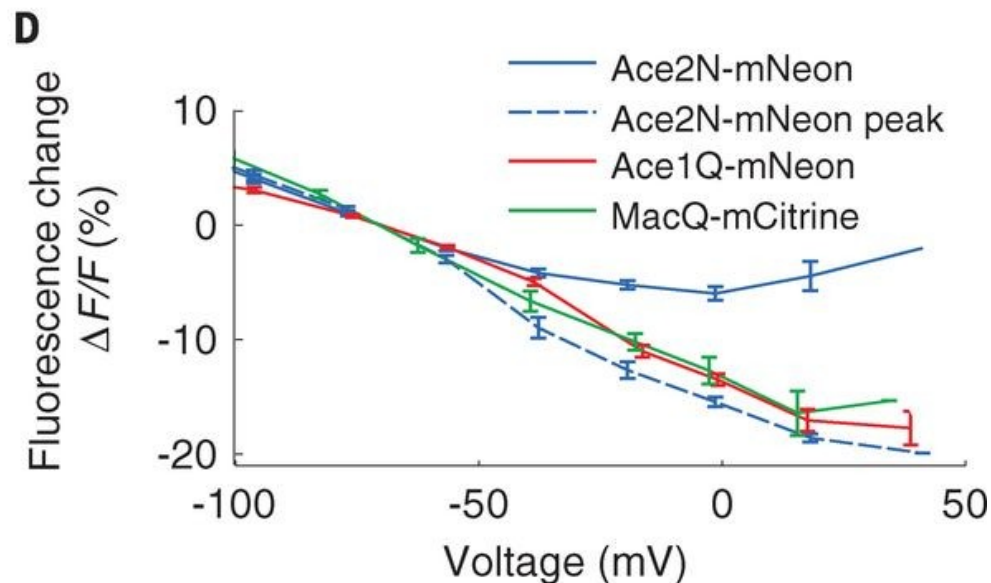
HEK293 cells  
100mV depolarization step

# indicator generation & characterization I



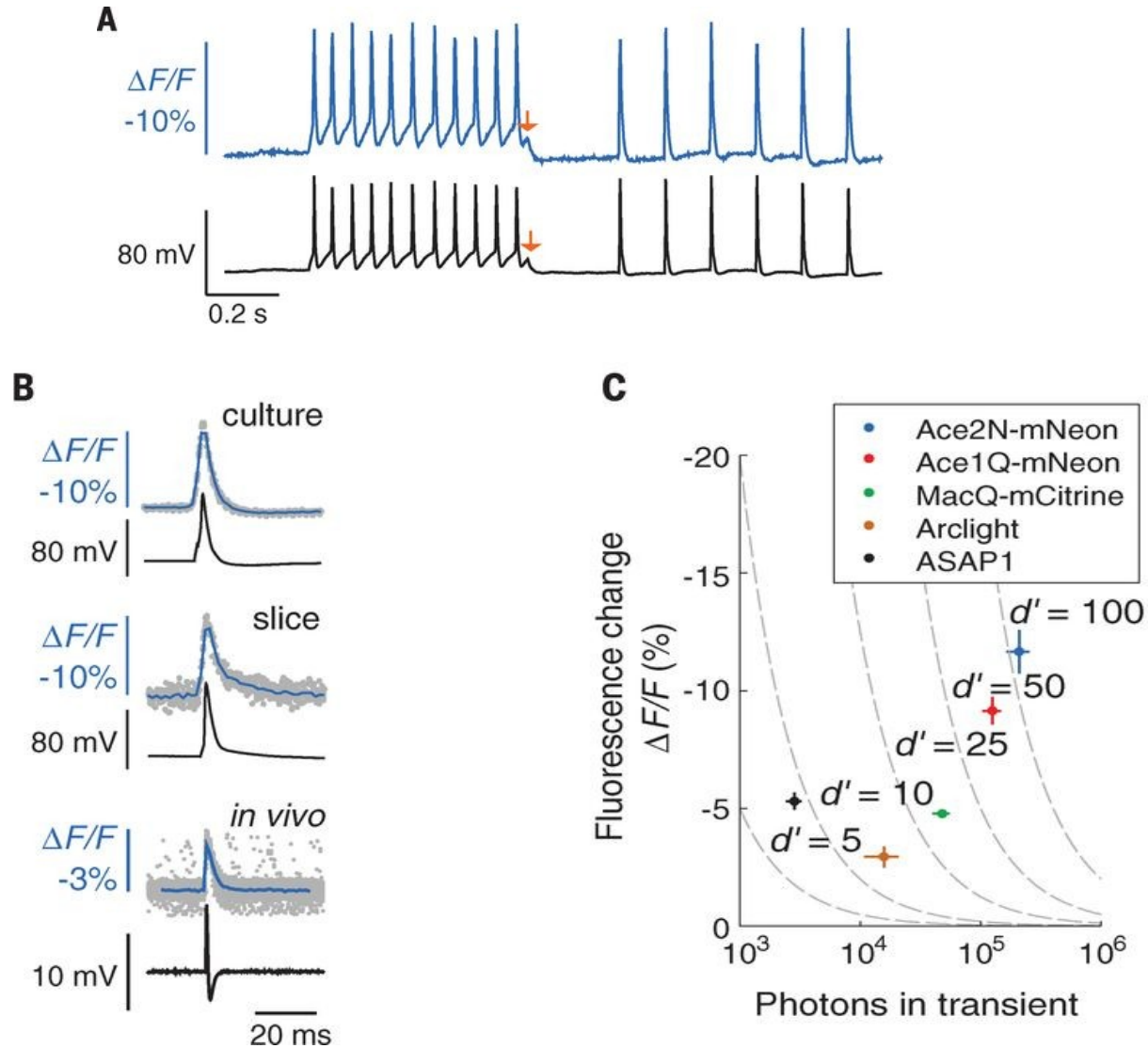
=> <1ms response time

=> linear relationship between membrane voltage and signal



HEK293 cells  
100mV depolarization step

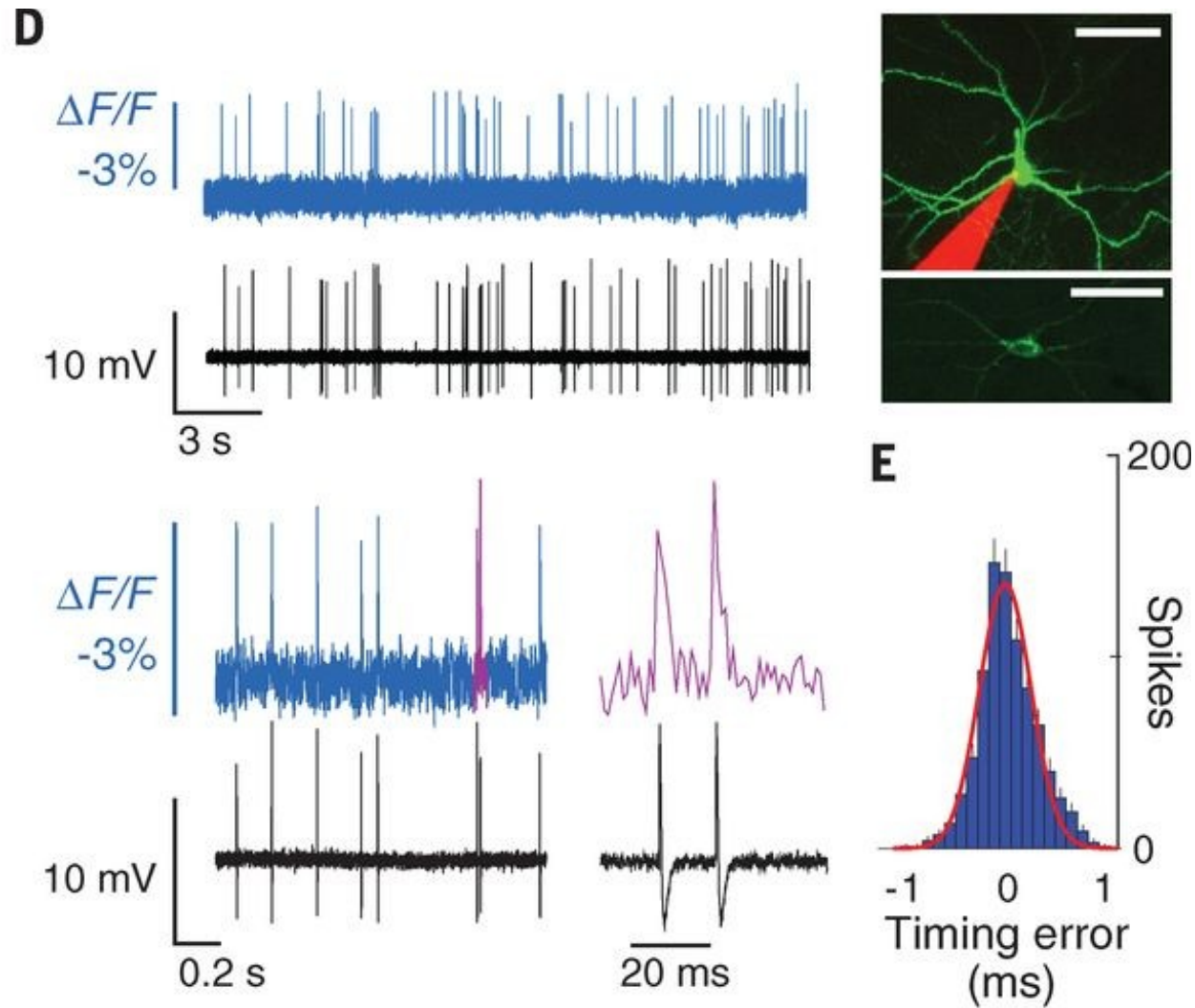
# performance assessment I



=> 3-10fold better spike detection fidelity



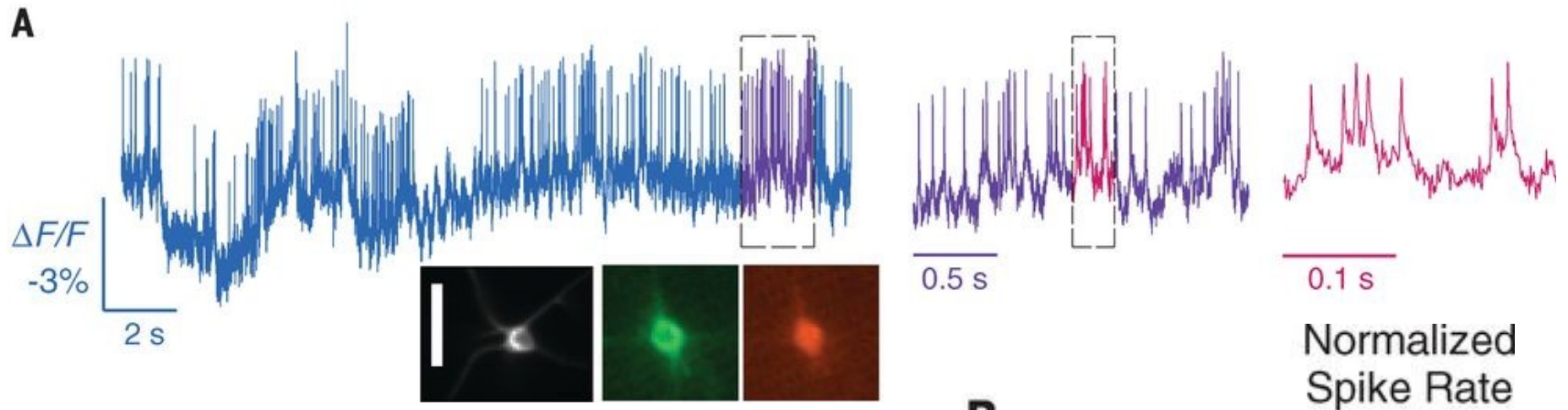
# performance assessment II



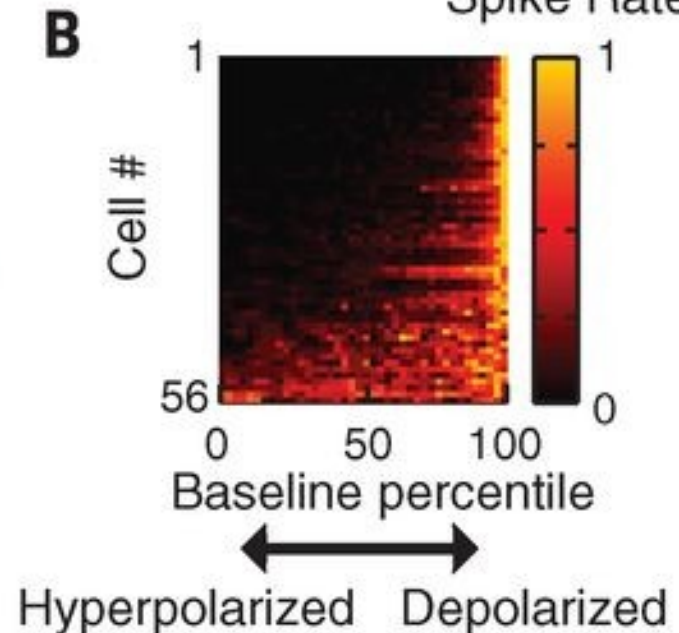
Anesthetized  
mouse,  
cortical  
neuron

=> detection of fast spiking with small timing error

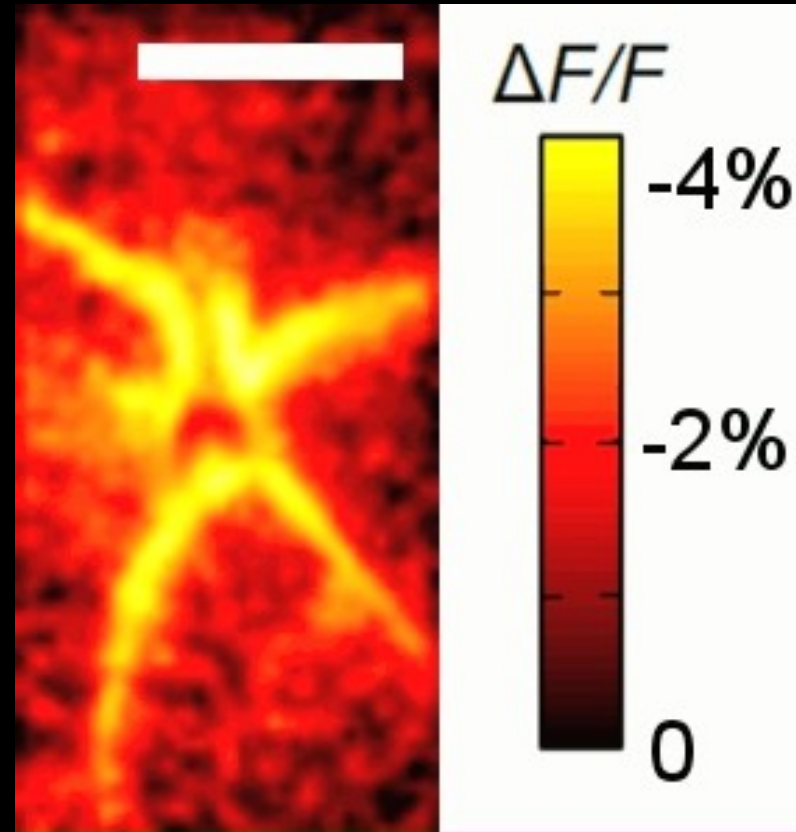
# *in vivo* recordings awake mouse



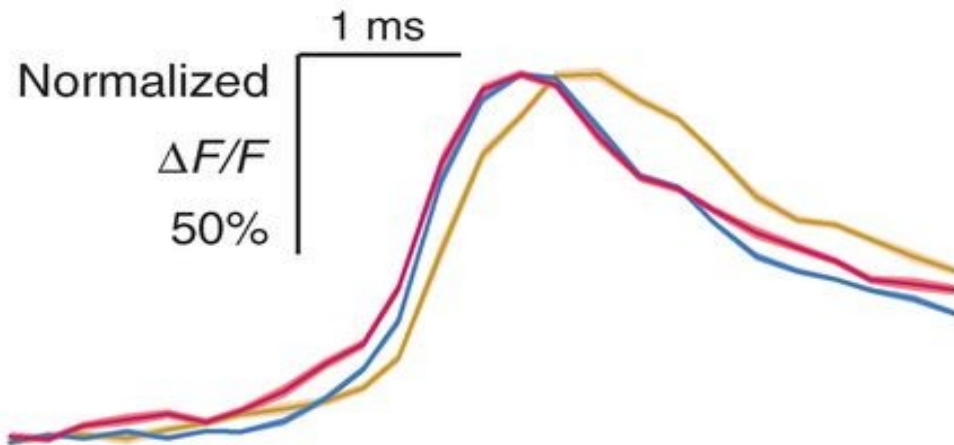
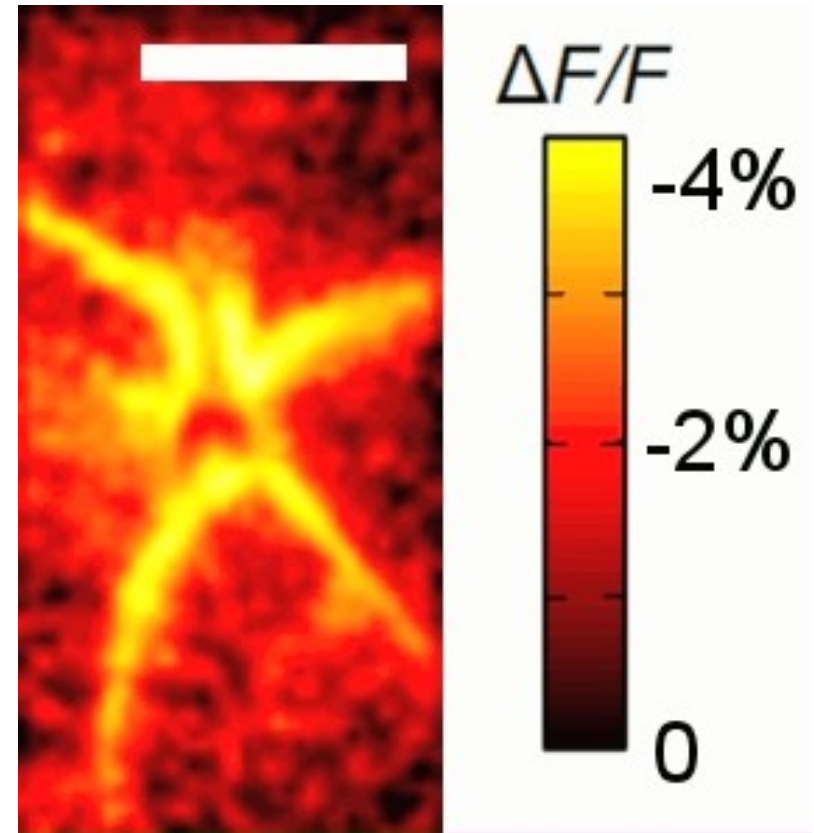
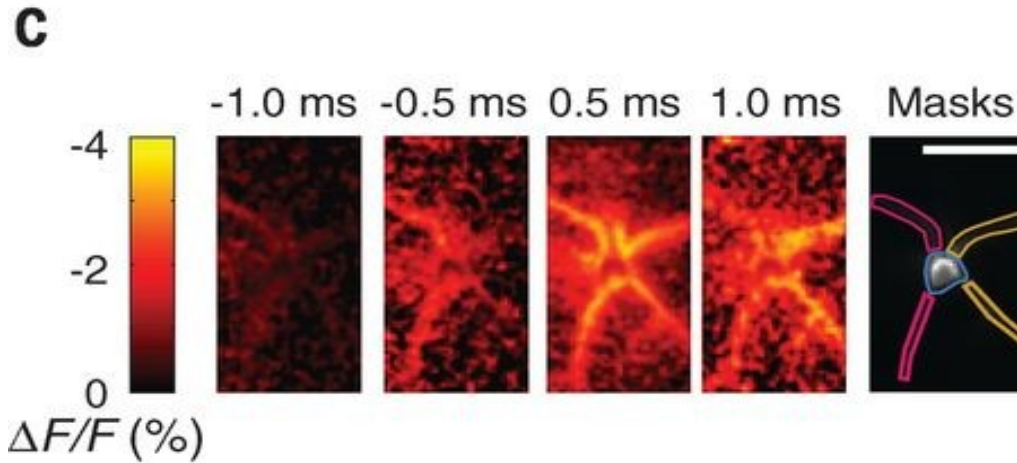
=> signals can be recorded from awake mice  
=> if Ace reports hyperpolarization, there is less spiking



# *in vivo* recordings awake mouse



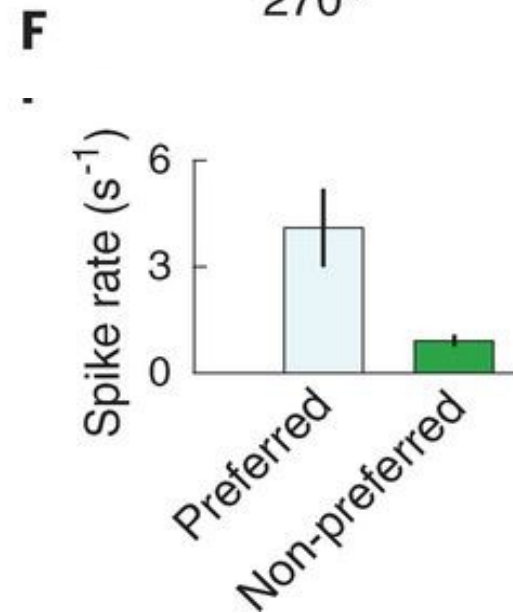
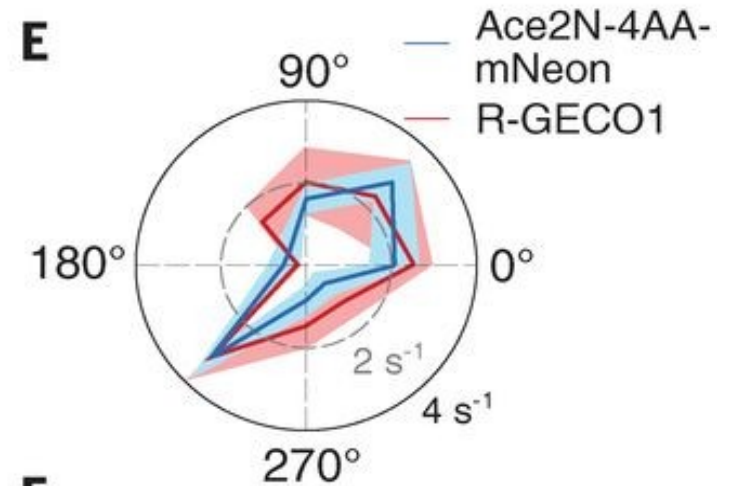
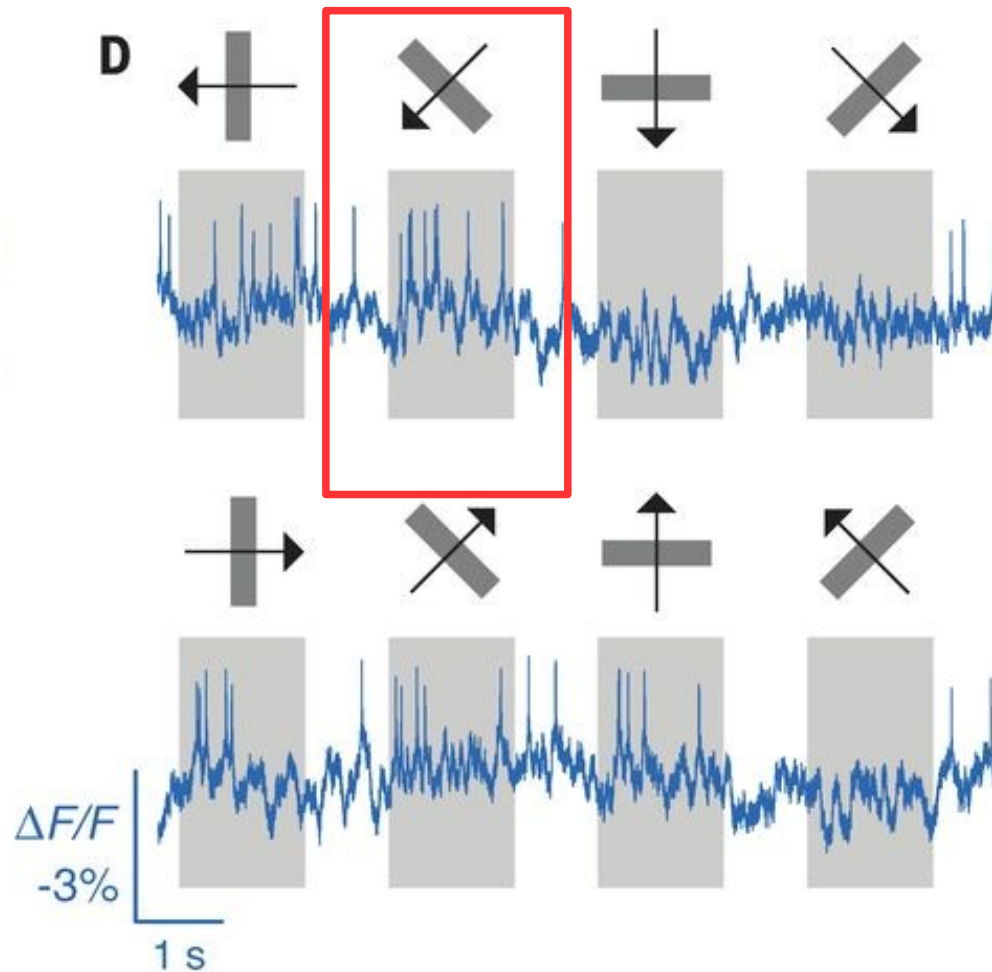
# *in vivo* recordings awake mouse



1900 spikes averaged

=> subcellular resolution possible

# *in vivo* recordings awake mouse



Visually evoked responses of a V1 cortical neuron to moving gratings (10 trials per stimulus)

# Conclusion paper 2

- New indicator outperforms
  - other published GEVIs
  - Calcium indicators
- But
  - only in sparse labelling
  - => development of new imaging techniques is required

# Paper 3

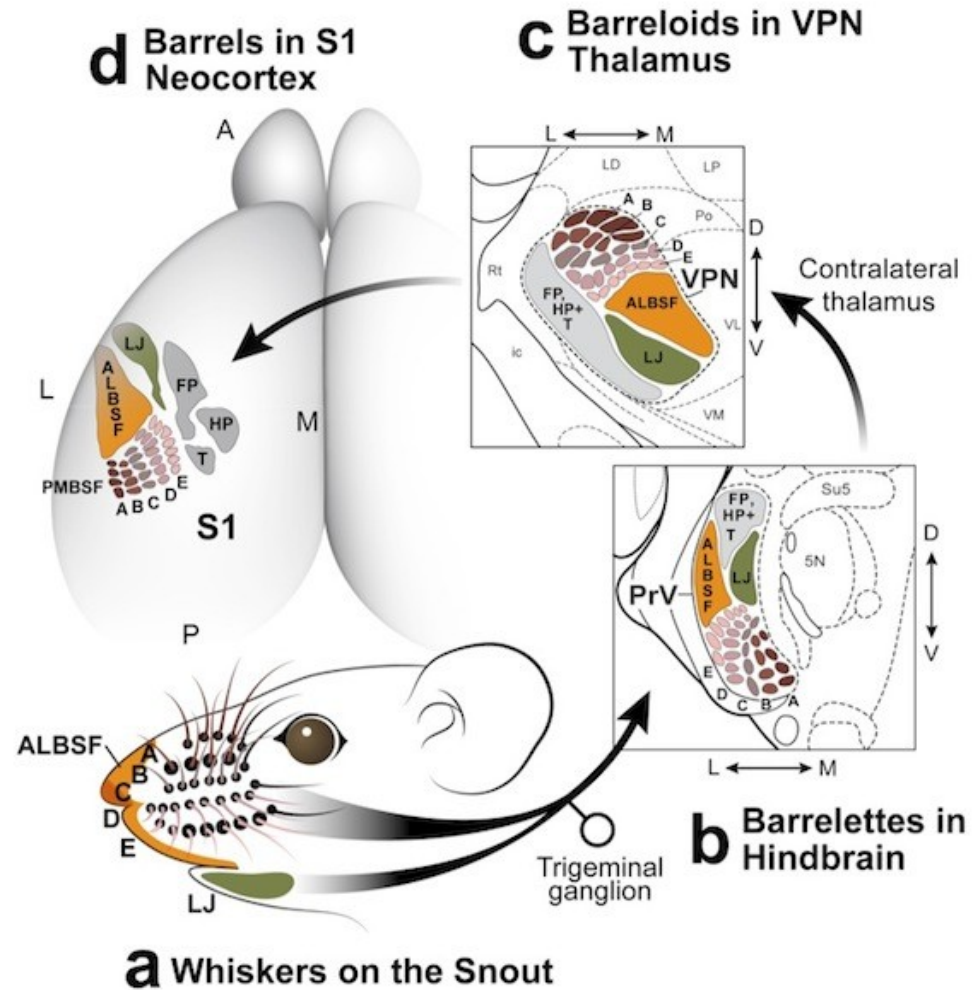
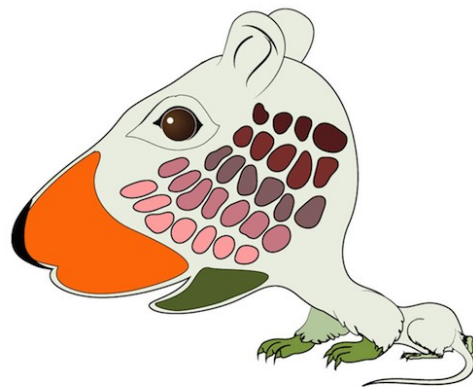
Sci Rep. 2015 Nov 27;5:17325. doi: 10.1038/srep17325.

## **In Vivo Voltage-Sensitive Dye Imaging of Subcortical Brain Function.**

Tang Q<sup>1</sup>, Tsytsarev V<sup>1,2</sup>, Liang CP<sup>1</sup>, Akkentli F<sup>2</sup>, Erzurumlu RS<sup>2</sup>, Chen Y<sup>1</sup>.

# The mouse whisker system

the mouseunculus





# Recording Configuration

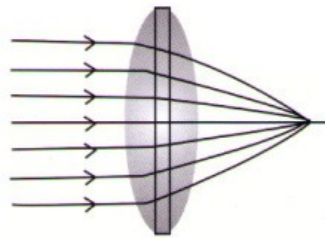
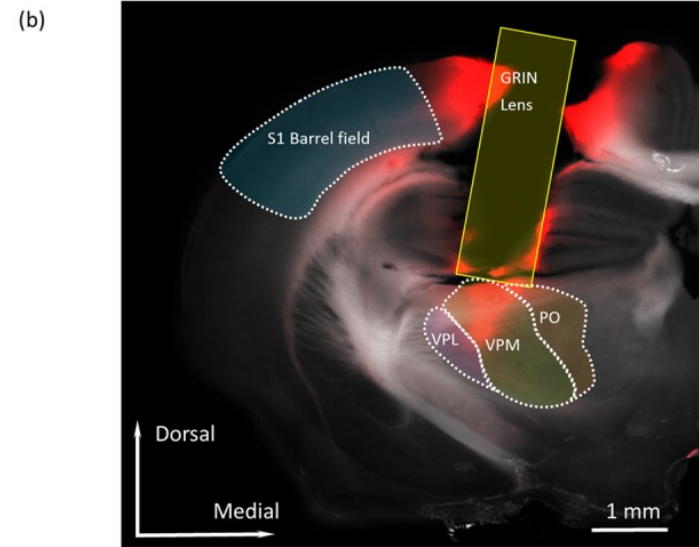
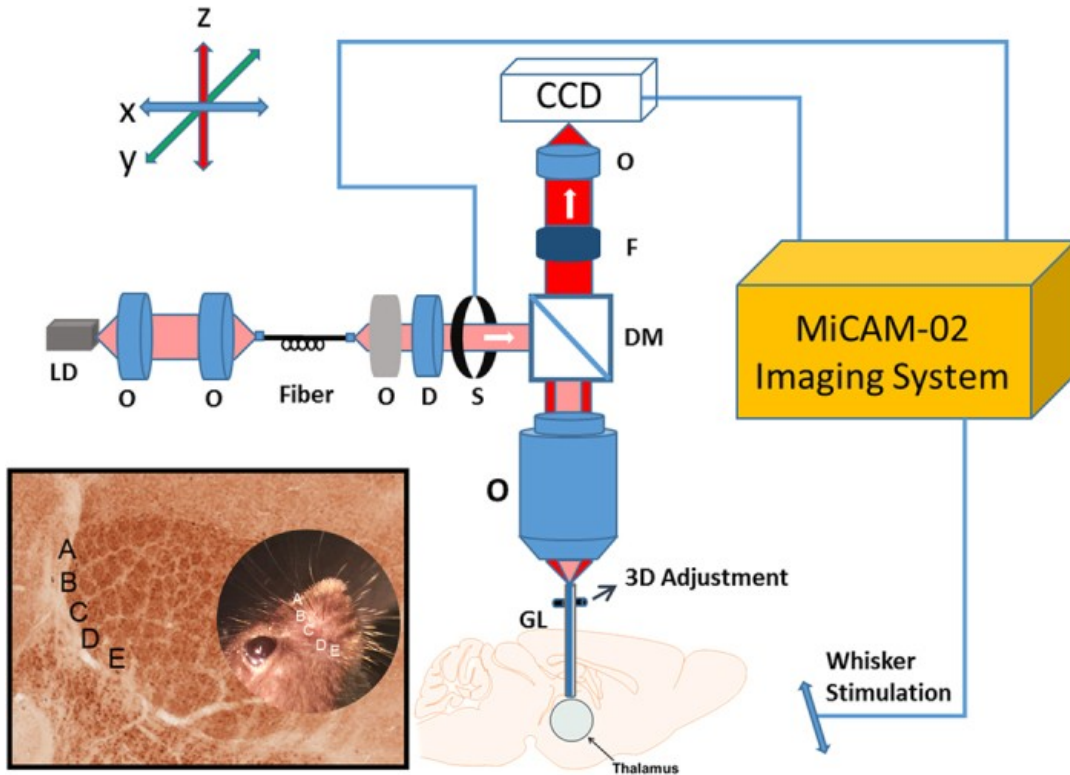


Figure 1: Conventional Lens

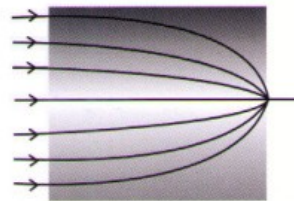
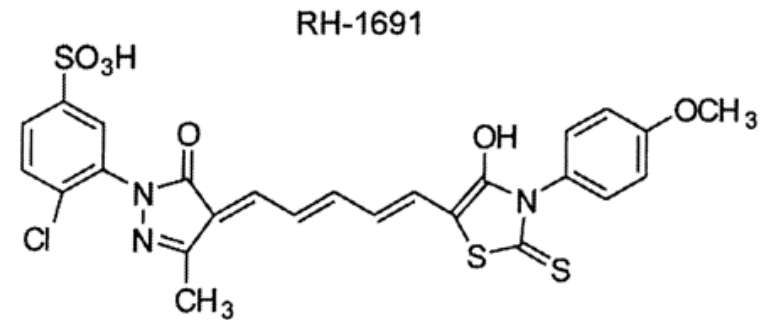
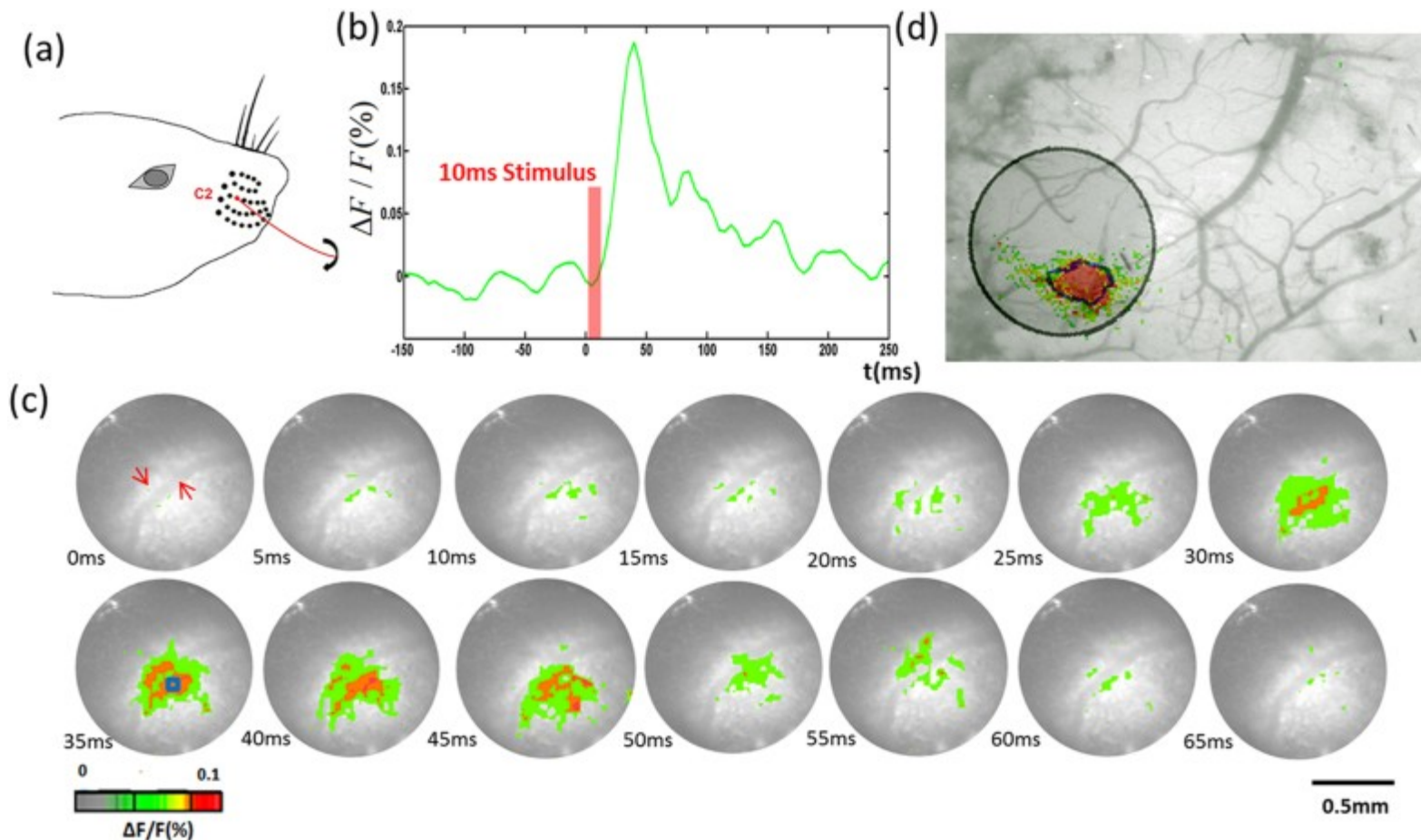


Figure 2: Grin Lens

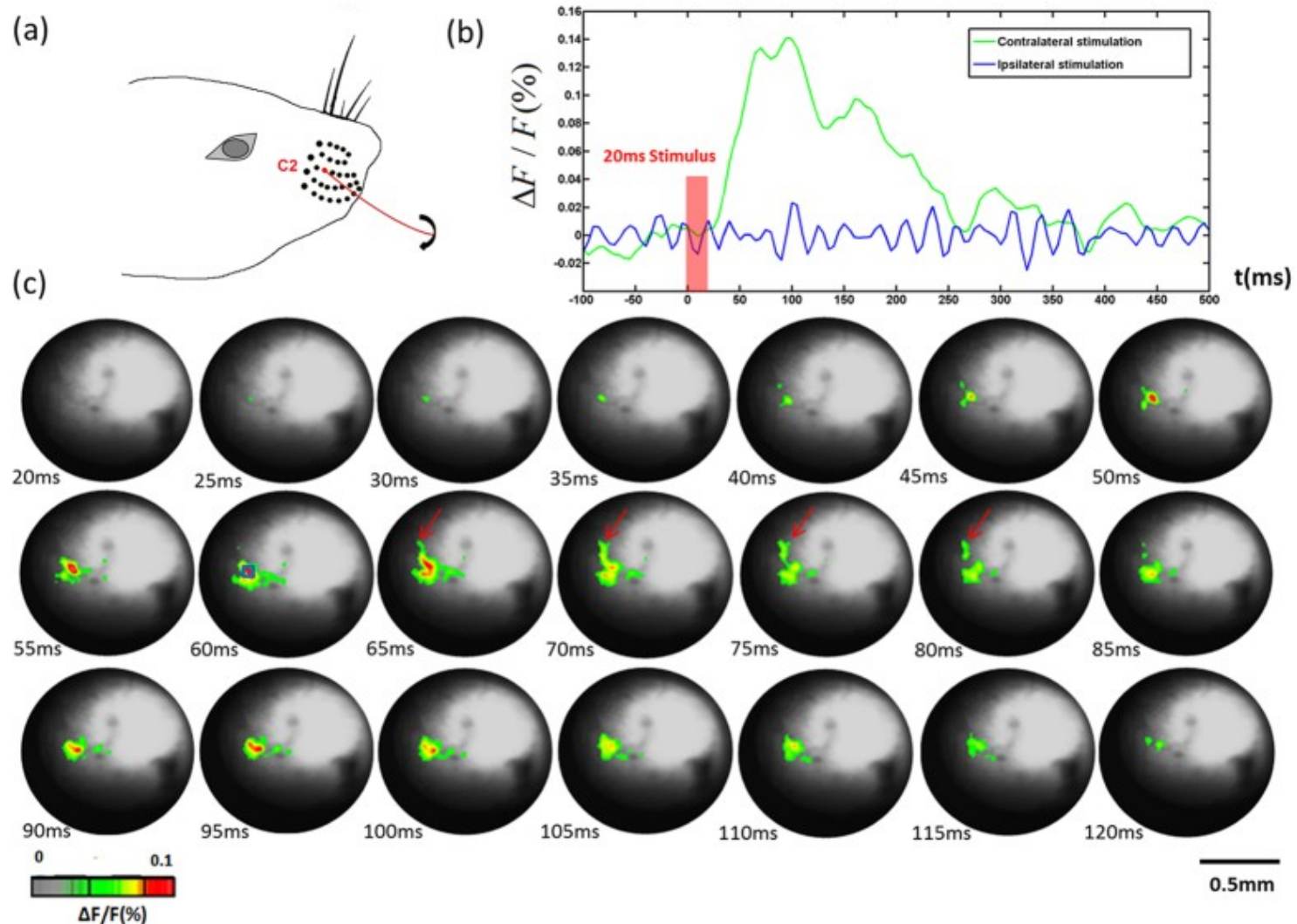


# Barrel Cortex imaging



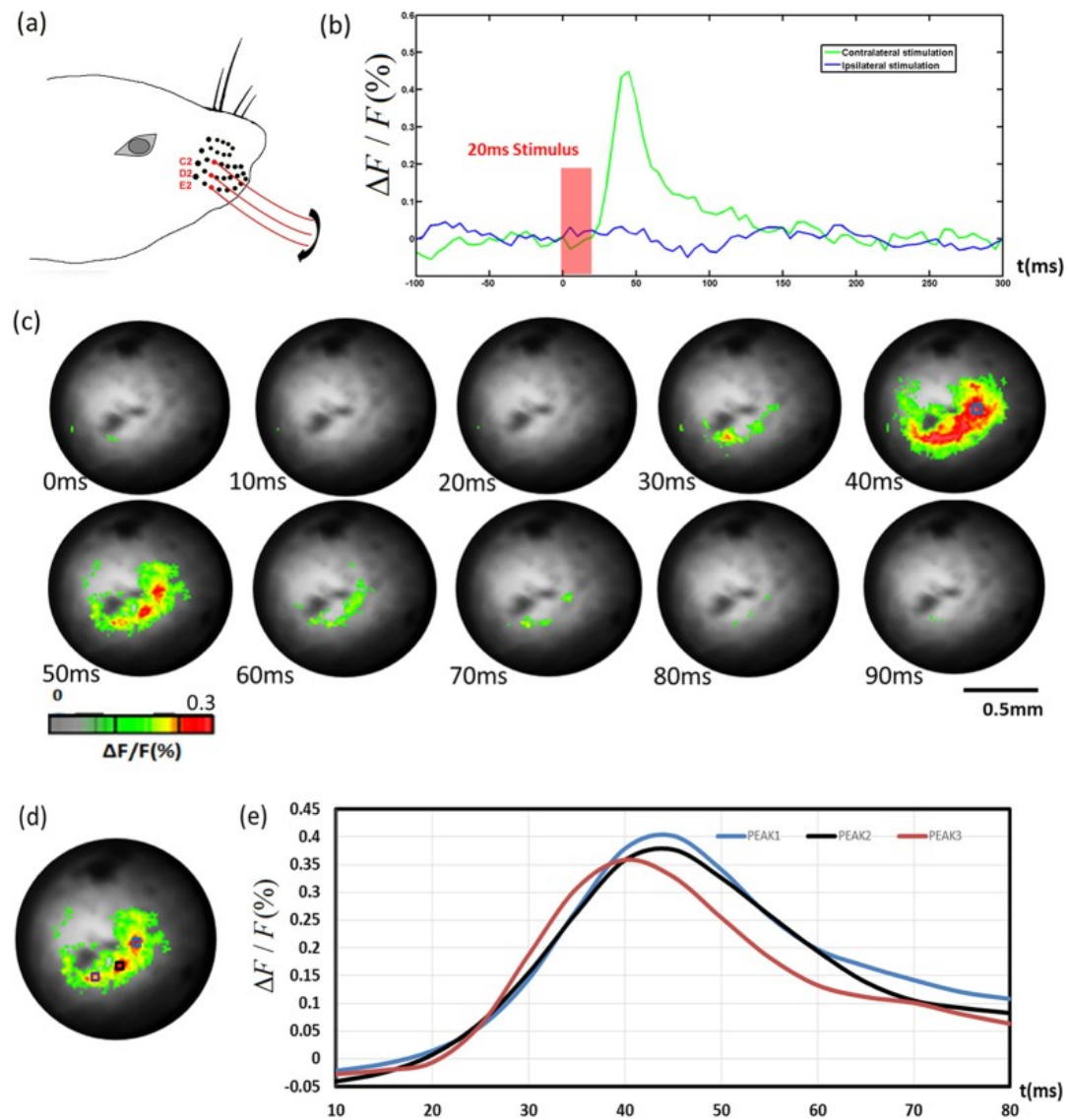
=> GRIN lens gives similar results

# Thalamic imaging: one whisker

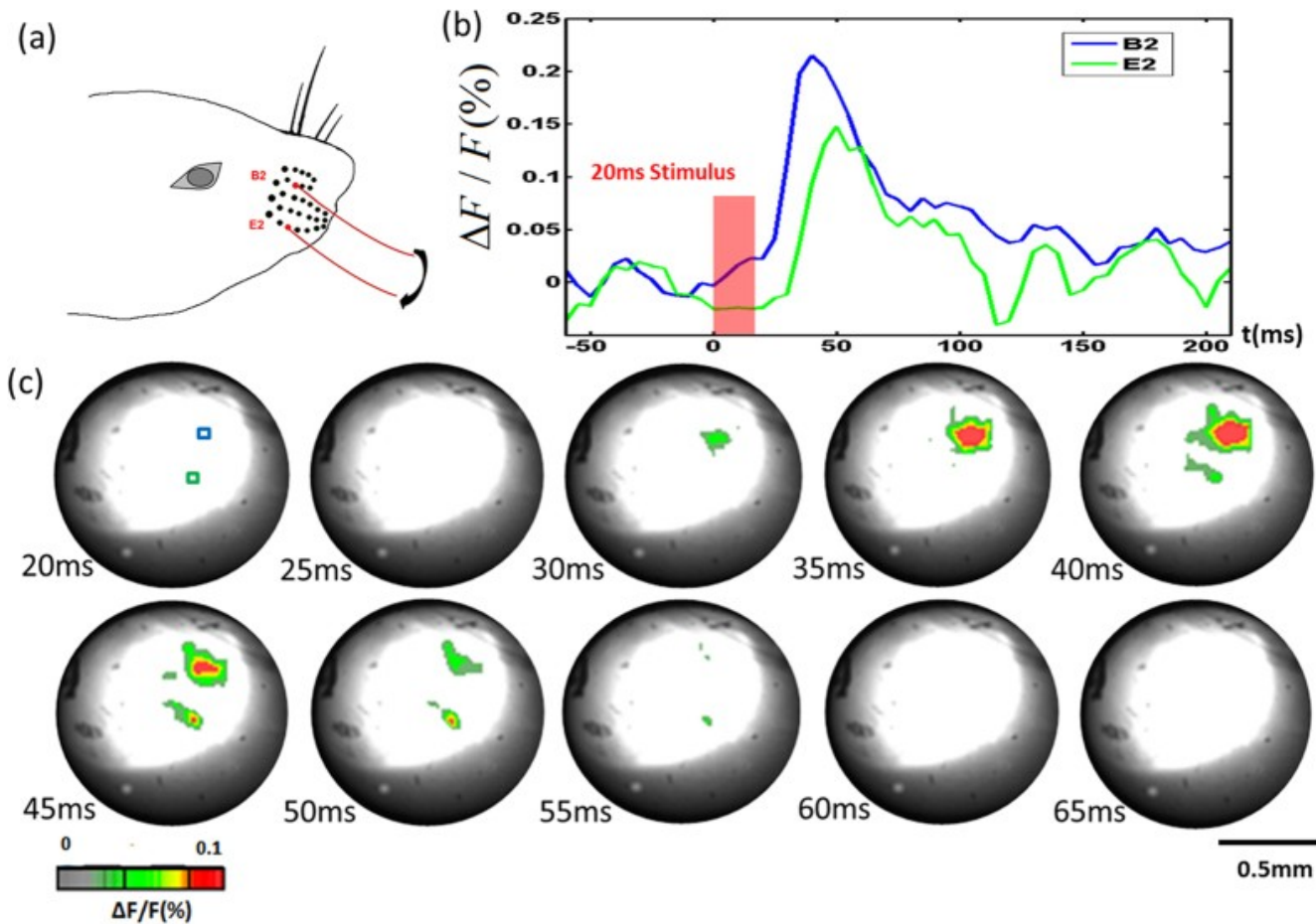


=> barreloid response to stimulation can be detected

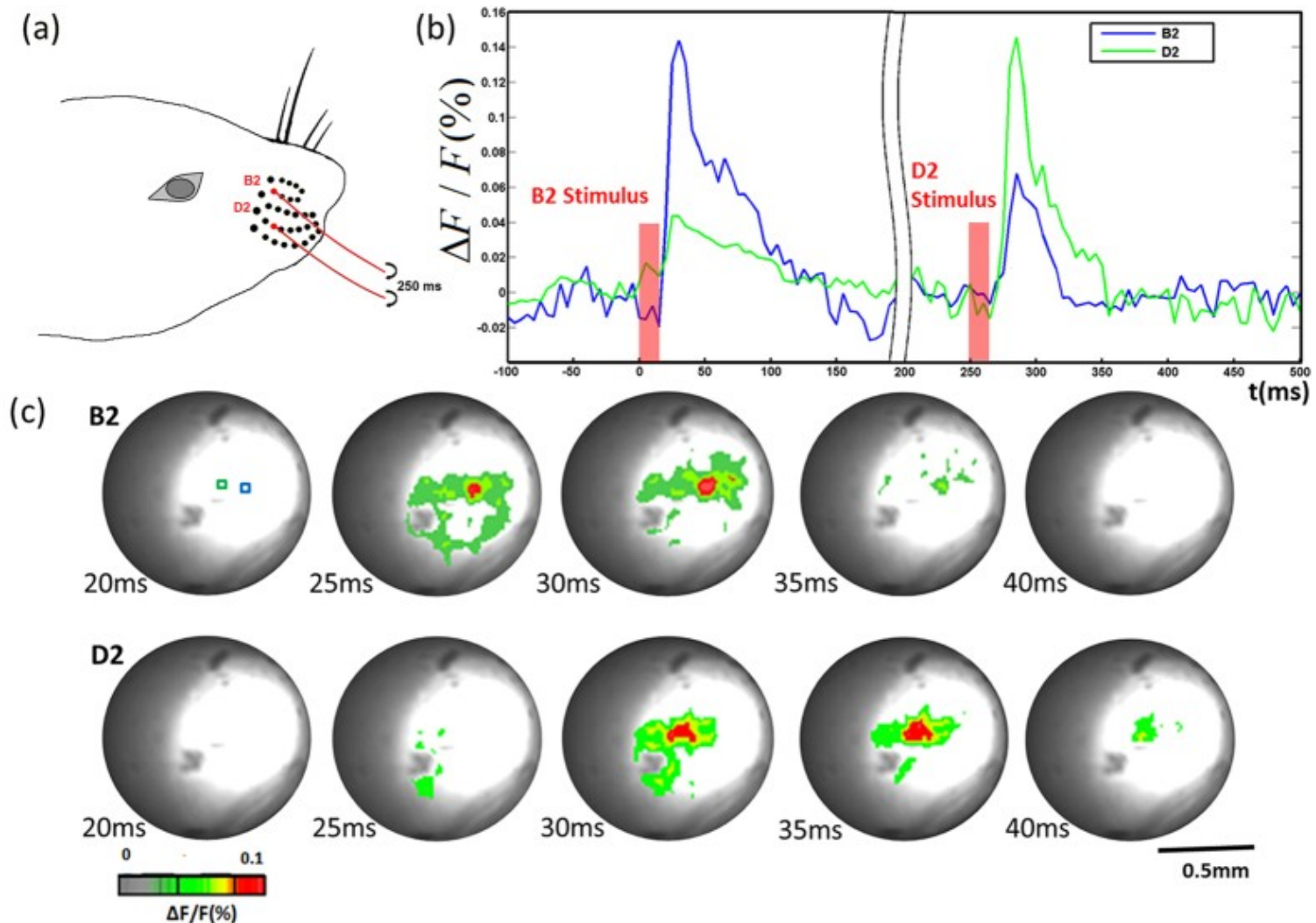
# whisker discrimination I



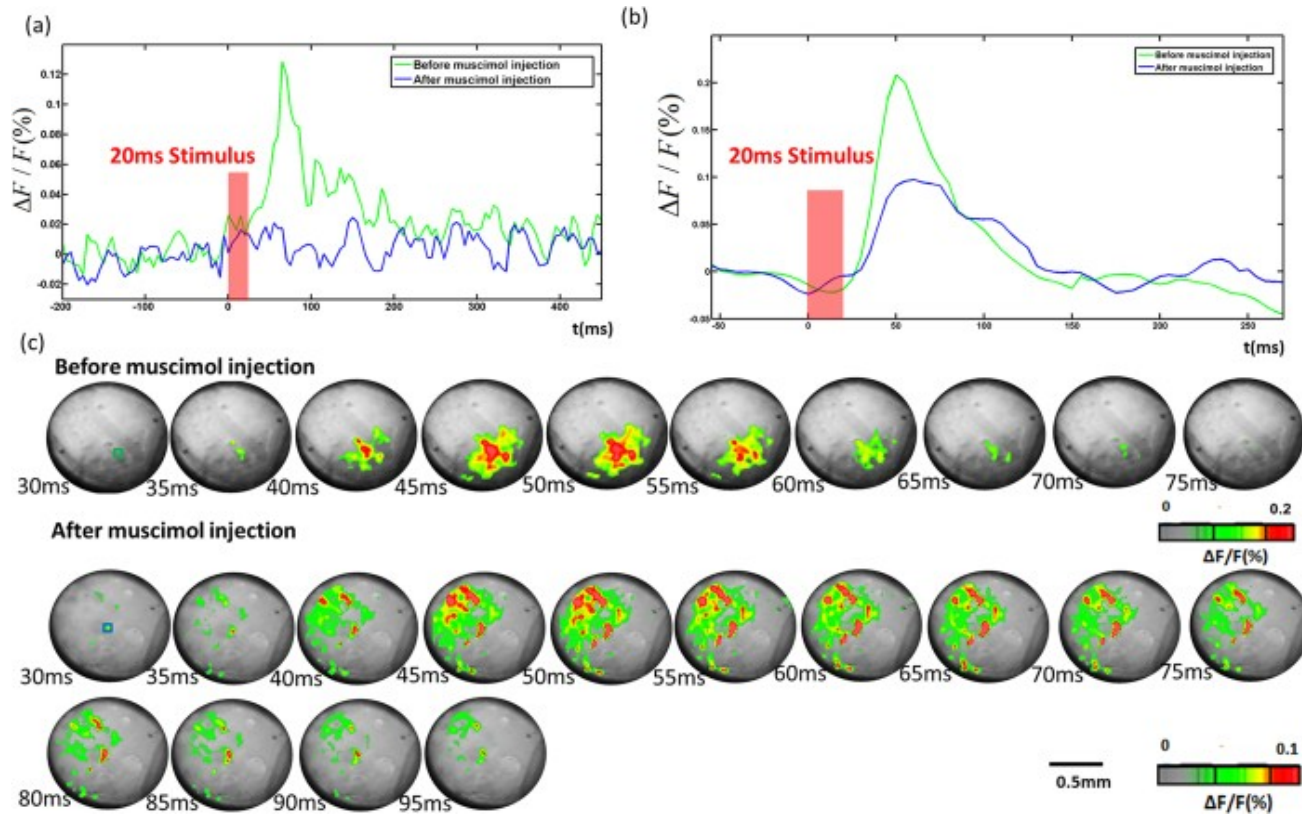
# whisker discrimination II



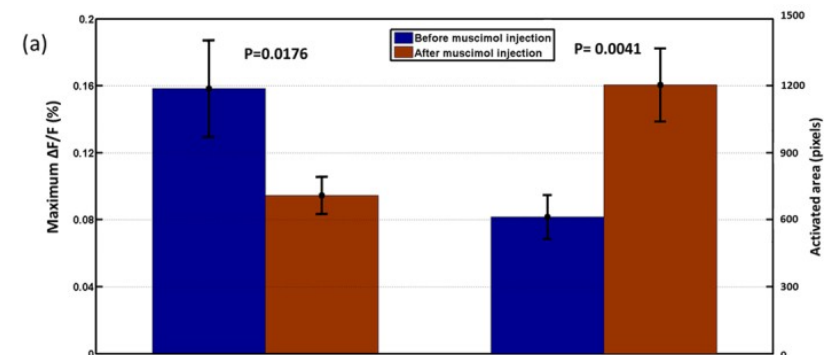
# Temporal discrimination of whisker activity



# Effect of corticothalamic inputs on barreloid activity



GABA<sub>A</sub>-activator



# Conclusion paper 3

- Provides a method to assess the function of subcortical regions while manipulating the cortex
- Good temporal and spatial resolution



# Overall conclusion and outlook

- VSDs are suitable for imaging in dissociated cultures or for „optical field recordings“ *in vivo*  
=> maybe several dyes have to be tested to find an appropriate one
- For *in vivo* applications with single cell resolution, GEVIs should be used
- Further advances in voltage-imaging will rely on
  - Development of faster two-photon microscopes
  - Development of dyes with special transition states

Thank you!