

A photograph showing several white microfluidic chips arranged in a clear plastic tray. Each chip has a grid of small, colored spots (yellow, orange, and red) on a dark background, representing synthetic gene networks. The chips are connected by a network of channels.

# Synthetic gene networks on paper

# Synthetic biology

Definition: "designing and constructing **biological devices** and **biological systems** for useful purposes." *\_Wikipedia*

Interdisciplinary science, combination of biotechnology, evolutionary biology, molecular biology, system biology and biophysics

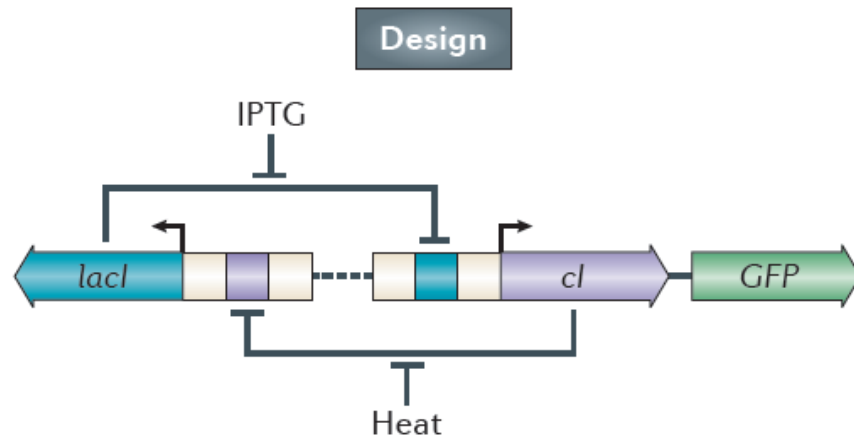
- First used by **Stéphane Leduc's** publication of « Théorie physico-chimique de la vie et générations spontanées » (1910) and « La Biologie Synthétique » (1912)
- Described by **Wacław Szybalski** in 1974: Up to now we are working on the descriptive phase of molecular biology. ... But the real challenge will start when we enter the **synthetic** phase of research in our field.... in the **synthetic biology**, in general.

**Synthetic gene networks** stands in the center of synthetic biology

# Milestones in synthetic gene networks

Toggle switches: Gardner et al, Nature, 2000

**a** Toggle switch



**Behaviour**

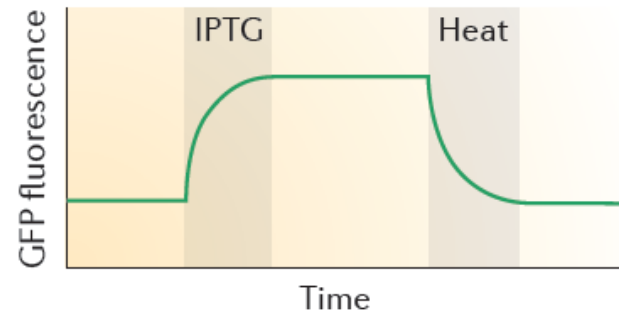


Figure from Cameron, et al, Nat Rev Microbiol, 2014

# Milestones in synthetic gene networks

Repressilator: Elowitz et al, Nature, 2000

## b Repressilator

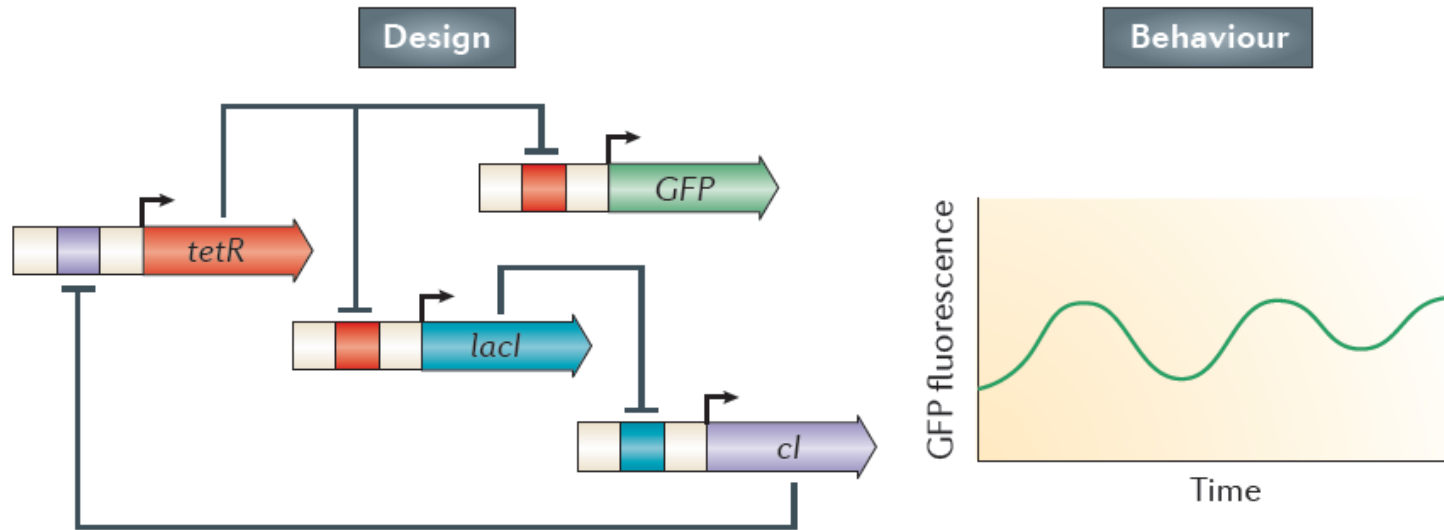
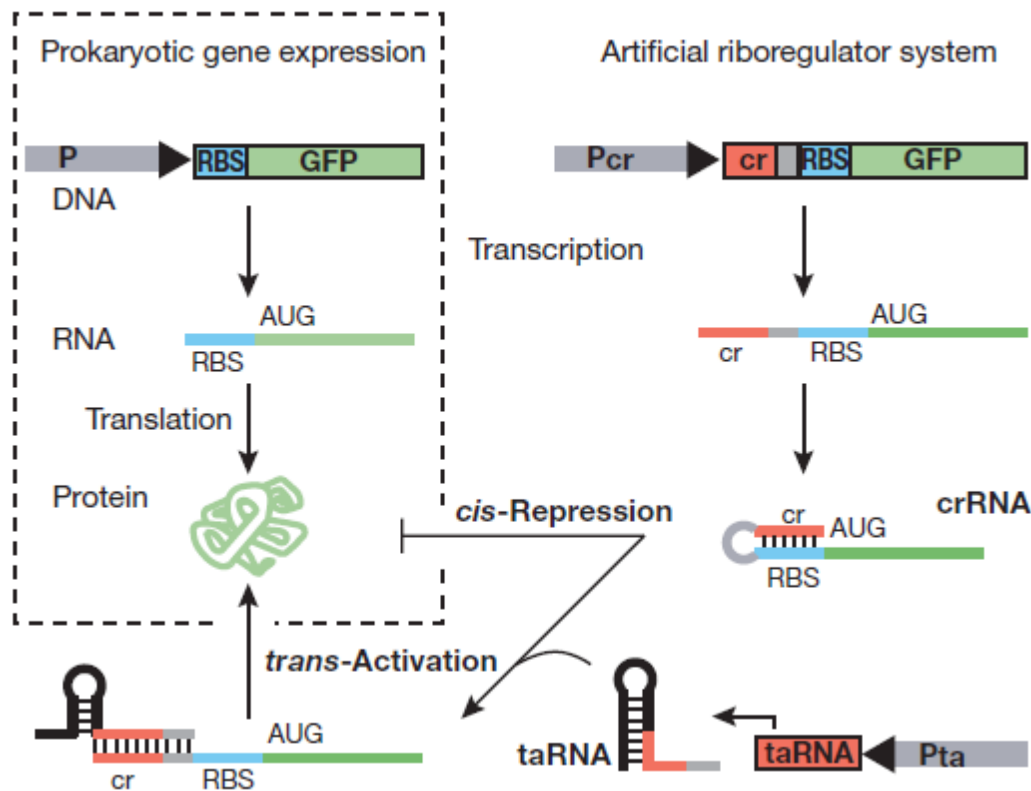


Figure from Cameron, et al, Nat Rev Microbiol, 2014

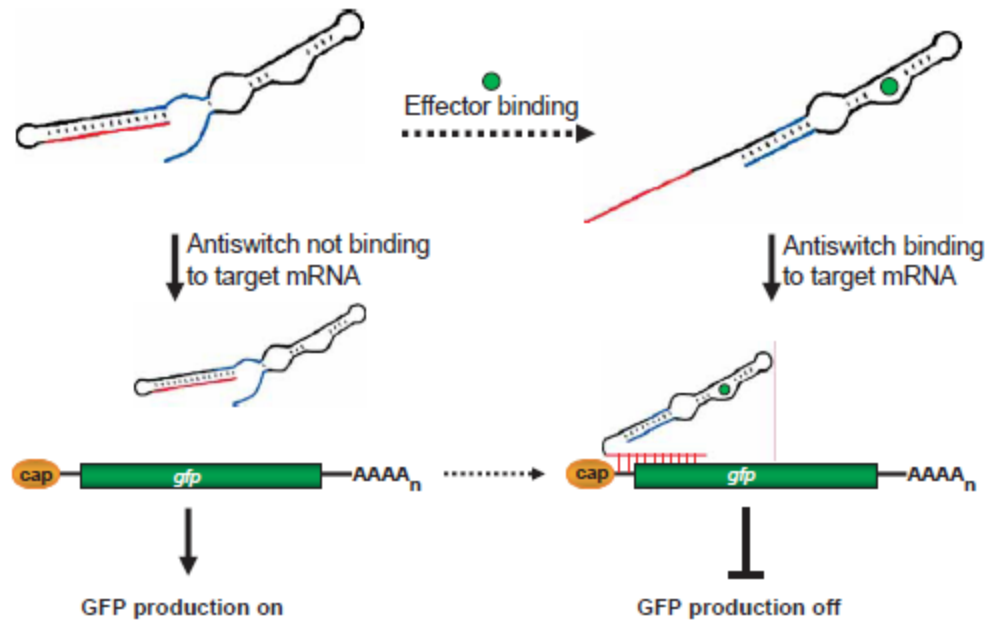
# Milestones in synthetic gene networks

Riboregulator: Isaacs et al, Nat Biotechnol, 2004



# Milestones in synthetic gene networks

Antiswitch riboregulator: Bayer et al, Nat Biotechnol, 2005



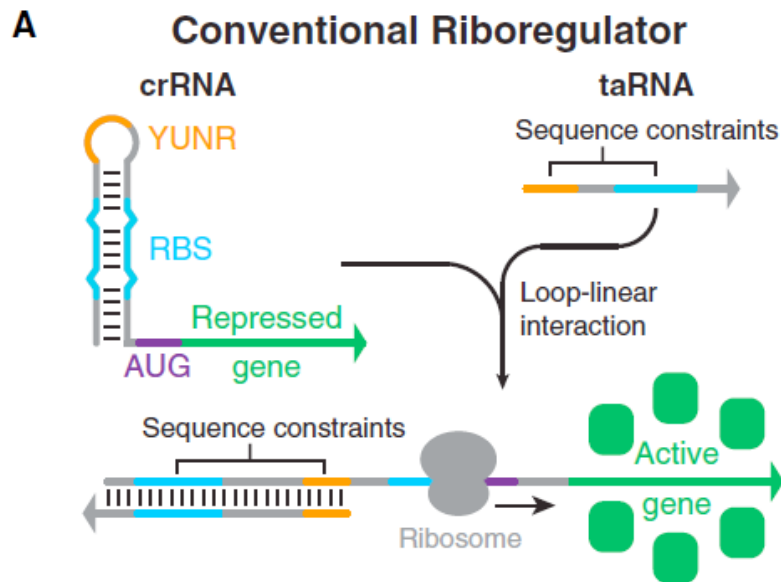
# Conventional riboregulator in Synthetic gene networks

- Programmability, but limited dynamic range: 55-fold activation, 10-fold repressing (v.s protein-based transcriptional regulators, 300~400-fold modulation)
- Due to the sequence constrains, limited number of composable, high-performance parts for constructing genetic circuits
- Difficulties in integrating multiple components into a large, complex synthetic network
- New regulatory components with wide dynamic range, low system crosstalk, high flexibility are required

# Toehold Switches: De-Novo-Designed Regulators of Gene Expression

Alexander A. Green,<sup>1</sup> Pamela A. Silver,<sup>1,2</sup> James J. Collins,<sup>1,3</sup> and Peng Yin<sup>1,2,\*</sup>

Cell 159, 925–939, November 6, 2014 ©2014 Elsevier Inc.

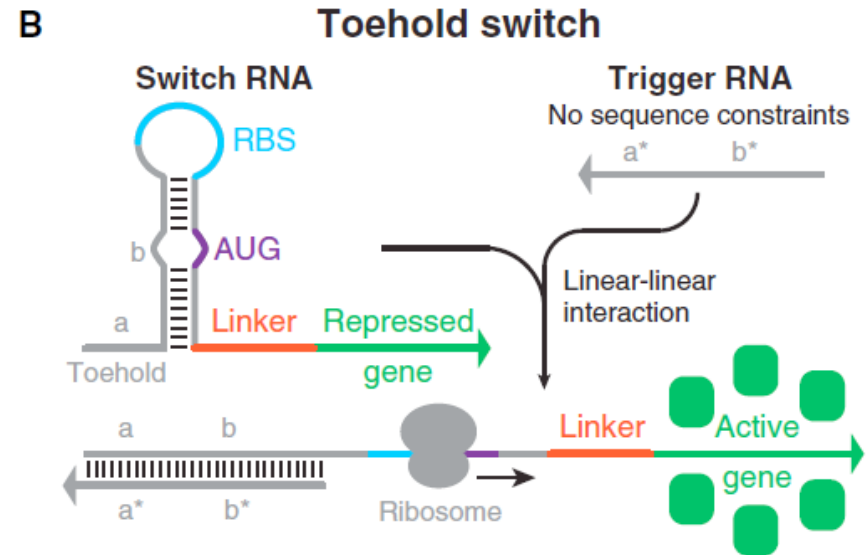


## crRNA

RBS: ribosome binding site, 19 nts

YUNR: pYrimidine-Uracil-Nucleotide-puRine, 4 nts

**taRNA:** 26 nts complementary to YUNR, RBS of crRNA



## Switch RNA

21 nts linker

11 nts loop

6 nts before AUG

9 nts after AUG

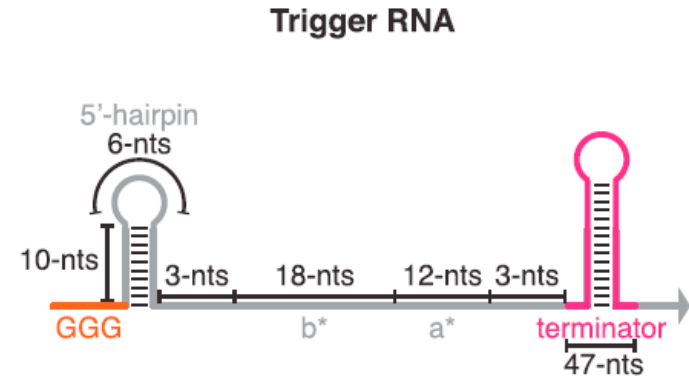
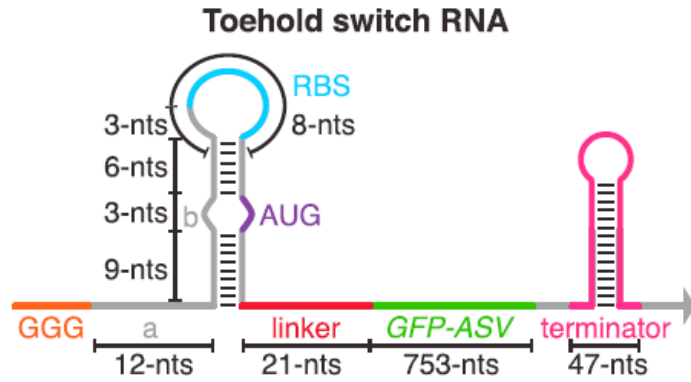
12 nts toehold

**Trigger RNA:** 30 nts complementary to Switch RNA

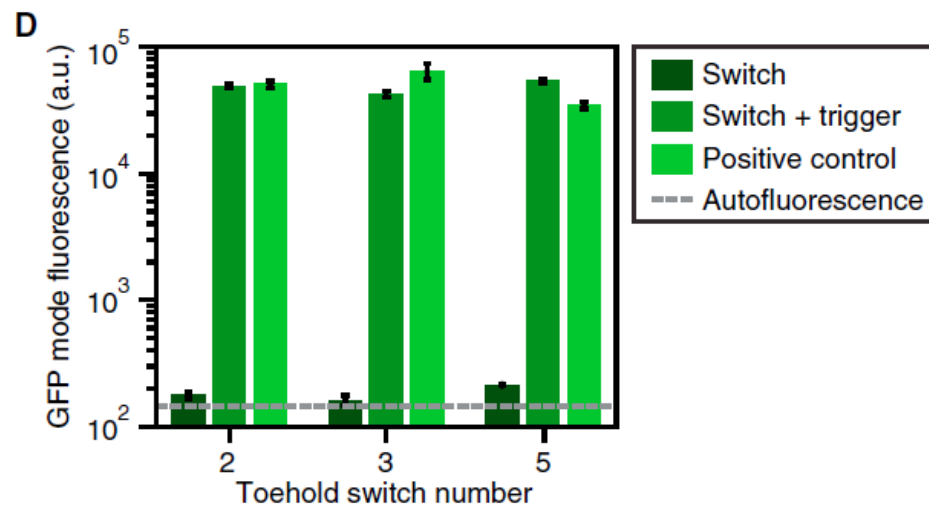
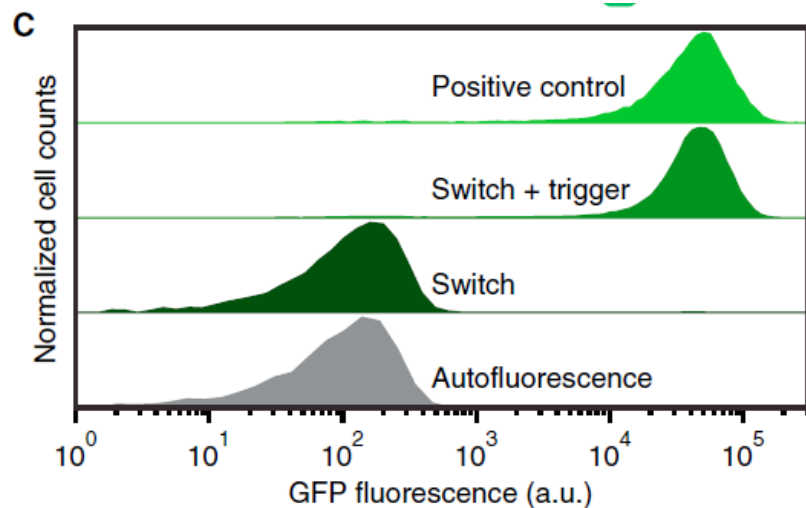


# Scheme of the first generation toehold switches

## i First-generation toehold switches



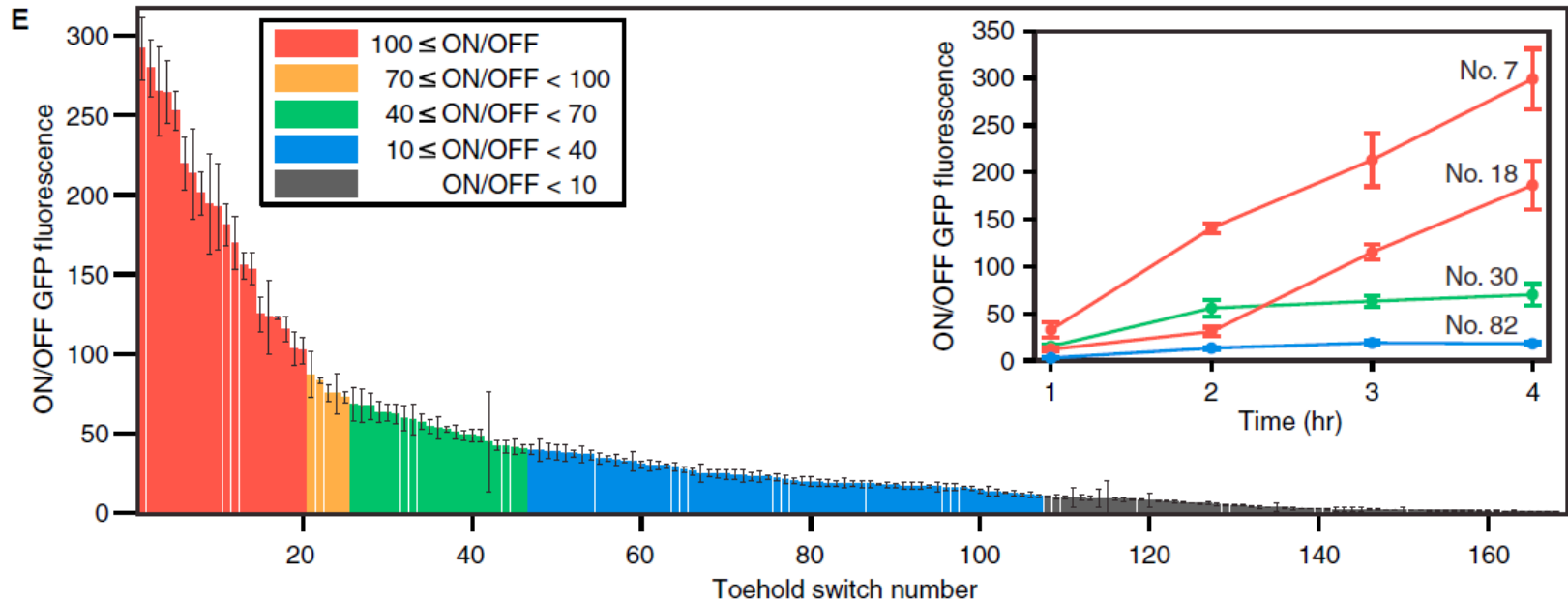
# Toehold switches mediated gene expression >100-fold modulation



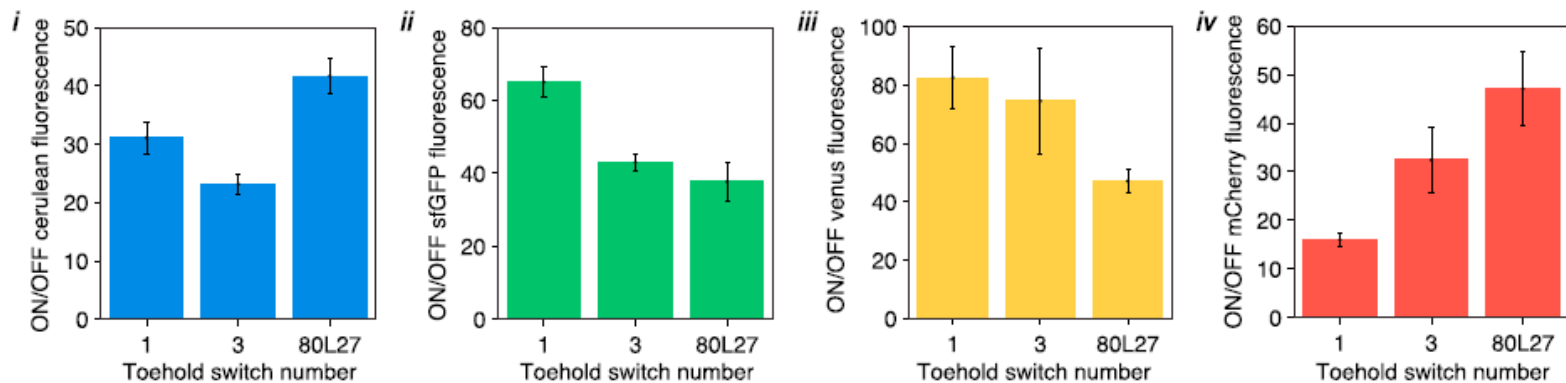
Switch RNA and Trigger RNA, induced by IPTG, T7 RNA polymerase

# ON/OFF ratio of toehold switches mediated gene expression

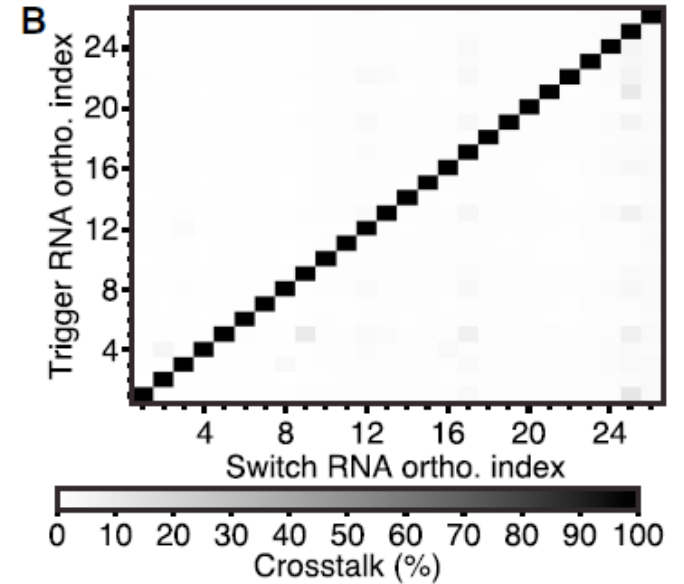
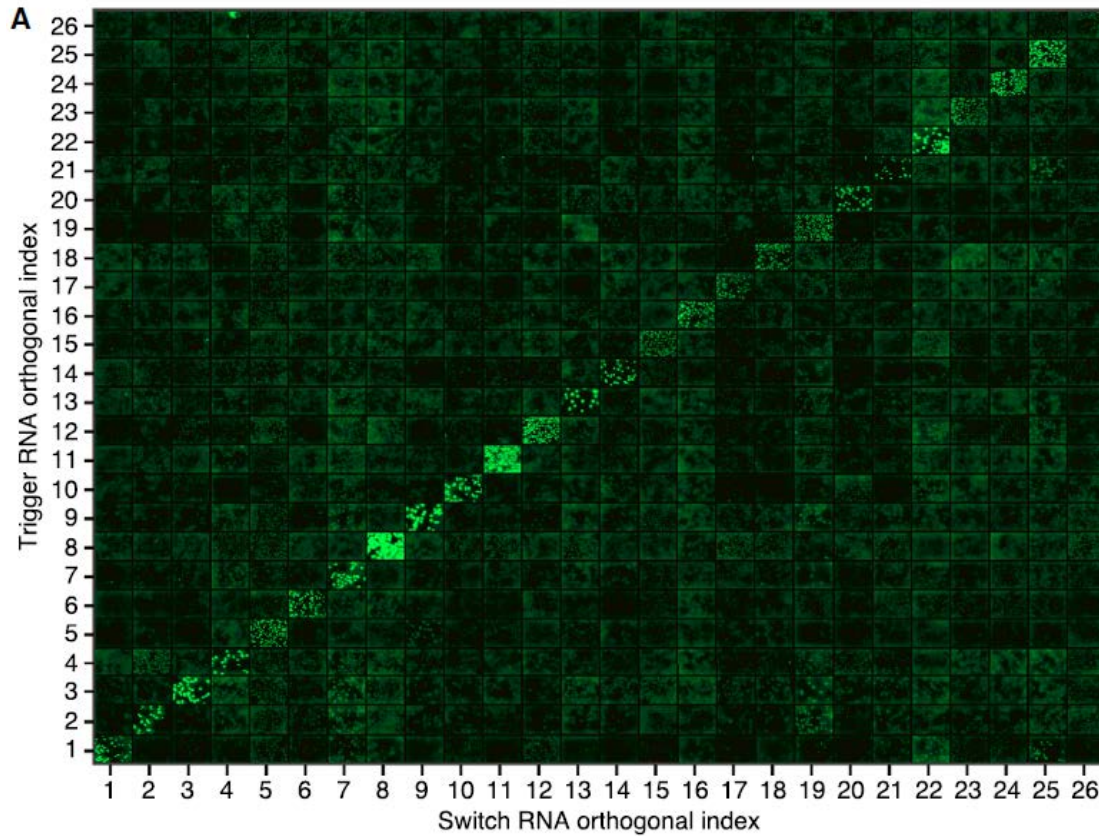
## 168 first generation toehold switches



## H Toehold switch performance with different output proteins



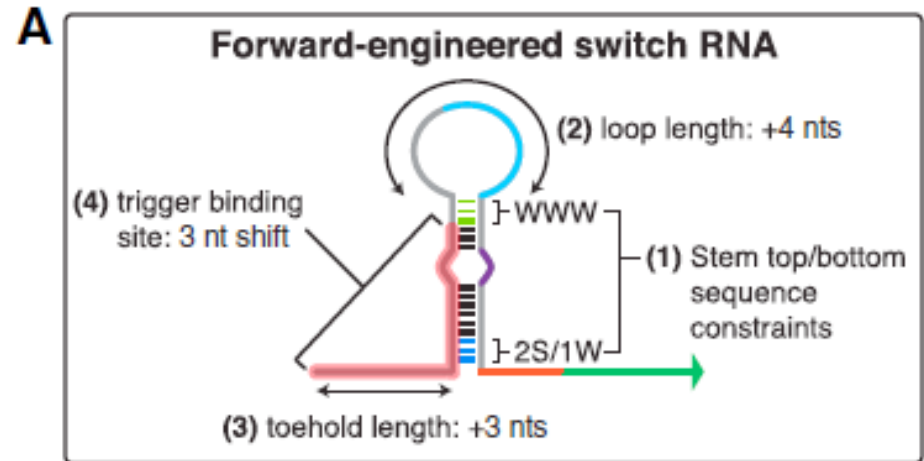
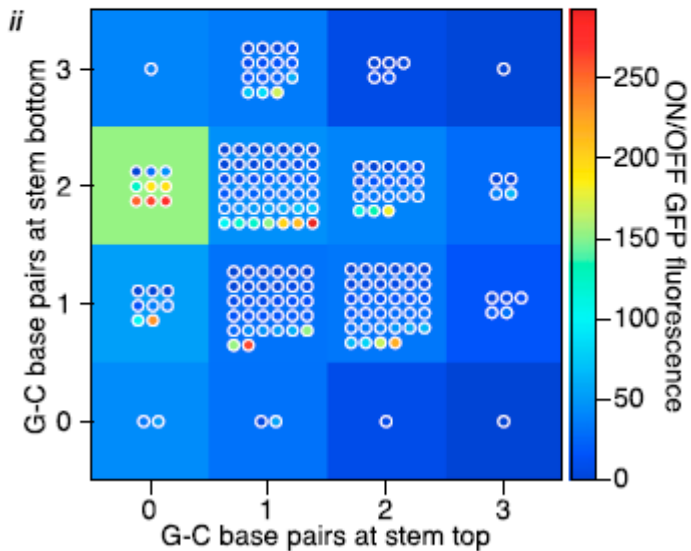
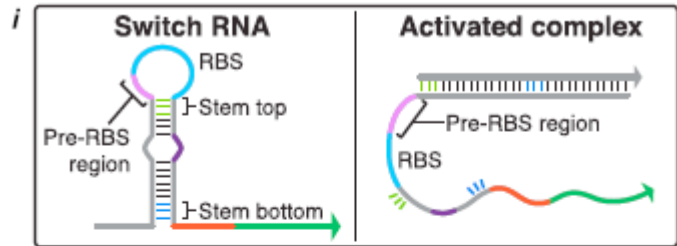
# Orthogonality of toehold switch mediated gene expression



**<12% crosstalk between switch RNA and noncognate triggering RNA**

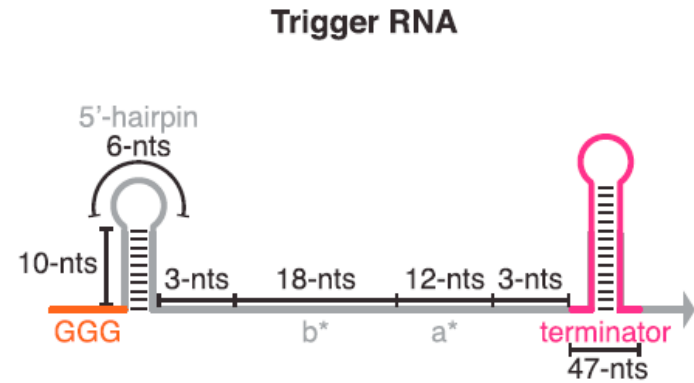
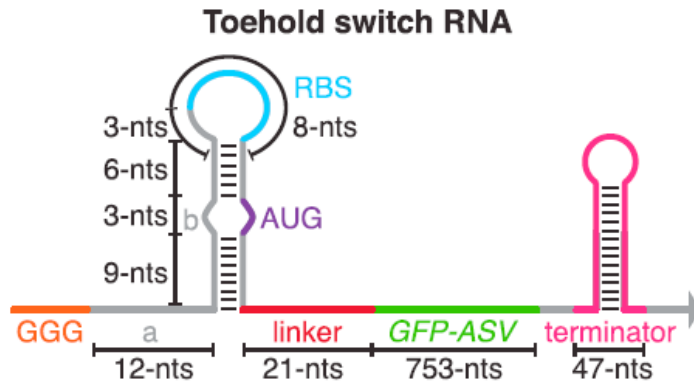
# Design of the forward-engineered toehold switches

## Sequence analysis of first-generation switches

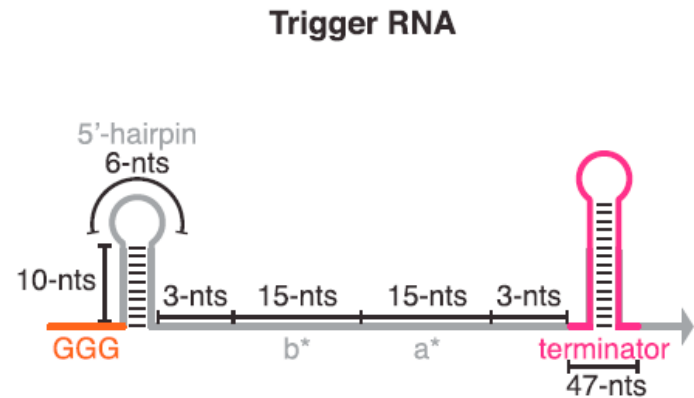
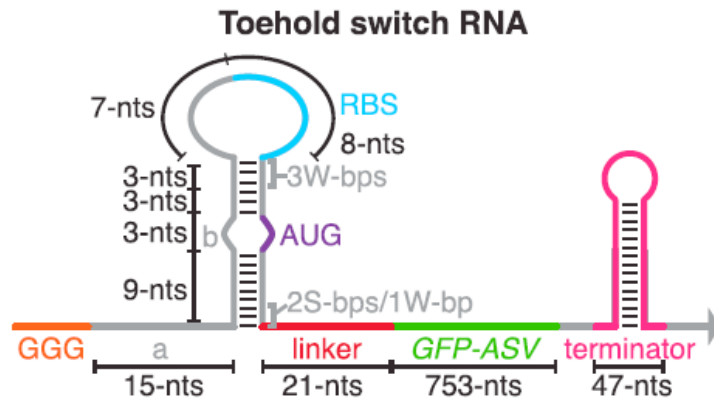


# Scheme of the forward-engineered toehold switches

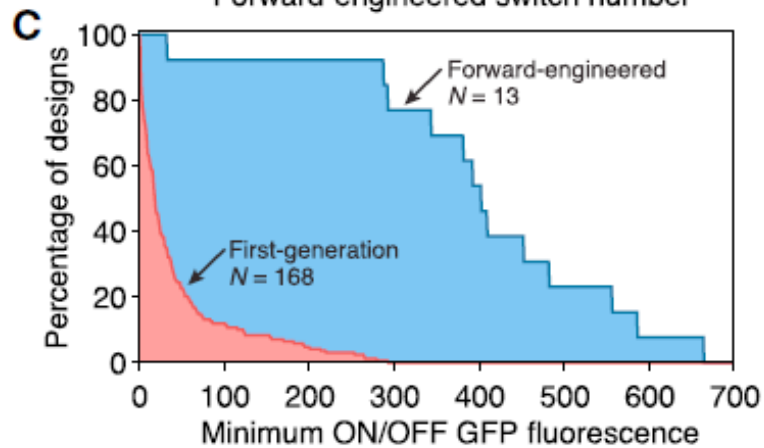
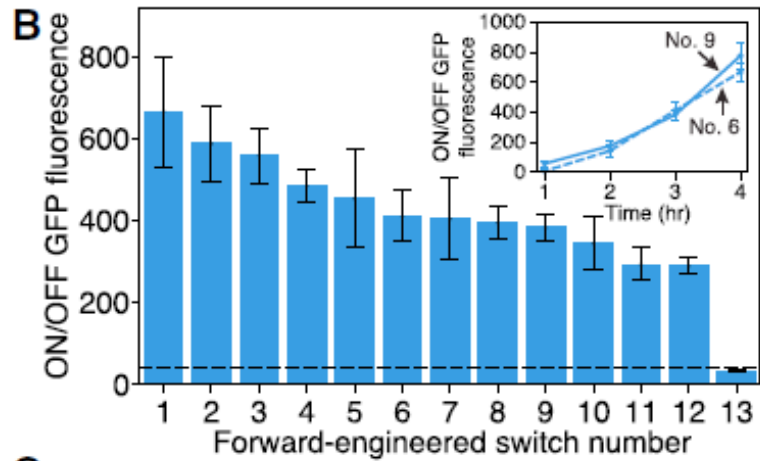
## i First-generation toehold switches



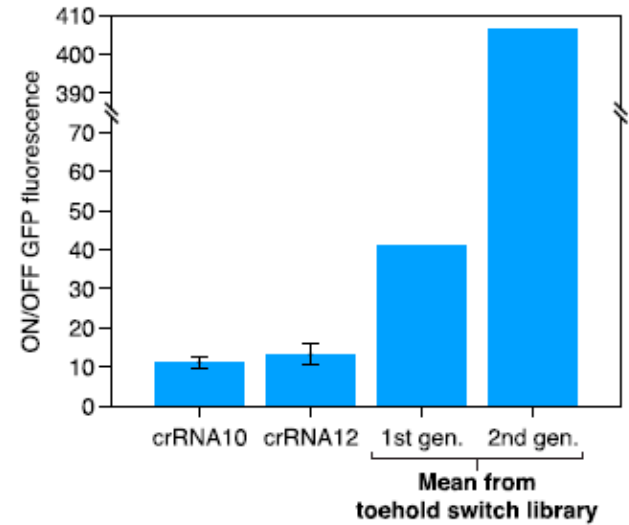
## ii Forward-engineered toehold switches



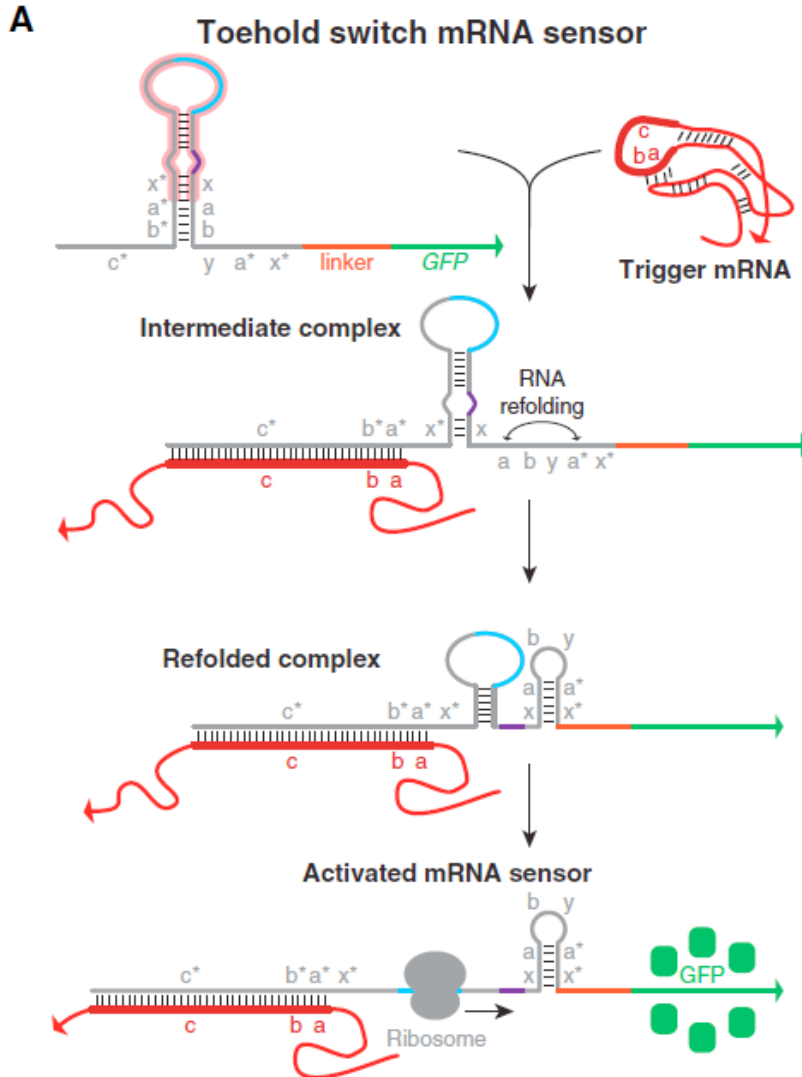
# ON/OFF ratio of toehold switches mediated gene expression



## F Riboregulator performance comparison



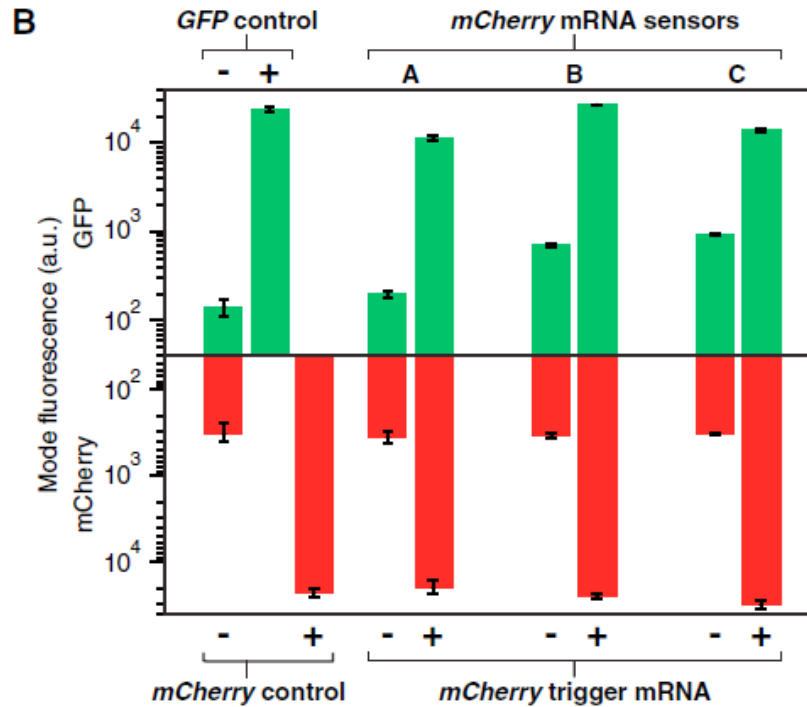
# The arbitrary sequence of trigger RNAs - Toehold switches triggered by mRNA?



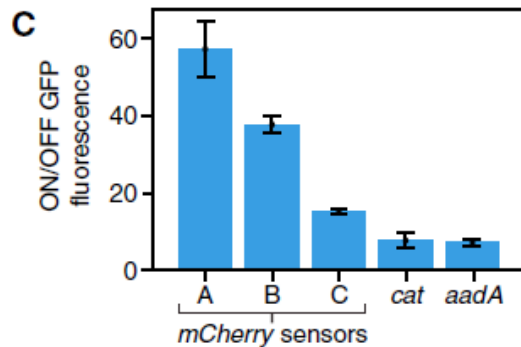
- Increased toehold domain from 12/15 nts to >24 nts
- Design sensor according to the previous toehold switches, common sequences
- RNA refolding mechanism to decrease the energy barrier to switch activation



# Toehold switches triggered by mCherry mRNA



- Sensors only express GFP upon toehold switch by mCherry mRNA
- Simultaneous detection of GFP and mCherry

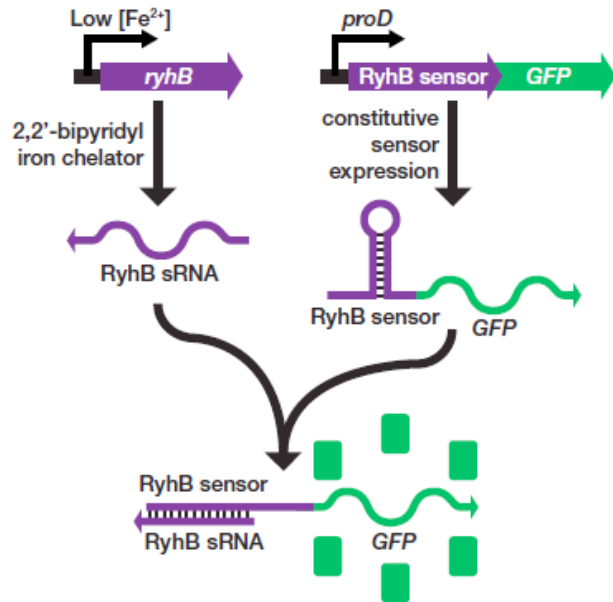


- ON/OFF ratio of sensors triggered by mCherry or Chloramphenicol acetyltransferase (*cat*) or spectinomycin resistance (*aadA*)

# Toehold switches triggered by endogenous RNA

- *ryhB*, 90 nts small RNA, downregulates iron-associated gene when iron level is low
- Increased expression of *ryhB* when iron level is decreased

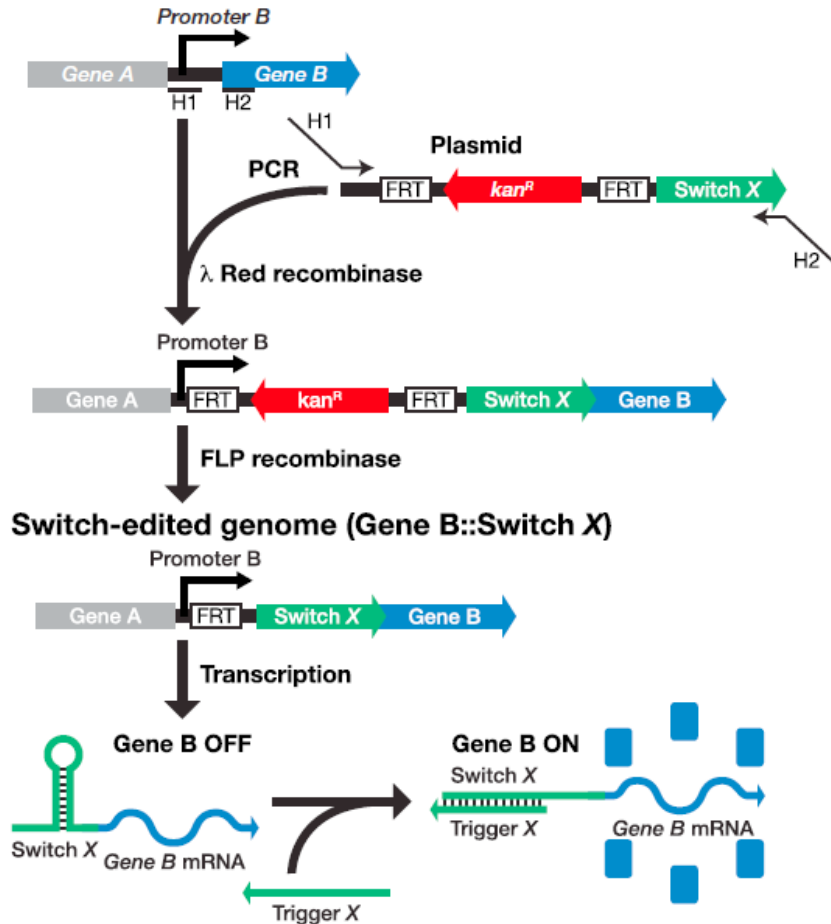
## D Toehold switch endogenous RNA sensor



- *ryhB* sensor increased GFP expression when treated with increasing level of iron-chelating compound
- Control GFP construct showed decreased GFP expression
- **Toehold switch acts as endogenous RNA indicator**

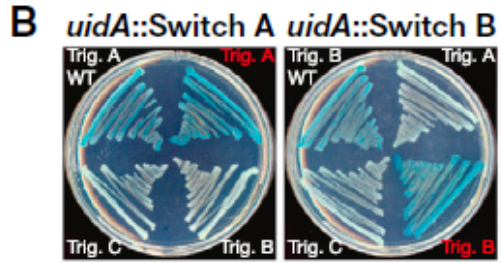
# Toehold switches regulate the translation of endogenous RNA

## A Wild-type genome



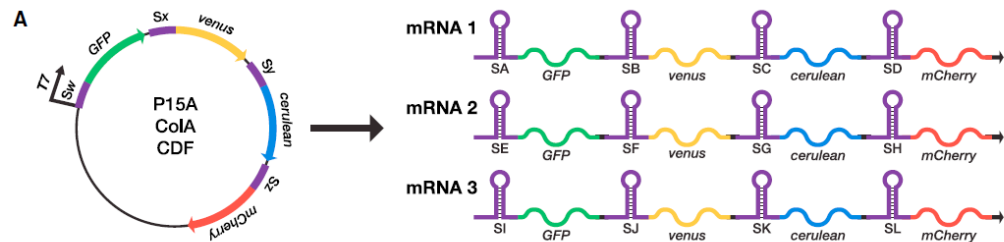
- Knock-in a switch into genome
- Add trigger to stimulate the expression of endogenous genes

# Toehold switches regulate the translation of endogenous RNA

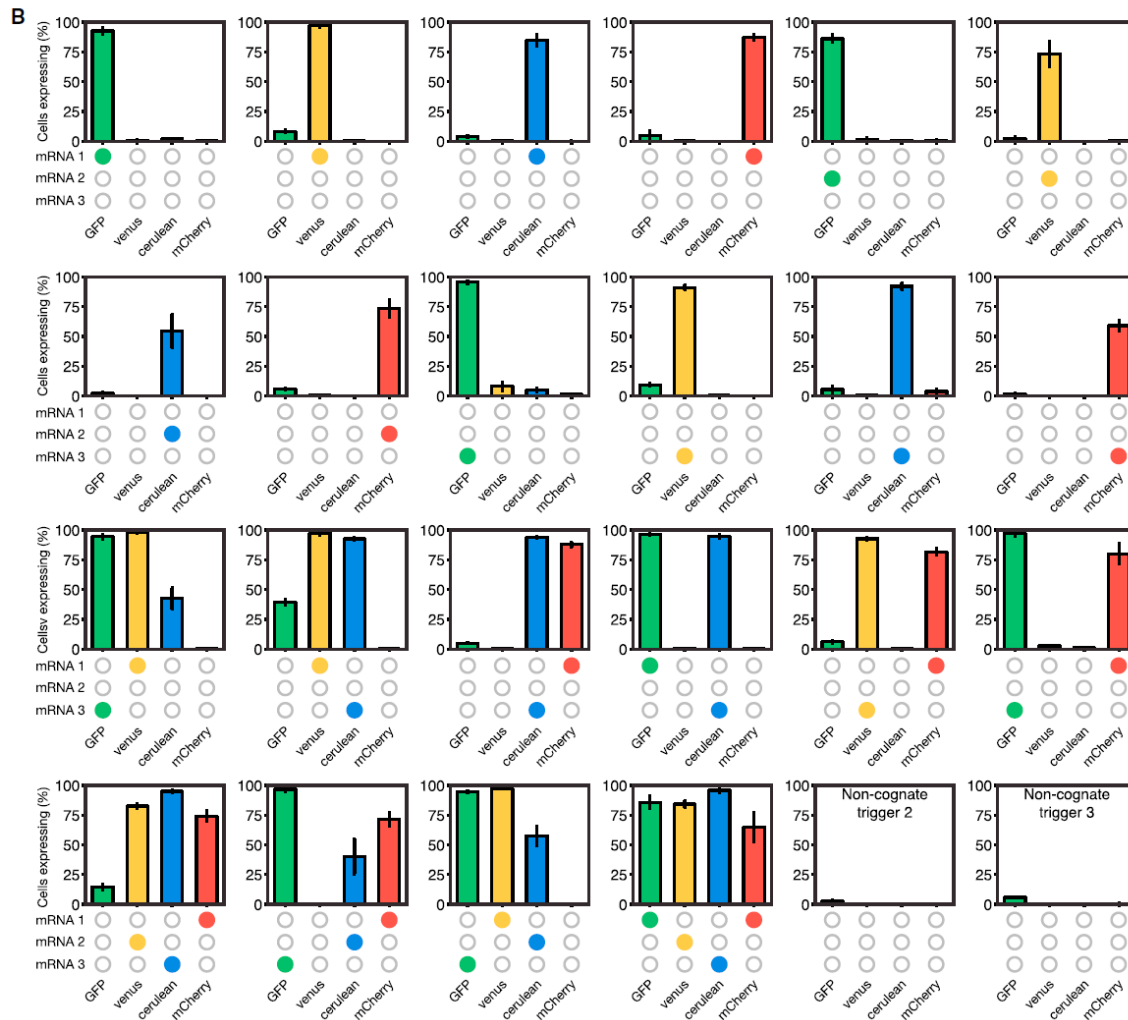


- *uidA*,  $\beta$ -glucuronidase, stained by X-Gluc

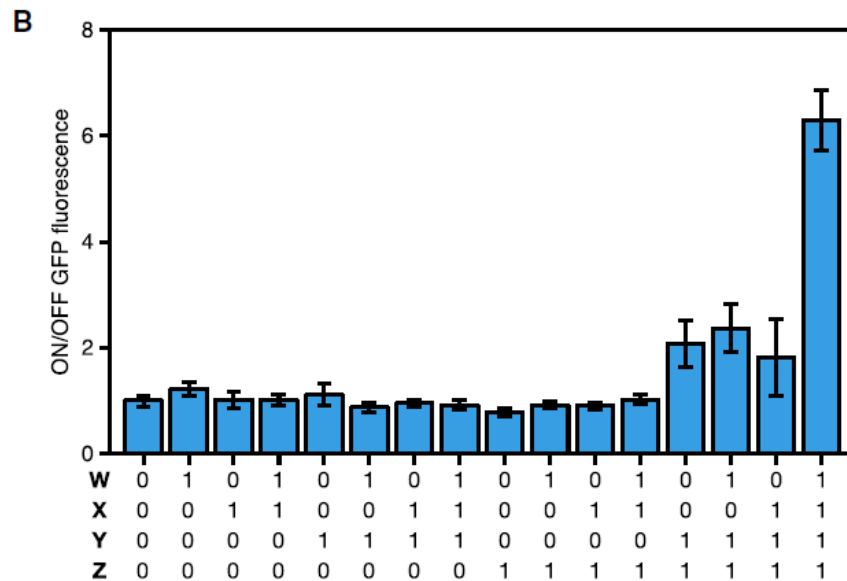
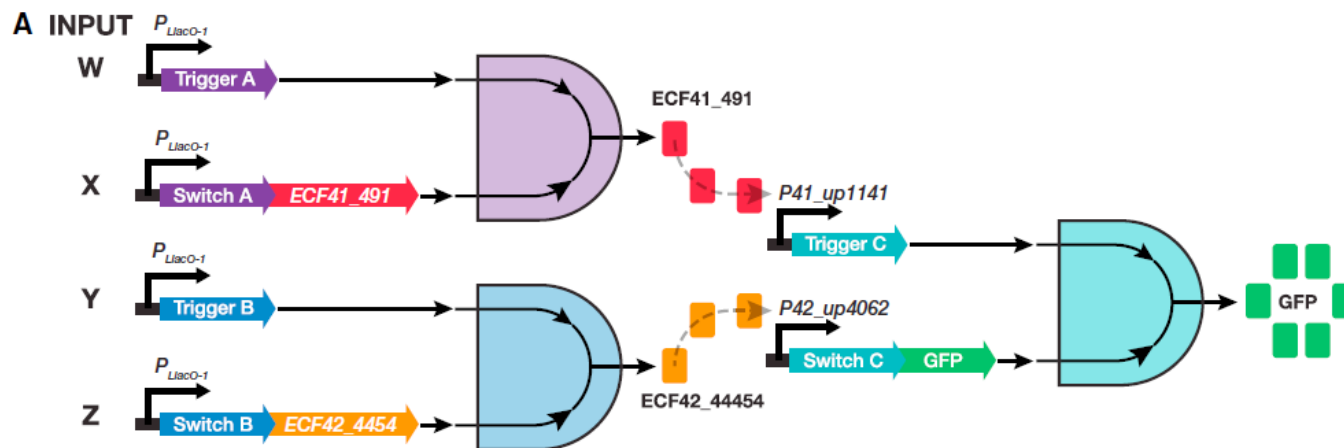
# Multiplexing regulation by toehold switches



- 12 toehold switches
- 4 fluorescent proteins



# Implementation of a 4-Input AND Circuit



ECF: transcription factors

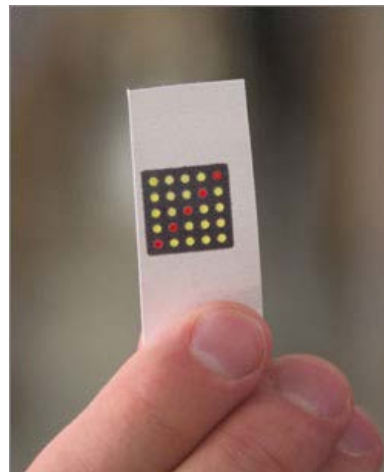
# Toehold switches in synthetic gene networks

- Versatile and powerful platform for posttranscriptional regulation, high performance, high dynamic range. High orthogonality
- Can be used to monitor the endogenous gene expression and to regulate endogenous gene expression
- Multiplexing
- All were done in cultured E.Coli

# Paper-Based Synthetic Gene Networks

Keith Pardee,<sup>1,2</sup> Alexander A. Green,<sup>1,2</sup> Tom Ferrante,<sup>1</sup> D. Ewen Cameron,<sup>2,3</sup> Ajay DaleyKeyser,<sup>1</sup> Peng Yin,<sup>1</sup> and James J. Collins<sup>1,2,3,\*</sup>

Cell 159, 940–954, November 6, 2014 ©2014 Elsevier Inc.

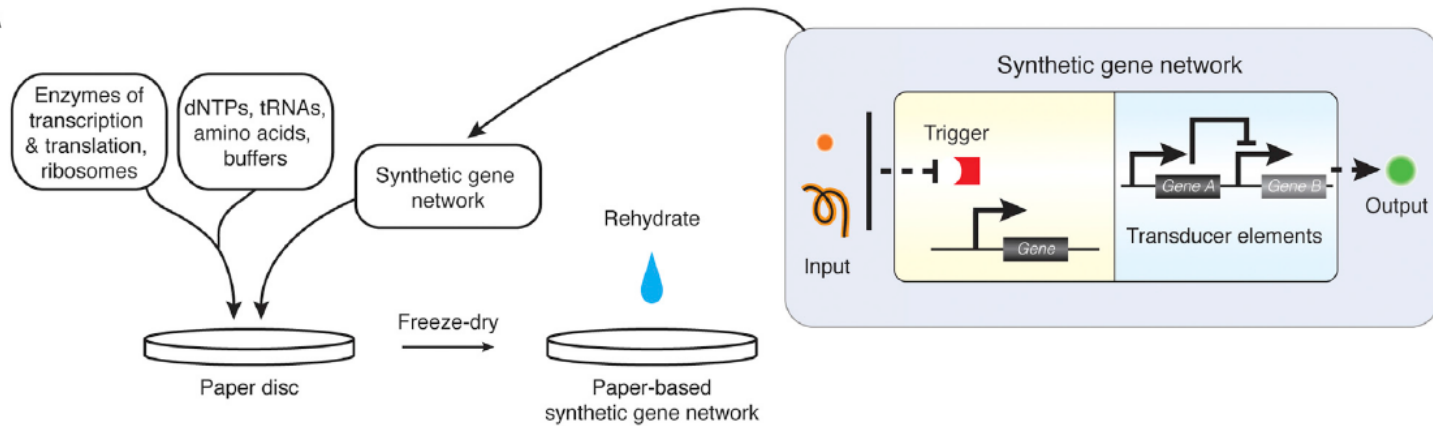




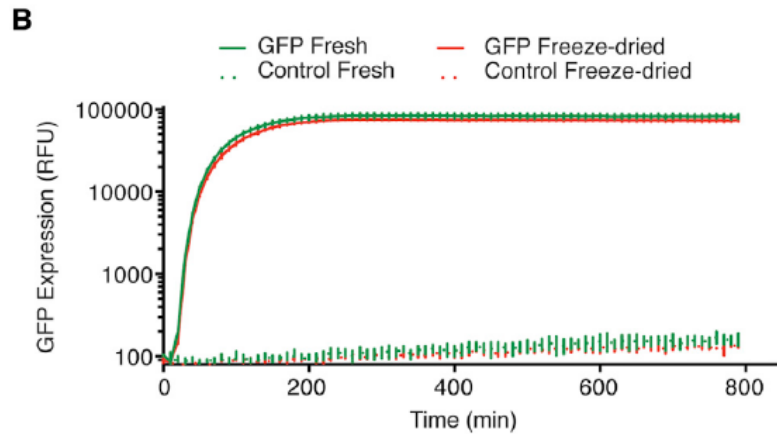
- Cell-based synthetic gene networks outside of laboratory are restricted due to the biosafety and practicality of the cellular host
- Synthetic gene networks have been studied in solution-phase reactions using freshly frozen cell-free systems, often liposomes. But solution-phase reactions are still not suitable and practical for handling outside of the lab
- Paper-based measurement or diagnostics mostly rely on chemical reaction

# Overview of synthetic gene networks on paper

A

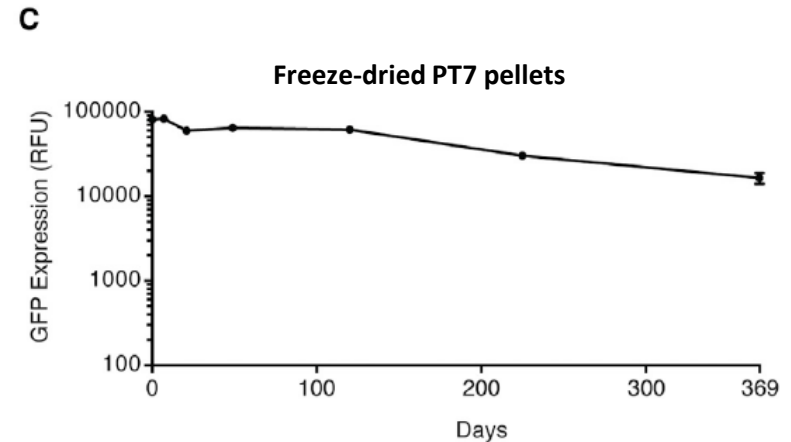


# GFP expression in solution phase from fresh and freeze-dried PT7 cell-free reactions



**PT7 system:** Ribosome + 35 purified bacterial proteins

Freeze-dry – rehydrate with H<sub>2</sub>O – 37°C incubation

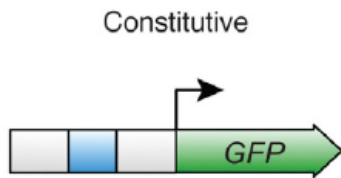


Freeze-dry – room temperature – rehydrate with H<sub>2</sub>O – 37°C incubation

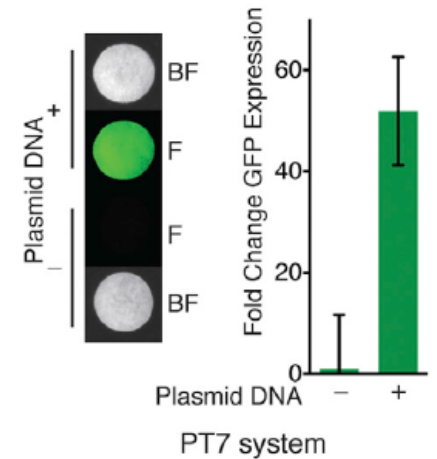
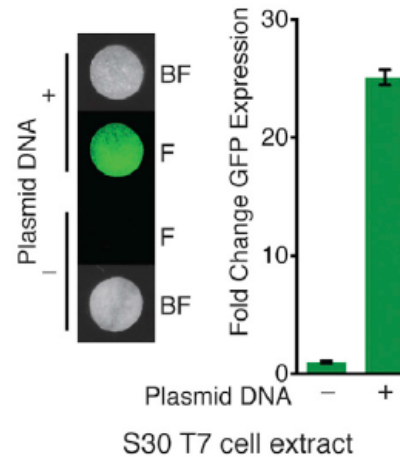
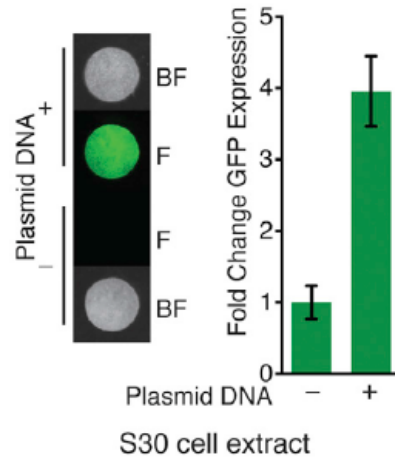
# GFP expression on cellulose matrix paper

2mm paper discs, 1.8ul reaction  
Freeze-dry  
Rehydrate with 1.8ul H<sub>2</sub>O  
Incubate at 37 °C

**D**



**E**



# Synthetic gene networks on cellulose matrix paper

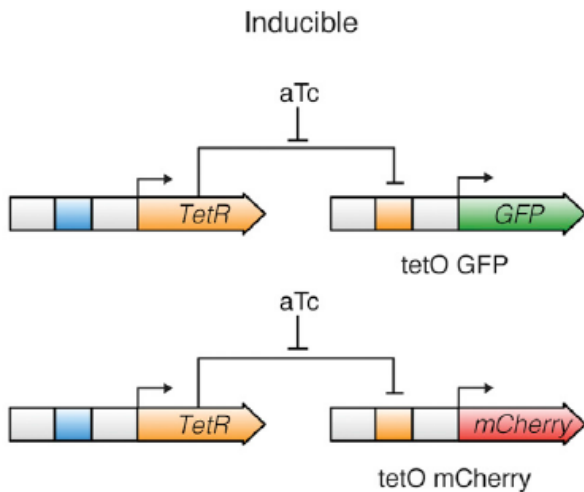
2mm paper discs, 1.8ul reaction

Freeze-dry

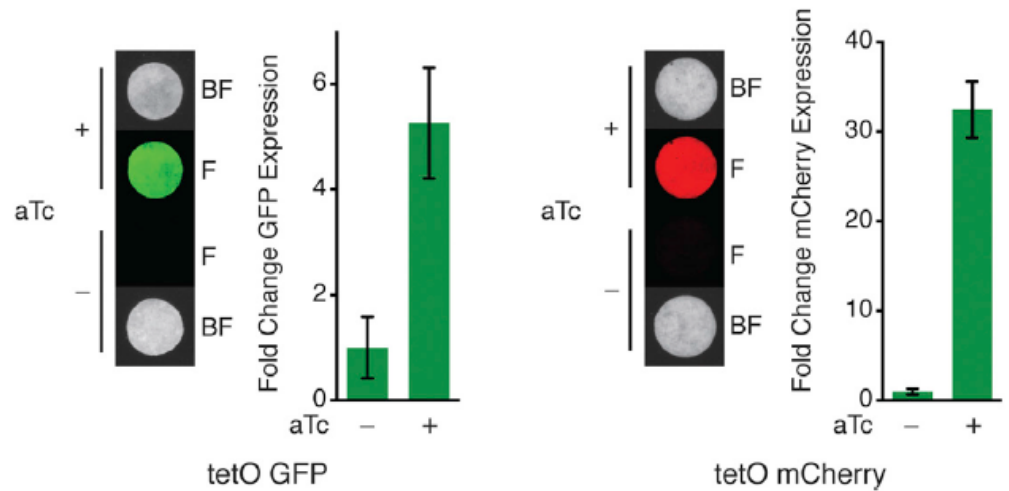
Rehydrate with 1.8ul Tetracycline analog: anhydrotetracycline (aTc)

Incubate at 37 °C, 2 hours

**F**



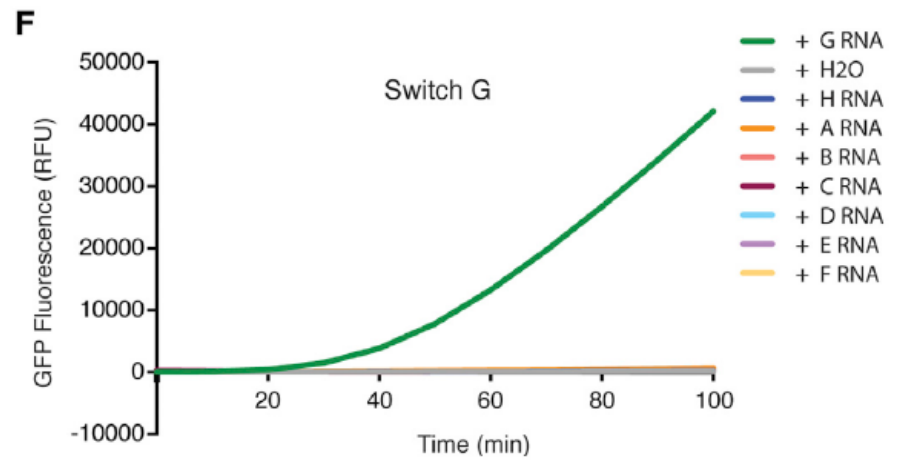
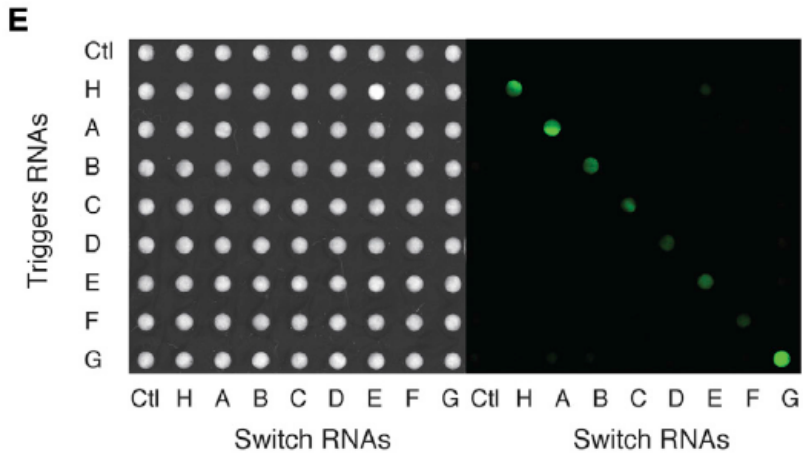
**G**



S30 E.coli cell extract

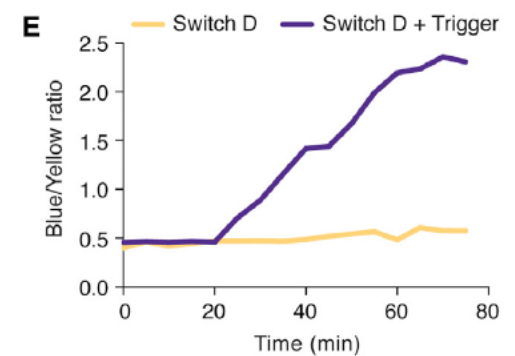
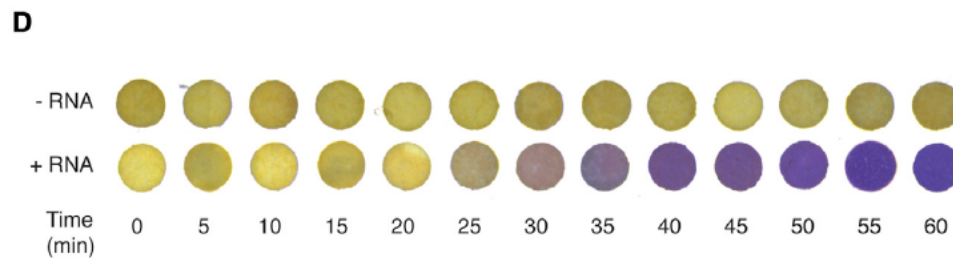
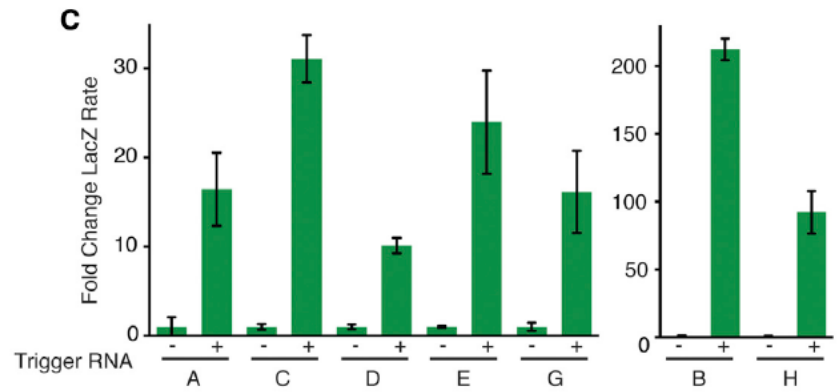
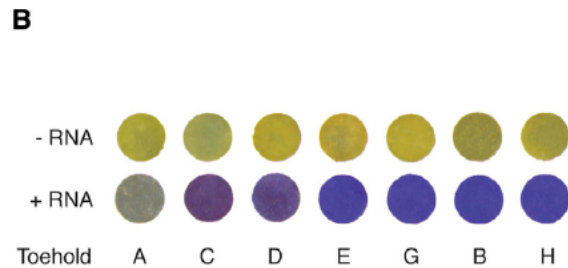
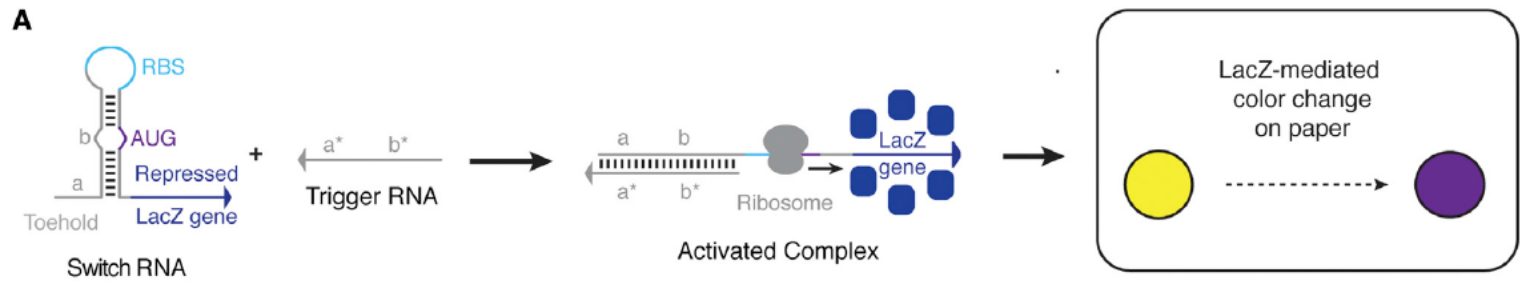


# Orthogonality of toehold switches on cellulose matrix paper



# LacZ expression on cellulose matrix paper

\_visible by naked eyes



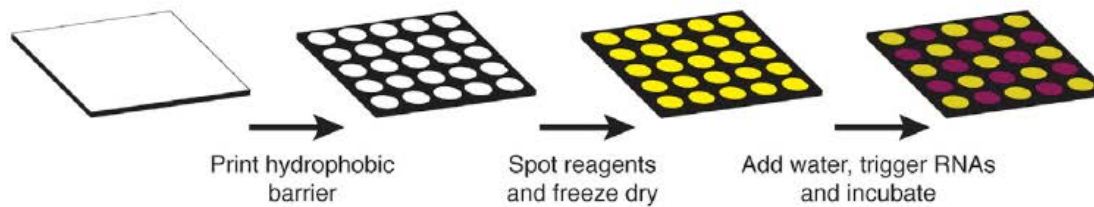


# LacZ expression on cellulose matrix paper

\_low cost and ease to manufacture

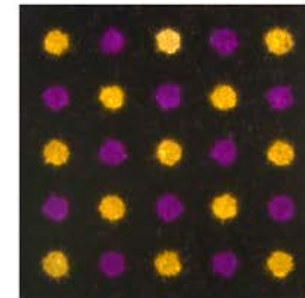
Standard computer printer+ chromatography paper + wax-based ink  
Electronic optic reader

F

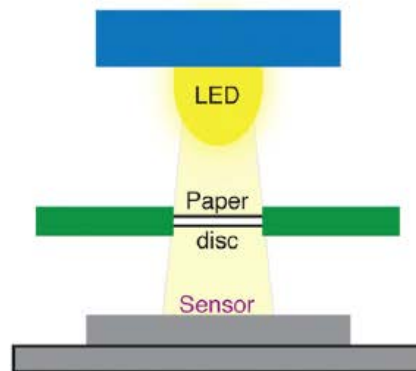
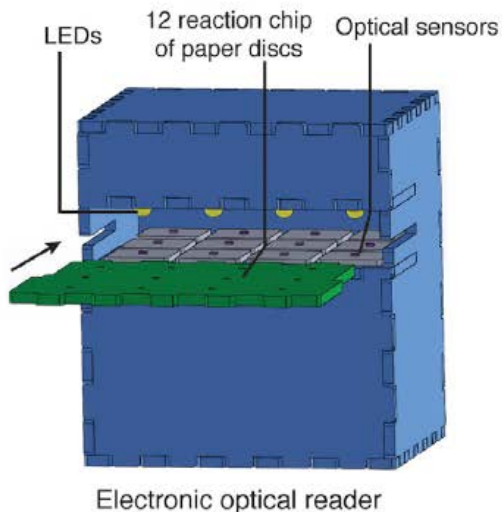


G

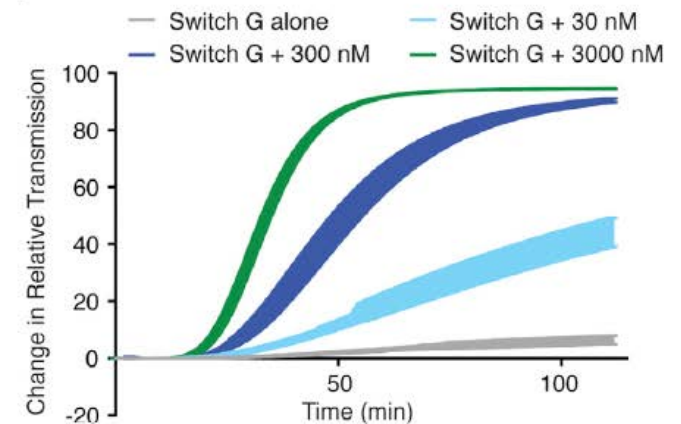
toehold switch E



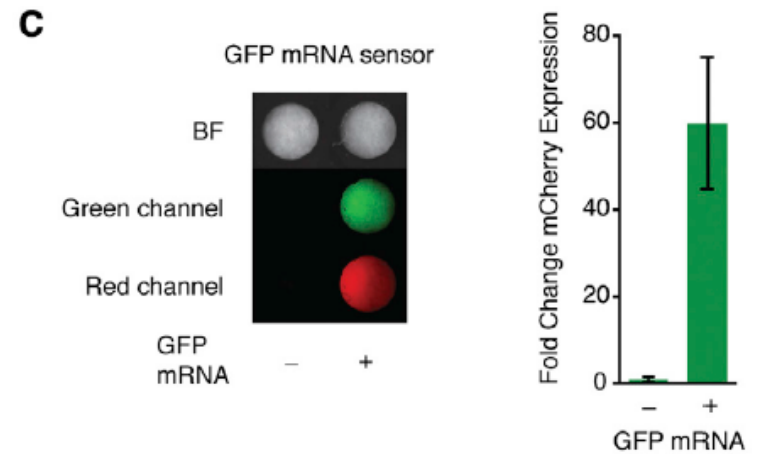
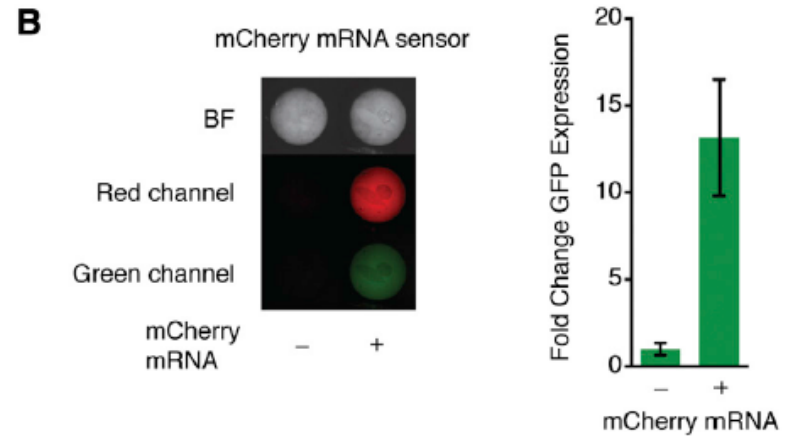
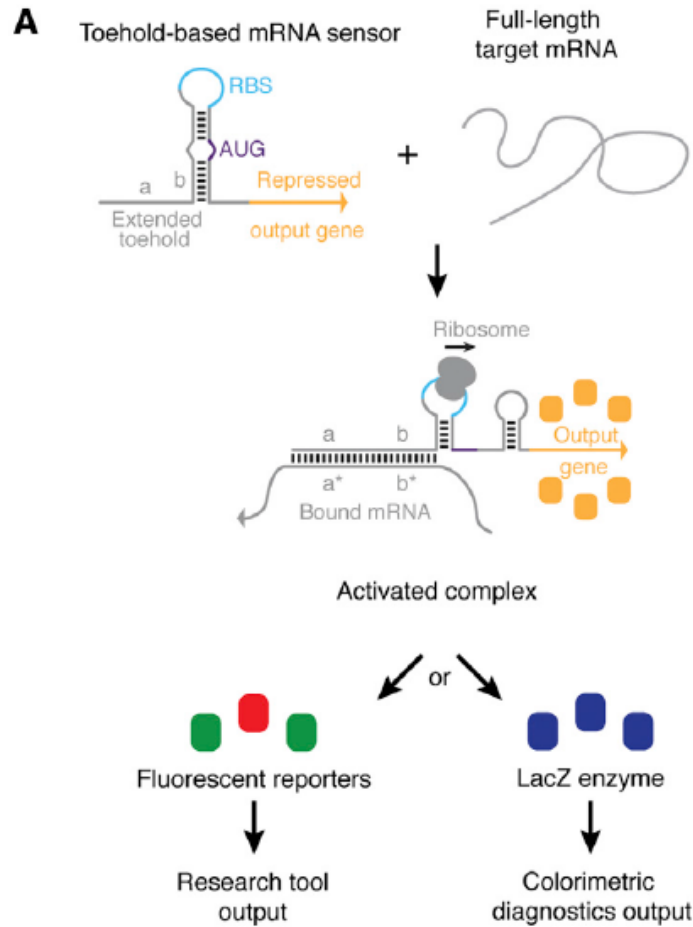
H



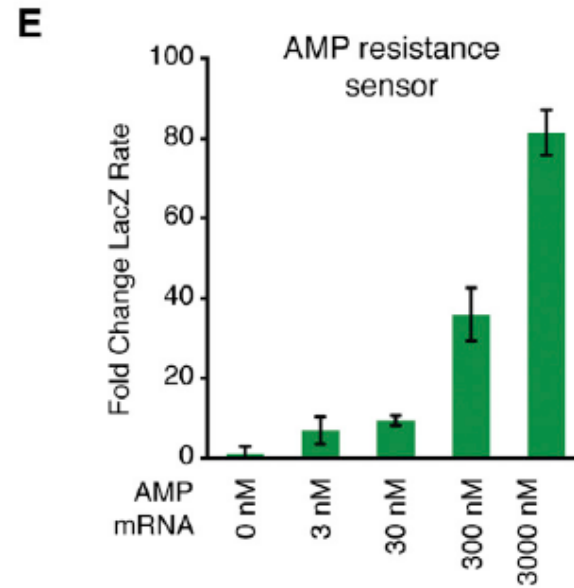
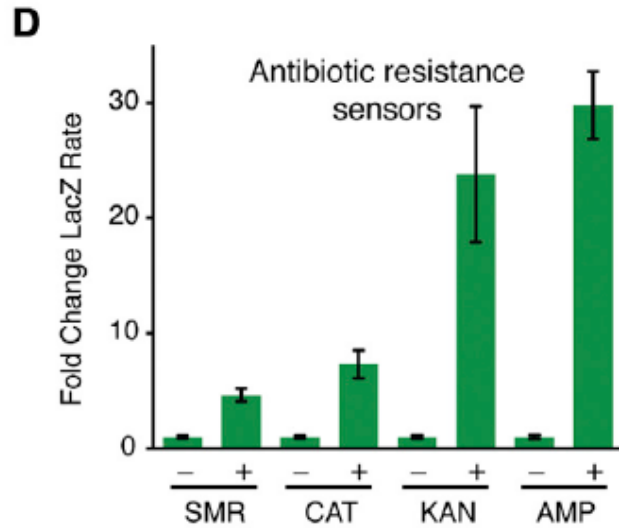
I



# Diagnostics by toehold switches on paper



# Diagnostics by toehold switches on paper \_antibiotic resistance genes

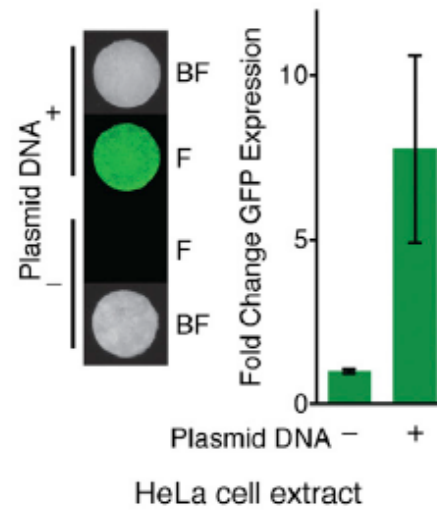


Purified mRNAs

# Diagnostics by toehold switches on paper

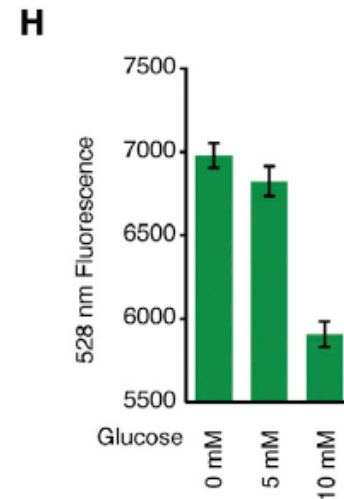
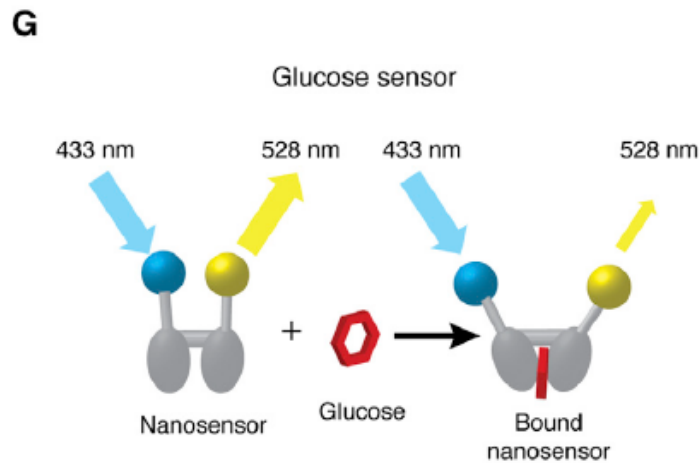
## \_Mammalian cells based system

**F**



# Diagnostics by FRET on paper

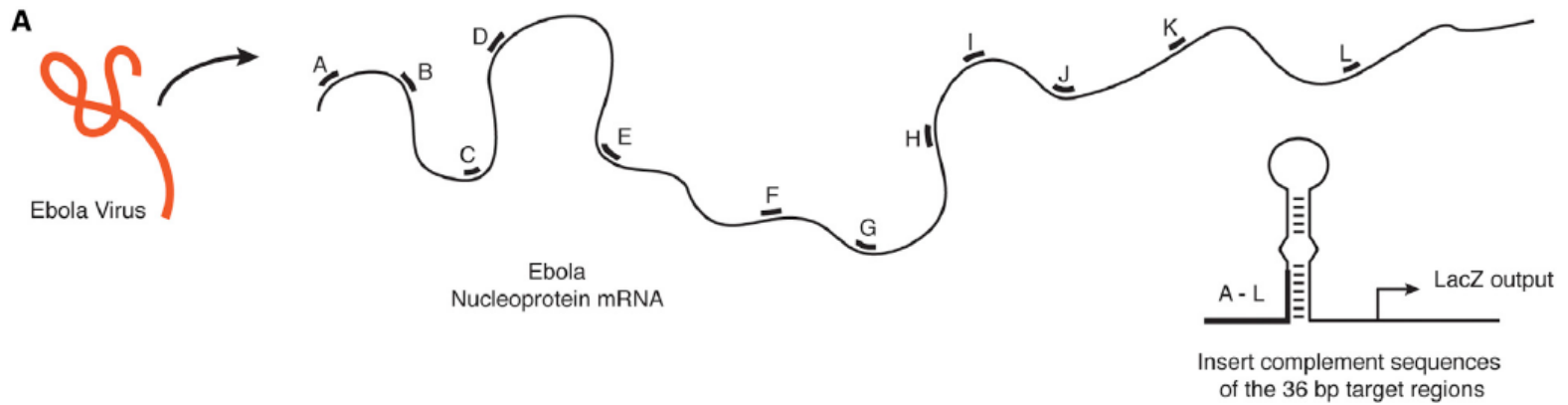
## \_Glucose measurement



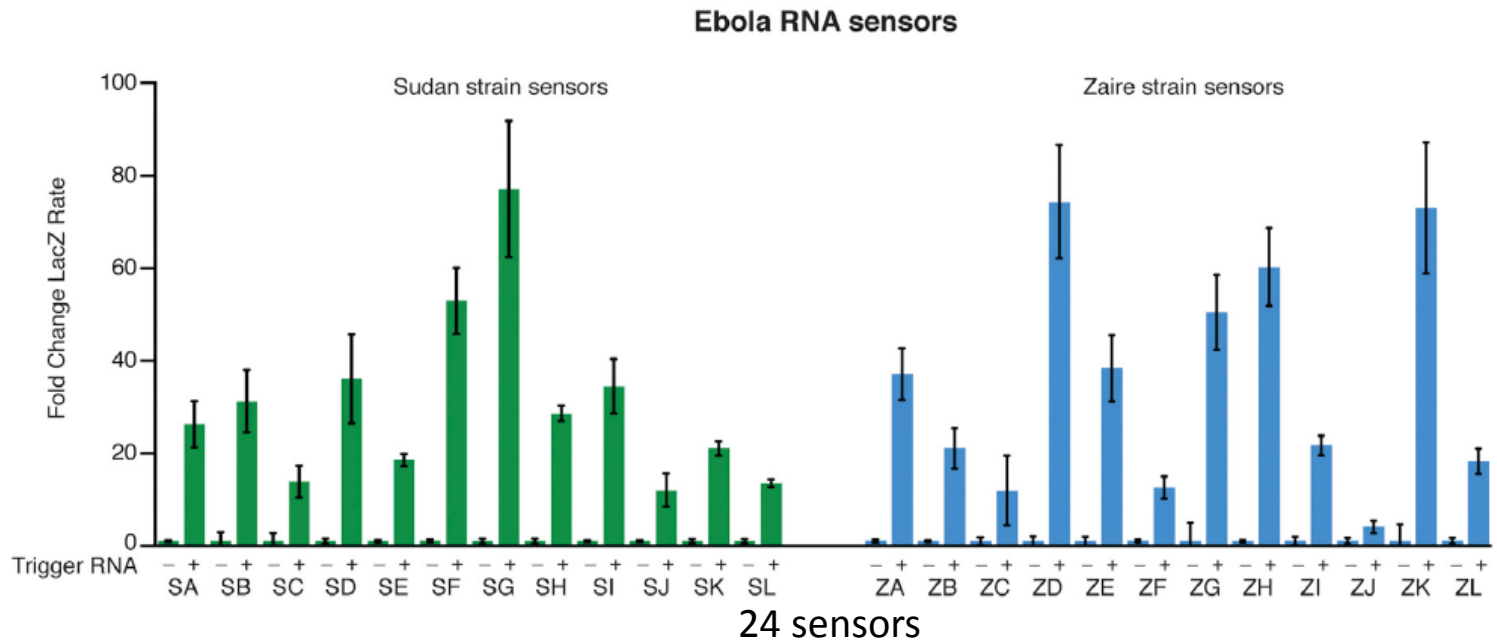
- FLIPglu-30 $\mu$  $\Delta$ 13V: express CFP, Venus, and periplasmic glucose binding
- Reaction in freeze-dried Hela cell extract on paper

# Rapid sensor prototyping

## \_strain typing for Ebola

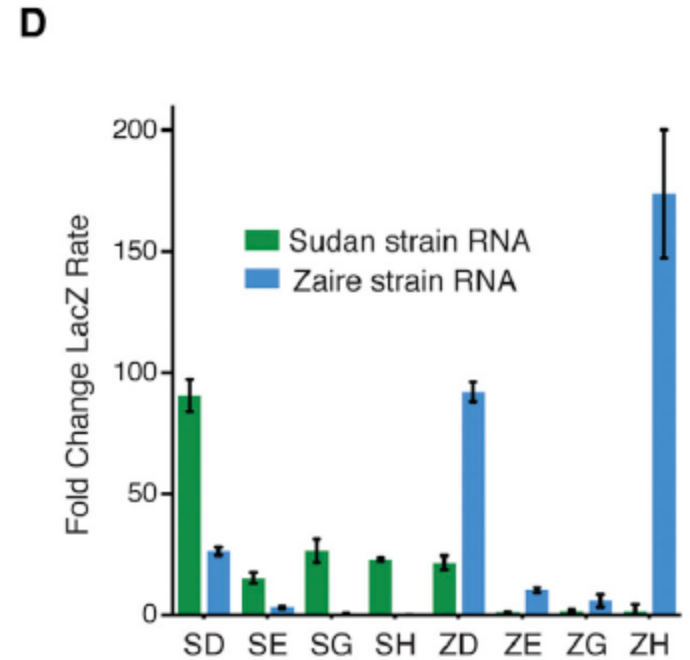
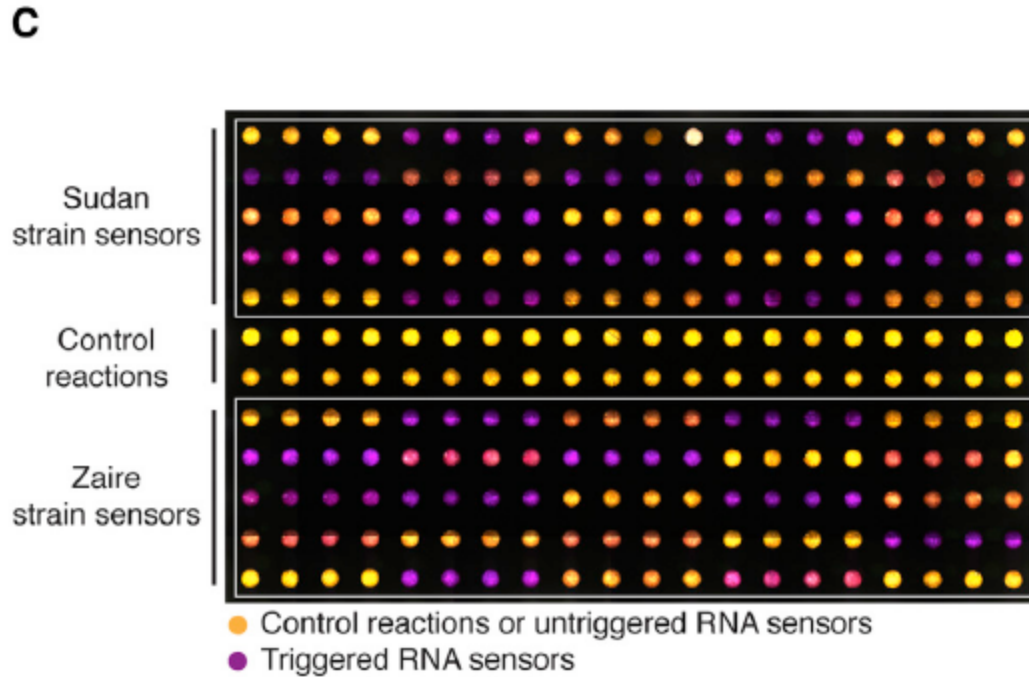


**B**



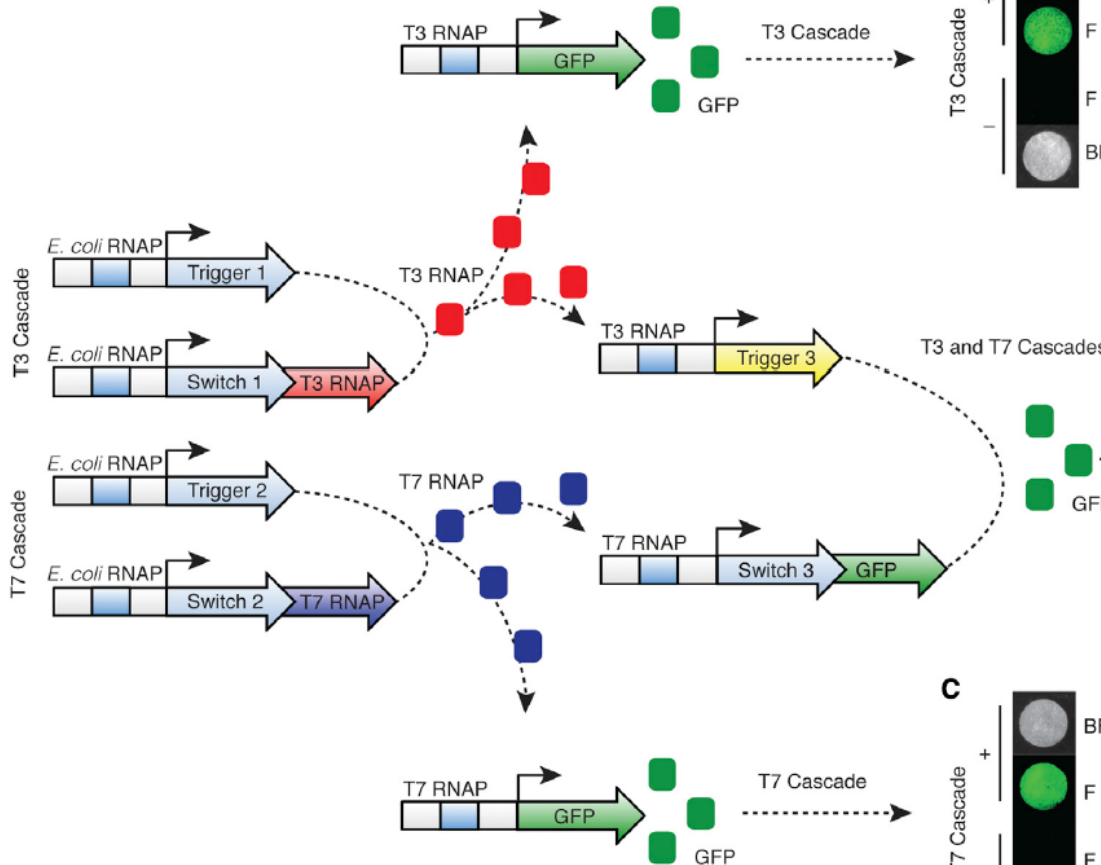
# Rapid sensor prototyping

\_strain typing for Ebola (<12 h)

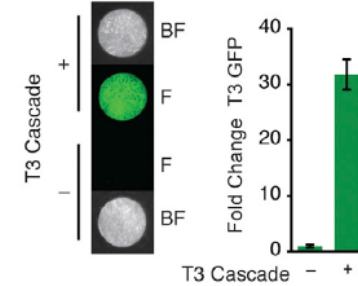


# Complex synthetic gene networks on paper

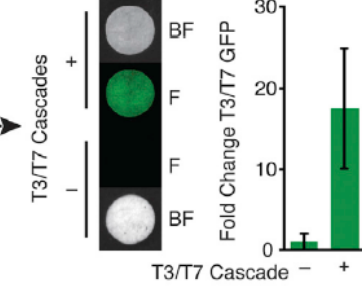
**A**



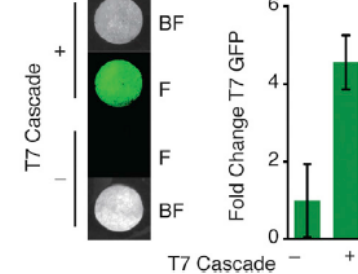
**B**



**D**



**C**





## Summary

Synthetic gene networks on paper or other porous materials

Integrated toehold switches or FRET into paper

Sensitivity: nanomolar, picomolar

Synthetic gene networks on paper has potential to translate existing constructs designed for basic research and biotechnology into portable and readily accessible molecule tools

*Synthetic biology has been predicted to heal us, feed us and fuel us.*

*-David Willetts*

**Thank you!**